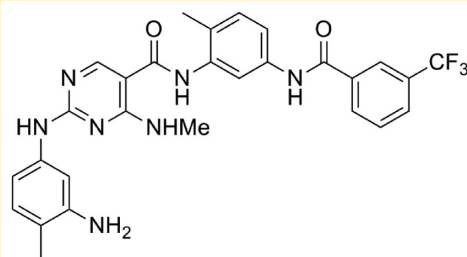


## Discovery of 2-((3-Amino-4-methylphenyl)amino)-N-(2-methyl-5-(3-(trifluoromethyl)benzamido)phenyl)-4-(methylamino)pyrimidine-5-carboxamide (CHMFL-ABL-053) as a Potent, Selective, and Orally Available BCR-ABL/SRC/p38 Kinase Inhibitor for Chronic Myeloid Leukemia

Xiaofei Liang,<sup>†,‡,○</sup> Xiaochuan Liu,<sup>†,§,○</sup> Beilei Wang,<sup>†,‡,○</sup> Fengming Zou,<sup>†,‡,○</sup> Aoli Wang,<sup>†,||</sup> Shuang Qi,<sup>†,‡</sup> Cheng Chen,<sup>†,‡</sup> Zheng Zhao,<sup>†,‡</sup> Wenchao Wang,<sup>†,‡</sup> Ziping Qi,<sup>†,‡</sup> Fengchao Lv,<sup>†,||</sup> Zhenquan Hu,<sup>†,‡</sup> Li Wang,<sup>†,‡</sup> Shanchun Zhang,<sup>‡,⊥</sup> Qingsong Liu,<sup>\*,†,‡,||,#</sup> and Jing Liu<sup>\*,†,‡</sup><sup>†</sup>High Magnetic Field Laboratory, Chinese Academy of Sciences, Mailbox 1110, 350 Shushanhu Road, Hefei, Anhui 230031, P. R. China<sup>‡</sup>CHMFL-HCMTC Target Therapy Joint Laboratory, 350 Shushanhu Road, Hefei, Anhui 230031, P. R. China<sup>§</sup>Department of Chemistry, University of Science and Technology of China, Hefei, Anhui 230036, P. R. China<sup>||</sup>University of Science and Technology of China, P. R. China, Anhui Hefei 230036, P. R. China<sup>⊥</sup>Hefei Cosource Medicine Technology Co. LTD., 358 Ganquan Road, Hefei, Anhui 230031, P. R. China<sup>#</sup>Hefei Science Center, Chinese Academy of Sciences, 350 Shushanhu Road, Hefei, Anhui 230031, P. R. China

## S Supporting Information

**K562: GI<sub>50</sub>: 14 nM**  
**KU812: GI<sub>50</sub>: 25 nM**  
**MEG-01: GI<sub>50</sub>: 16 nM**  
**pABL1: EC<sub>50</sub>: about 100 nM**  
**ABL1: IC<sub>50</sub>: 70 nM**  
**S Score(1)=0.02**  
**T<sub>1/2</sub>: 4.3 h**  
**Bioavailability: 24%**

**ABSTRACT:** Starting from a dihydropyrimidopyrimidine core scaffold based compound 27 (GNF-7), we discovered a highly potent (ABL1: IC<sub>50</sub> of 70 nM) and selective (S score (1) = 0.02) BCR-ABL inhibitor 18a (CHMFL-ABL-053). Compound 18a did not exhibit apparent inhibitory activity against c-KIT kinase, which is the common target of currently clinically used BCR-ABL inhibitors. Through significant suppression of the BCR-ABL autophosphorylation (EC<sub>50</sub> about 100 nM) and downstream mediators such as STAT5, Crkl, and ERK's phosphorylation, 18a inhibited the proliferation of CML cell lines K562 (GI<sub>50</sub> = 14 nM), KU812 (GI<sub>50</sub> = 25 nM), and MEG-01 (GI<sub>50</sub> = 16 nM). A pharmacokinetic study revealed that 18a had over 4 h of half-life and 24% bioavailability in rats. A 50 mg/kg/day dosage treatment could almost completely suppress tumor progression in the K562 cells inoculated xenograft mouse model. As a potential useful drug candidate for CML, 18a is under extensive preclinical safety evaluation now.

## ■ INTRODUCTION

Chronic myeloid leukemia (CML), a hematological cancer of bone marrow white blood cells, constitutes about 15% of adult leukemia, and usually 1–2 patients are diagnosed with CML per 100,000 people/per year in the US.<sup>1</sup> It is characterized by a reciprocal chromosomal translocation between chromosomes 9 and 22 of the break point cluster region (BCR) gene with the Abelson (ABL) gene for ABL1 kinase, which leads to a shortened chromosome 22 (i.e., Philadelphia chromosome).<sup>2,3</sup> The fusion tyrosine kinase BCR-ABL is constitutively active and leads to uncontrolled myeloid cell proliferation through downstream mediators such as Stat5 and ERK.<sup>4</sup> The seminal

discovery of the small molecule inhibitor Imatinib<sup>5,6</sup> has validated BCR-ABL as the drug discovery target for CML. Since then, several ABL kinase inhibitors have been approved for clinical use such as Nilotinib,<sup>7</sup> Dasatinib,<sup>8</sup> Bosutinib,<sup>9</sup> and Ponatinib,<sup>10</sup> and a few are in clinical trials now including Bafetinib,<sup>11</sup> Danusertib,<sup>12</sup> and Rebastinib,<sup>13</sup> etc. In addition, a number of newly discovered inhibitors are in an extensive preclinical study such as GZD824<sup>14</sup> and allosteric inhibitor GNF2, GNFS,<sup>15</sup> etc. Despite the great clinical success, the FDA

Received: October 15, 2015

Published: January 20, 2016

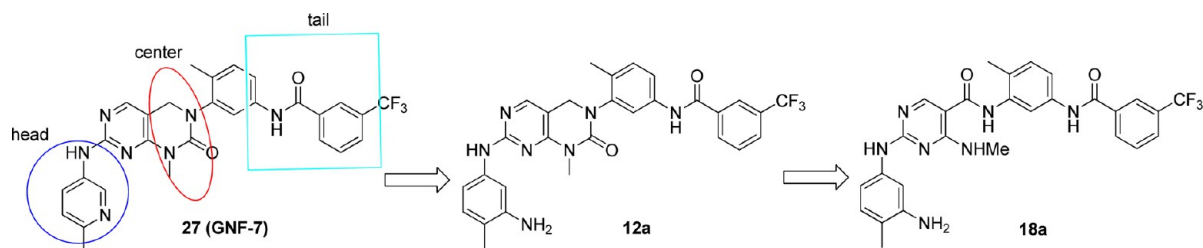
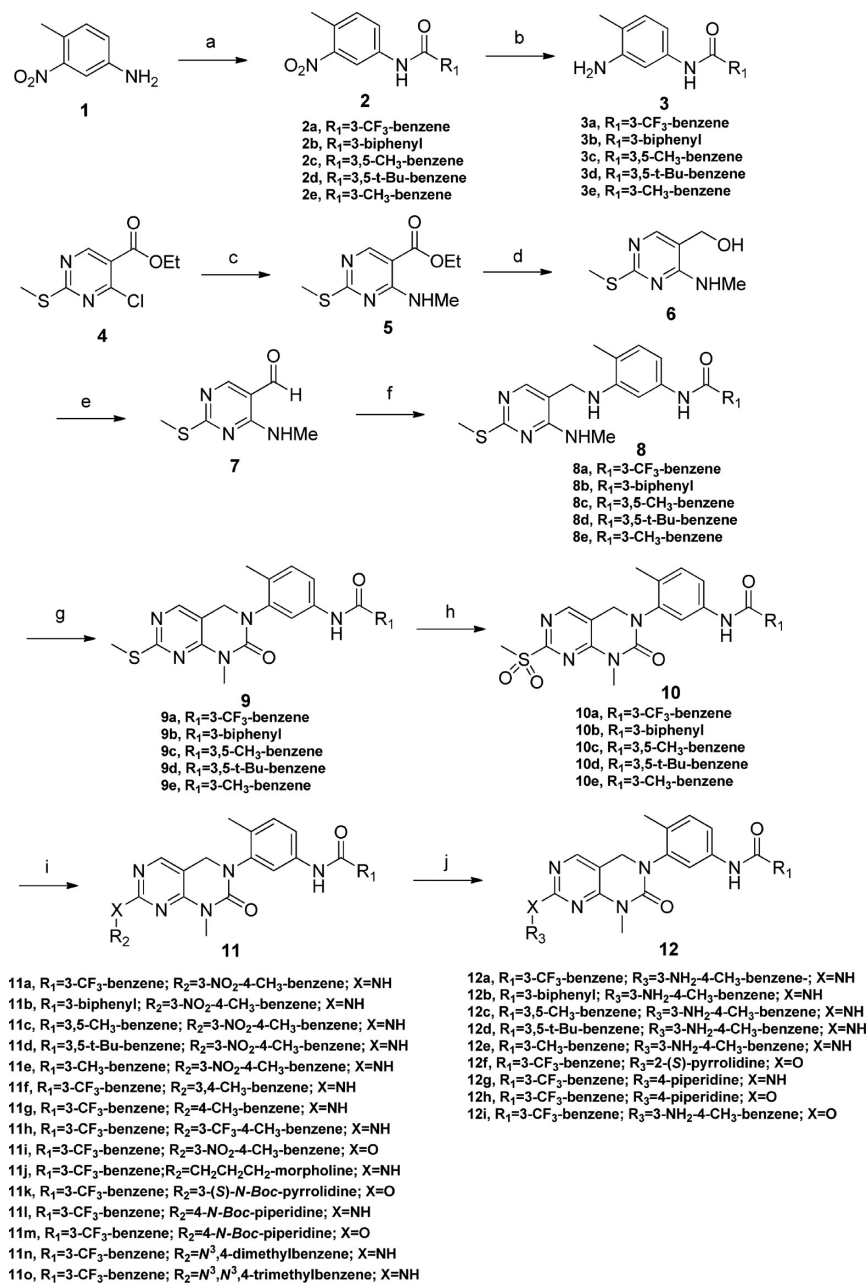


Figure 1. Schematic illustration of the discovery of 18a.

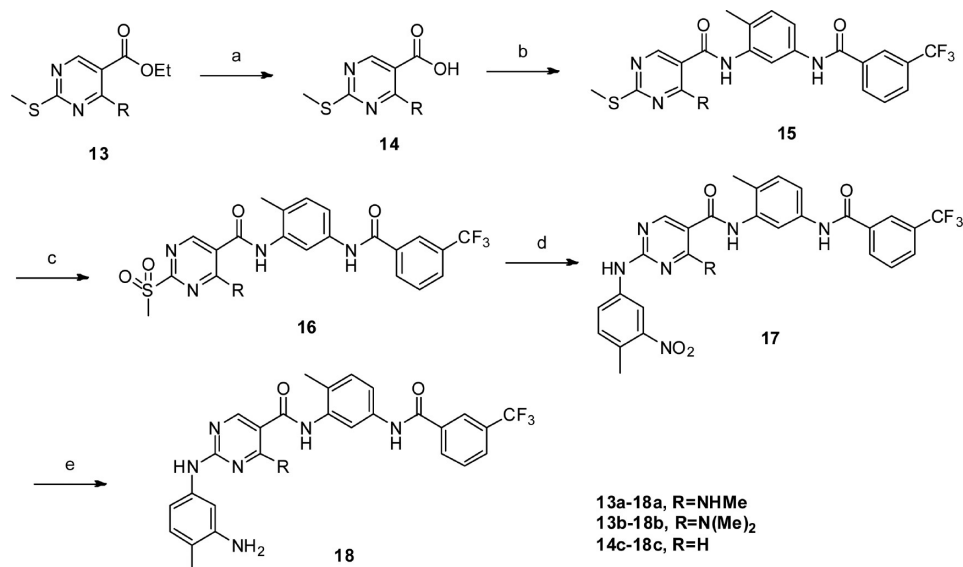
### Scheme 1. Synthetic Route to Dihydropyrimidopyrimidine 12<sup>a</sup>



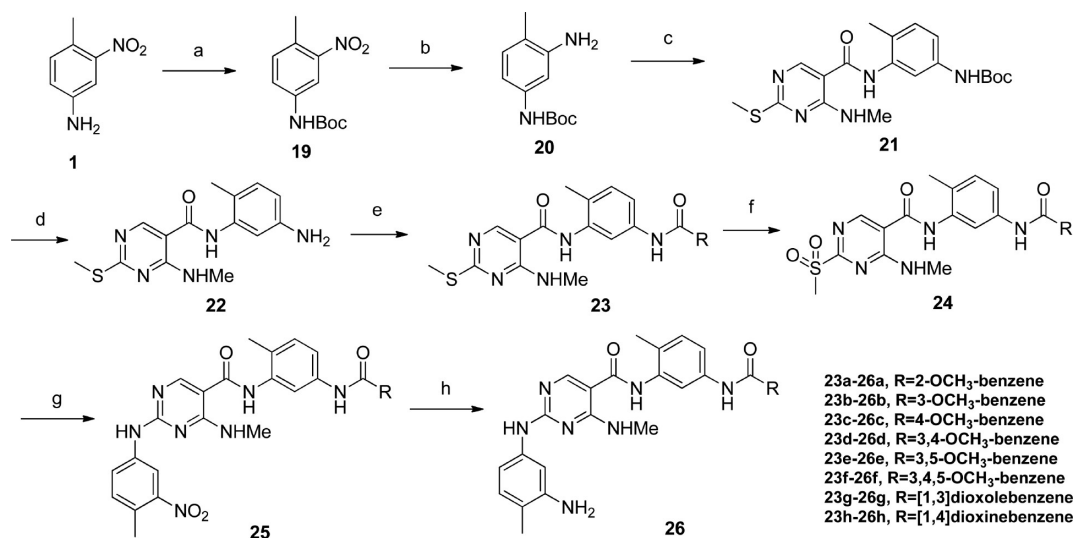
<sup>a</sup>Reagents and conditions: (a) R<sub>1</sub>COCl, THF, DIPEA, 0 °C; (b) H<sub>2</sub>, 10% Pd/C, MeOH, rt; (c) THF, MeNH<sub>2</sub>, TEA, 0 °C; (d) LAH, THF, 0 °C to reflux; (e) DCM, MnO<sub>2</sub>, rt; (f) 3, Na(CN)BH<sub>3</sub>, MeOH, AcOH, rt; (g) triphosgene, DCM, DIPEA, 0 °C to rt; (h) m-CPBA, DCM, 0 °C; (i) R<sub>2</sub>NH<sub>2</sub>, dioxane, TFA, 120 °C or R<sub>2</sub>OH, dioxane, K<sub>2</sub>CO<sub>3</sub>, rt; (j) SnCl<sub>2</sub>·2H<sub>2</sub>O, MeOH, reflux or 4 M HCl, dioxane, rt.

approved drugs such as Imatinib, Nilotinib, Bosutinib, and Dasatinib all potently inhibit other targets such as DDR1/2, c-

KIT, and so on besides ABL1 kinase. Although the role of off-target inhibition is not very clear in the clinical aspect, the

Scheme 2. Synthetic Route to Compound 18<sup>a</sup>

<sup>a</sup>Reagents and conditions: (a) for the synthesis of 14a–b, NaOH, MeOH, H<sub>2</sub>O, rt; (b) 3a, HATU, DIPEA, DMF, rt; (c) m-CPBA, DCM, 0 °C; (d) 4-methyl-3-nitroaniline, dioxane, TFA, 120 °C; (e) SnCl<sub>2</sub>·2H<sub>2</sub>O, MeOH, reflux.

Scheme 3. Synthetic Route to Compound 26<sup>a</sup>

<sup>a</sup>Reagents and conditions: (a) (Boc)<sub>2</sub>O, THF, DMAP, reflux; (b) H<sub>2</sub>, 10% Pd/C, MeOH, rt; (c) 14a, HATU, DMF, DIPEA, rt; (d) 4 M HCl, MeOH, rt; (e) RCOCl, THF, DIPEA, 0 °C; (f) m-CPBA, DCM, 0 °C; (g) 4-methyl-3-nitroaniline, dioxane, TFA, 120 °C; (h) SnCl<sub>2</sub>·2H<sub>2</sub>O, MeOH, reflux.

highly selective BCR-ABL inhibitor is still highly demanded from both the preclinical pathological and clinical side effect mechanistic study points of view. Here, we describe our medicinal chemistry effort from a dihydropyrimidopyrimidine scaffold based multiple target BCR-ABL inhibitor GNF-7<sup>16,17</sup> (compound 27) to a pyrimidine scaffold based highly potent and selective BCR-ABL inhibitor compound 18a (CHMFL-ABL-053), which completely abolished the c-KIT kinase activity (Figure 1).

## RESULTS AND DISCUSSION

**Chemistry.** The synthesis of 12 (Scheme 1) began with amide coupling of 4-methyl-3-nitroaniline with an appropriate carbonyl chloride. The nitrobenzene was then converted to the

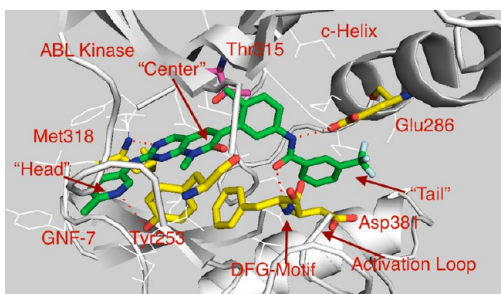
corresponding aniline 3 by reduction with hydrogen under 10% Pd/C conditions. Intermediate pyrimidine aldehyde 7 was obtained from ethyl 4-chloro-2-(methylthio)pyrimidine-5-carboxylate 4. Methylamine pyrimidine 5 was prepared with methylamine in almost quantitative yield. Subsequent reduction of 5 in the presence of LAH in anhydrous THF afforded the desired alcohol 6, which was then oxidized into aldehyde 7. Reductive amination with 7 and 3 afforded diamine derivative 8. The diamine was then converted into the dihydropyrimidopyrimidine 9 with triphosgene, which was subsequently oxidized to the corresponding sulfone 10. Alcohol or aniline derivatives were readily reacted with this sulfone under etherification with base or simple thermal amination conditions to provide compound 11. Finally, SnCl<sub>2</sub> mediated reduction of

the nitrobenzene or N-Boc deprotection furnished compound 12.

Hydrolysis of the ethyl ester of pyrimidine 13 followed by amide-coupling conditions furnished amide 15 (Scheme 2). The sulfide of 15 was then subjected to oxidation, amination, and subsequent reduction to provide aniline compounds 18.

As shown in Scheme 3, compound 26 was prepared starting from 4-methyl-3-nitroaniline 1. After Boc protection (19) and hydrogenation (20), amide 21 was obtained under the standard amide coupling condition with compound 14a. Removal of the Boc protection (22) followed by acylation with appropriate acyl chloride formed amide 23, which was then subjected to oxidation (24), amination (25), and reduction to provide aniline compound 26.

**Structure–Activity Relationship.** The dihydropyrimidopyrimidine compound 27 (GNF-7) has been reported as a type II BCR-ABL inhibitor which could also overcome “gatekeeper” T315I mutation. However, it also potently inhibited other kinases such as JAK1, 2, 3, FGFR3, FLT3, PDGFR, TRKC, etc.<sup>17</sup> After careful analysis of the molecular modeling results, we envisioned that the methylpyridine occupied hinge binding area (the so-called “head” part), the dihydropyrimidopyrimidine moiety occupied “gatekeeper” residue Thr315 adjacent area (the so-called “center” part), and the trifluoromethylbenzene occupied DFG shifting created hydrophobic area (the so-called “tail” part) still have medicinal exploration space for better selectivity (Figure 1). We then chose to systematically optimize the “head”, “center”, and “tail” parts of 27 to obtain a full spectrum of the structure–activity relationship (SAR) as illustrated in Figure 2 with cell based assays using ABL transfused isogenic BaF3 cells and intact CML cancer cell line K562 as the primary readout.



**Figure 2.** Illustration of SAR exploration rationale. ABL kinase was shown in white. GNF-7 was labeled in color by atoms (carbon in green, nitrogen in blue, and oxygen in red). The hydrogen bonds were labeled as red dashed lines. The key amino acid residues for the binding were labeled as follows: carbon in yellow, nitrogen in blue, and oxygen in red. The gatekeeper residue Thr315 was labeled as follows: carbon in magenta, nitrogen in blue, and oxygen in red. The “Head”, “Center”, “Tail”, the DFG-motif, and the “Activation Loop” are pointed out with red arrows.

The “head” moiety occupied the hydrophobic pocket located proximal to the hinge binding region in Abl kinase. Removal of the hydrogen bond acceptor nitrogen atom in 2-methylpyridine of 27 by replacement with a 2-methylbenzene (11g) led to significant loss of activity against parental BaF3 cells ( $GI_{50}$ : 8.0 versus 0.12  $\mu\text{M}$ ) but not in K562 cells ( $GI_{50}$ : 0.018  $\mu\text{M}$  versus 0.009  $\mu\text{M}$ ) (Table 1) which suggested a better selectivity profile and potent on-target (ABL) antiproliferation effect in the TEL and BCR transfused isogenic BaF3 cells whose growth was dependent on the constitutively activated ABL kinase ( $GI_{50}$ :

**Table 1.** Anti-proliferation Efficacies against Intact and Isogenic Cancer Cell Lines of Dihydropyrimidopyrimidine Derivatives<sup>a</sup>

Compd	Structure	BaF3 ( $\mu\text{M}$ )	Tel-Abl-BaF3 ( $\mu\text{M}$ )	P210-BaF3 ( $\mu\text{M}$ )	K562 ( $\mu\text{M}$ )
27		0.12±0.0004	0.005±0.0003	0.014±0.0005	0.009±0.0006
11f		3.76±0.781	0.061±0.001	0.066±0.013	0.017±0.014
11g		8.00±1.136	0.050±0.007	0.019±0.004	0.018±0.005
11h		>10	0.185±0.001	0.067±0.032	0.029±0.006
11j		5.60±0.0002	0.009±0.0004	0.015±0.002	0.007±0.0002
11k		8.38±2.569	0.292±0.0014	0.126±0.003	0.030±0.0009
11l		7.52±0.778	0.105±0.000	0.055±0.044	0.018±0.003
11m		>10	0.288±0.0027	0.132±0.009	0.029±0.010
11n		>10	0.020±0.001	0.014±0.006	0.019±0.001
11o		>10	0.042±0.005	0.030±0.004	0.028±0.008
12a		0.44±0.093	0.005±0.000	0.005±0.000 2	0.003±0.000
12b		>10	0.039±0.007	0.046±0.013	0.023±0.001
12c		>10	0.031±0.0062	0.042±0.006	0.026±0.0021
12d		>10	0.096±0.0062	0.12±0.006	1.3±0.236
12e		>10	0.006±0.0001	0.014±0.005	0.0688±0.001 2
12f		>10	1.380±0.021	1.26±0.03	0.45±0.0022
12g		3-10	0.100±0.1	0.15±0.005	0.019±0.0007
12h		>10	2.043±0.035	2.705±0.327	0.220±0.038
12i		>10	0.015±0.0003	0.032±0.003	0.013±0.001

<sup>a</sup>All  $GI_{50}$  values are presented as the mean  $\pm$  SEM ( $n = 3$ ).

0.050  $\mu\text{M}$  and 0.019  $\mu\text{M}$ , respectively). Increasing the size of this moiety (11f, 11h, 11n, and 11o) retained the potency and selectivity suggesting a large sized pocket residing in this region. However, when the aniline analogue was introduced (12a), it started to potently inhibit parental BaF3 cells again ( $GI_{50}$ : 0.44

Table 2. Anti-proliferation Efficacies against Intact and Isogenic Cancer Cell Lines of Compound 18a and Its Analogues<sup>a</sup>

Compd	Structure	BaF3 (μM)	Tel-ABL-BaF3 (μM)	P210-BaF3 (μM)	K562 (μM)
17a		>10	0.087 ± 0.006	0.036 ± 0.006	0.052 ± 0.011
17b		>10	>10	>10	>10
17c		8.70 ± 3.655	1.400 ± 0.059	1.20 ± 0.096	0.38 ± 0.027
18a		>10	0.053 ± 0.0012	0.007 ± 0.002	0.014 ± 0.001
18b		>10	1.030 ± 0.200	1.849 ± 0.196	1.211 ± 0.031
18c		>10	0.590 ± 0.247	0.47 ± 0.055	0.33 ± 0.025
25a		>10	>10	>10	>10
25b		6.31 ± 0.1	0.983 ± 0.067	1.1 ± 0.154	0.48 ± 0.021
25c		>10	2.900 ± 0.188	3.6 ± 0.399	1.53 ± 0.110
25d		7.30 ± 1.292	1.400 ± 0.176	1.6 ± 0.100	0.68 ± 0.045
25e		>10	2.034 ± 0.112	0.294 ± 0.083	1.07 ± 0.061

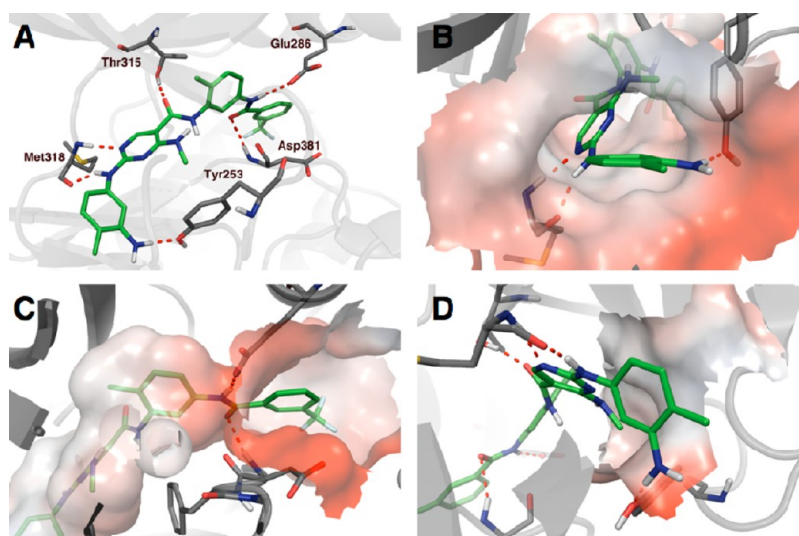
Compd	Structure	BaF3 (μM)	Tel-ABL-BaF3 (μM)	P210-BaF3 (μM)	K562 (μM)
25f		>10	0.931 ± 0.08	1.5 ± 0.171	0.43 ± 0.063
25g		>10	1.290 ± 0.040	0.214 ± 0.095	0.499 ± 0.030
25h		>10	4.240 ± 0.0763	4.1 ± 0.617	0.32 ± 0.017
26a		>10	5.20 ± 0.202	6.4 ± 0.212	1.98 ± 0.351
26b		>10	0.524 ± 0.003	0.57 ± 0.097	0.33 ± 0.155
26c		>10	1.300 ± 0.017	2.1 ± 0.181	0.61 ± 0.079
26d		>10	1.100 ± 0.259	1.6 ± 0.174	0.38 ± 0.078
26e		>10	0.628 ± 0.004	0.311 ± 0.084	0.230 ± 0.151
26f		>10	0.445 ± 0.111	0.67 ± 0.117	0.25 ± 0.047
26g		>10	0.570 ± 0.010	0.124 ± 0.020	0.353 ± 0.006
26h		>10	0.787 ± 0.129	1.1 ± 0.153	0.32 ± 0.086

<sup>a</sup>All GI<sub>50</sub> values are presented as the mean ± SEM (*n* = 3).

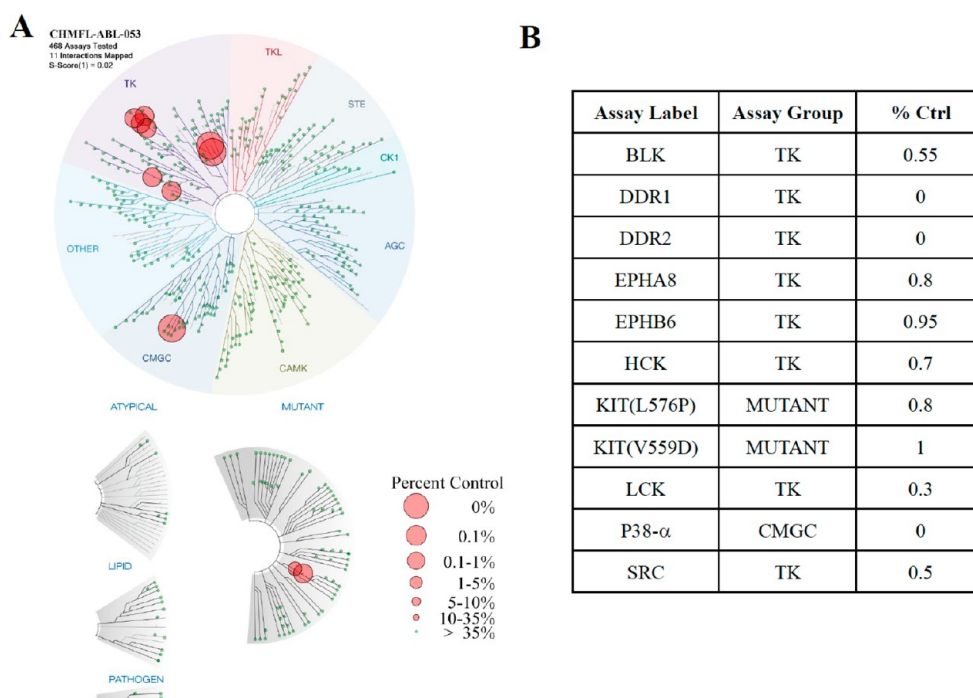
μM). Interestingly, replacement of the aromatic substituents with cyclic aliphatic rings such as piperidine (**12g**), *N*-Boc piperidine (**11i**), or the aliphatic chain (**11j**) did not affect the potency. This suggested that the hydrophobic pocket in this area was flexible. Since the modeling study revealed that the 2-aminopyrimidine part of **27** formed two hydrogen bonds in the hinge-binding region with Met318, we then tried to replace the -NH with an oxygen atom to see if the activity remained. Interestingly, compound **12i** still retained the activities against the K562 cells, Tel-ABL-BaF3 cells, and P210-BaF3 cells (GI<sub>50</sub>: 0.013 μM, 0.015 μM, and 0.032 μM, respectively) and still exhibited good selectivity over parental BaF3 cells (GI<sub>50</sub>: > 10 μM). Unfortunately, when the "head" moiety was replaced by aliphatic rings with the O-bridged hinge binding such as (*S*)-*N*-Boc 3-hydroxypyrrolidine (**11k**), *N*-Boc 4-hydroxypiperidine (**11m**), (*S*)-pyrrolidin-3-ol (**12f**), and piperidin-4-ol (**12h**), significant activity loss were observed in K562 cells, Tel-ABL-BaF3 cells, and P210-BaF3 cells. This suggested that the hinge binding factors and "head" hydrophobic binding factors are dependent on each other to contribute to the binding. We next explored the "tail" moiety, which occupied the hydrophobic pocket formed by the "DFG" motif shift in the inactive conformation. Keeping the "head" and "center" moiety as in **12a** while switching the trifluoromethyl group in the "tail" to a methyl group (**12e**) resulted in about 23-fold potency loss against K562 cells (GI<sub>50</sub>: 0.069 μM versus 0.003 μM), though the activities against TEL-ABL-BAF3 cells (GI<sub>50</sub>: 0.006 μM)

and P210-BaF3 cells (GI<sub>50</sub>: 0.014 μM) and selectivity against parental BaF3 cells (GI<sub>50</sub>: > 10 μM) were retained. In addition, replacement of the trifluoromethyl group with a bulky aromatic group (**12b** and **12c**) did not result in significant activity loss in K562 cells (GI<sub>50</sub>: 0.023 μM and 0.026 μM), TEL-ABL-BAF3 cells (GI<sub>50</sub>: 0.039 μM and 0.031 μM), and P210-BaF3 cells (GI<sub>50</sub>: 0.046 μM and 0.042 μM). However, modification of the 3,5-position of the tail group with two *tert*-butyl groups (**12d**) significantly lowered the activity against K562 cells (GI<sub>50</sub>: 1.3 μM versus 0.003 μM), which suggested that the hydrophobic pocket created by "DFG" motif shifting in Abl kinase could only tolerate medium size of hydrophobic moiety.

Despite the similar potencies of **12a** against model cell lines such as **27**, the narrow selectivity window between the parental BaF3 and isogenic BaF3 cell lines led us to explore more of the "center" moiety by opening the cyclic urea ring of **12a**, which presumably would increase the flexibility and improve the solubility. Removal of the carbonyl group offered **18a**, which displayed potent antiproliferation efficacy against K562 cells (GI<sub>50</sub>: 0.014 μM) and P210-BaF3 cells (GI<sub>50</sub>: 0.007 μM) and exhibited good selectivity over parental BaF3 cells (GI<sub>50</sub>: > 10 μM) (Table 2). Either the introduction of one more methyl group at this position (**18b**) or the removal of the 4-methylamino group (**18c**) resulted in significant activity loss against K562 cells (GI<sub>50</sub>: 1.211 μM and 0.33 μM). Compared to **18a–c**, the nitro group bearing compounds **17a–c** all showed a loss of potency slightly or significantly. We then



**Figure 3.** Compound **18a** was docked into Abl kinase (PDB ID: 2HYY). Hydrogen bonds are indicated by red dashed lines to key amino acid residues. (A) Cartoon view of the binding mode of **18a** with ABL1 kinase. (B) Solid surface view of the shallow hydrophobic pocket located adjacent to the hinge binding area. (C) Solid surface view of the hydrophobic pocket formed by the D(out)FG inactive conformation. (D) Solid surface view of the small hydrophobic pocket formed by Leu248, Val256, and Try253 to accommodate the methylamine moiety.



**Figure 4.** Kinome wide selectivity profiling of **18a** with DiscoverX KinomeScan technology. Measurements were performed at a concentration of 1  $\mu$ M of the inhibitor. The affinity was defined with respect to a DMSO control. (A) Treemap demonstration of **18a**'s selectivity in 468 kinase targets. (B) Other targets that demonstrated strong binding to **18a** with a percent control number less than 1.

turned our attention to the “tail” part of the **18a** scaffold. Replacement of the 3-trifluoromethyl group with 3-methoxy group in the tail phenyl ring (**25b** and **26b**) dramatically decreased their potency against K562 cells ( $GI_{50}$ : 0.48  $\mu$ M and 0.33  $\mu$ M, respectively). Shifting the 3-methoxy group to the 2-position (**25a**, **26a**), 4-position (**25c**, **26c**), or one bearing the dimethoxy group (**25d**, **25e**, **26d**, **26e**), trimethoxy group (**25f**, **26f**), [1,3]dioxo (**25g**, **26g**), and [1,4]dioxine (**25h**, **26h**) substituents in the tail phenyl ring all led to the significant activity loss.

Since compound **18a** exhibited the best activity and selectivity profile, we then moved forward to study the binding mode of **18a** with ABL kinase via molecular modeling. The model illustrated that it did prefer to adopt a type II binding mode as designed. The aminopyrimidine formed two hydrogen bonds with Met318 in the hinge area (Figure 3A). The Glu286 residue in the c-Helix and “DFG” residue Asp381 formed two typical hydrogen bonds with amide linkage between the “center” moiety and the “tail” moiety. The gatekeeper residue Thr315 formed a hydrogen bond with the carbonyl group linking the aminopyrimidine moiety and the “tail” part.

Furthermore, Tyr253 also formed a hydrogen bond with the amino group in the “head” moiety, which provided the explanation for the activity difference between **17a–c** and **18a–c**. The hydrophobic pocket in the hinge binding area is shallow and flat, which explained why both aromatic and aliphatic rings and chains could be tolerated (Figure 3B). 3-Trifluoromethylbenzene in the “tail” occupied the hydrophobic pocket formed by “DFG” motif through van der Waals interactions (Figure 3C). In addition, Leu248, Val256, and Tyr253 formed a small hydrophobic pocket, which could accommodate the aminomethyl group in the aminopyrimidine “center” moiety, and this could explain the reason why compounds **18b** and **18c** lost activity (Figure 3D).

We next examined the kinome wide selectivity profile of **18a** (1  $\mu\text{M}$ ) on the DiscoverRx's KinomeScan profiling platform. The results demonstrated that **18a** was highly selective among 468 kinases tested and exhibited an S score (1) = 0.02 (Figure 4 and Supplemental Table 1). The data also indicated that **18a** might have strong binding (with a percent control number less than 1) against BLK, DDR1, DDR2, EPHA8, EphB6, HCK, LCK, p38 $\alpha$ , and SRC kinases. Further confirmation with an Invitrogen SelectScreen biochemical assay revealed that **18a** exhibited an  $\text{IC}_{50}$  of 70 nM against ABL1 kinase and also strongly inhibited p38 $\alpha$  ( $\text{IC}_{50}$ : 62 nM) and SRC kinases ( $\text{IC}_{50}$ : 90 nM) (Table 3). However, it was less potent with DDR1

**Table 3. Invitrogen SelectScreen Biochemical Characterization of 18a (Values = Mean  $\pm$  SEM,  $n = 2$ )**

kinase	<b>18a</b> ( $\text{IC}_{50}$ : nM)
ABL1	70 $\pm$ 5
DDR1	292 $\pm$ 53
DDR2	457 $\pm$ 23
c-KIT	>10000
P38	62 $\pm$ 6
SRC	90 $\pm$ 3

( $\text{IC}_{50}$ : 292 nM) and DDR2 ( $\text{IC}_{50}$ : 457 nM). Previous reports also showed that Dasatinib and Nilotinib had strong binding to p38 $\alpha$  ( $K_i$ : 27 nM and 460 nM respectively) (Supplemental Table 2). Bosutinib and Dasatinib showed strong binding to

SRC ( $K_i$ : 1 nM and 0.21 nM respectively). In addition, **18a** did not exhibit apparent potency against c-KIT kinase ( $\text{IC}_{50}$ : over 10000 nM), which is the common off-target for clinically used BCR-ABL inhibitors Imatinib, Nilotinib, Botutinib, and Dasatinib (Table 3 and Supplemental Table 2). In the TEL-fused isogenic BaF3 cells, **18a** displayed great selectivity between the BCR-ABL and other potential off-targets including SRC, DDR1, DDR2, LCK, BLK, and HCK (Table 4). We also tested **18a** against a variety of clinically important mutations of ABL in the p210 fused BaF3 assay system. Interestingly, it was sensitive against ABL F317L, F317I, and M351T mutants but was relatively resistant against E255K, Q252H, Y253F, and H369P mutants and completely lost the activity to the gatekeeper mutant T315I (Table 4).

It is intriguing that compound **18a** uniquely abolished c-KIT kinase activity in comparison to that of other ABL inhibitors. In order to understand this from a structure point of view, we then docked compound **18a** into c-KIT kinase (PDB ID: 1T46) (Figure 5). The modeling results suggested that **18a** might adopt a similar type II binding mode as it binds to ABL kinase (Figure 5A). However, when c-KIT and ABL kinase were superimposed, we found that in ABL kinase the Tyr253 located in the p-loop which could provide a key hydrogen bond with the methyl aniline moiety in the “head” part of **18a** was replaced by Gly596 residue in c-KIT kinase (Figure 5B). Lack of this key hydrogen bond may result in the loss of inhibitory potency of **18a** against c-KIT kinase.

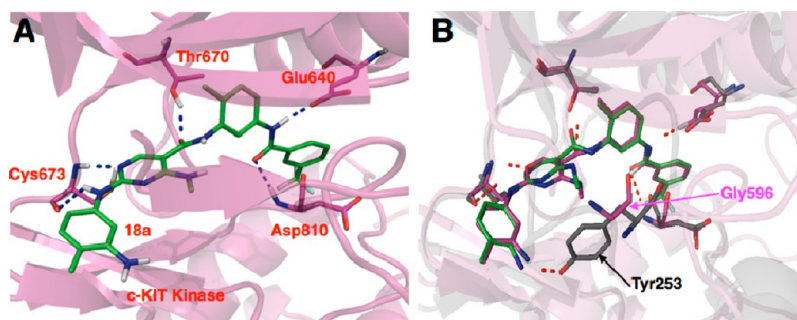
We then compared **18a** with Imatinib and Nilotinib in a panel of intact leukemia cancer cell lines including the CML and AML (Table 5). Compound **18a** exhibited potent antiproliferation efficacies against all of the three BCR-ABL driven CML cell lines K562 ( $\text{GI}_{50}$ : 14 nM), KU812 ( $\text{GI}_{50}$ : 25 nM), and MEG-01 ( $\text{GI}_{50}$ : 16 nM) but not other AML cell lines, implying strong and selective on-target effects. In addition, **18a** did not display any apparent activity against the CHL cell line, indicating a good nonspecific toxicity profile.

We then investigated **18a**'s effects on the BCR-ABL mediated signals in BCR-ABL driven CML cell lines K562, KU812, and MEG-01 (Figure 6). Compound **18a** almost completely suppressed BCR-ABL kinase autophosphorylation at the Y245 site in K562, KU812, and MEG-01 at the

**Table 4. Confirmation of Target Inhibition Revealed in the KinomeScan with Isogenic BaF3 Cell Lines<sup>a</sup>**

cell line	<b>18a</b> ( $\mu\text{M}$ )	Imatinib ( $\mu\text{M}$ )	Nilotinib ( $\mu\text{M}$ )	Dasatinib ( $\mu\text{M}$ )
parental BaF3	>10	6.7 $\pm$ 0.2	2.1 $\pm$ 0.05	>10
BaF3/p210	0.007 $\pm$ 0.004	0.38 $\pm$ 0.03	0.004 $\pm$ 0.0005	0.003 $\pm$ 0.0001
TEL-SRC	0.2 $\pm$ 0.005	2.1 $\pm$ 0.05	0.47 $\pm$ 0.02	<0.0003
TEL-DDR1-BaF3	3.3 $\pm$ 0.08	3–10	1.1 $\pm$ 0.02	10–3
TEL-DDR2-BaF3	>10	7.7 $\pm$ 0.5	1.4 $\pm$ 0.05	10–3
TEL-LCK-BaF3	0.6 $\pm$ 0.01	0.5 $\pm$ 0.06	0.87 $\pm$ 0.21	0.001 $\pm$ 0.00009
TEL-BLK-BaF3	1.1 $\pm$ 0.17	4.1 $\pm$ 1.6	1.3 $\pm$ 0.03	0.005 $\pm$ 0.00002
TEL-HCK-BaF3	0.98 $\pm$ 0.004	9.7 $\pm$ 2	4.1 $\pm$ 0.2	0.039 $\pm$ 0.0001
p210-E255 K-BaF3	0.313 $\pm$ 0.032	1.93 $\pm$ 0.253	0.021 $\pm$ 0.006	0.017 $\pm$ 0.0004
p210-F317L-BaF3	0.045 $\pm$ 0.002	2.169 $\pm$ 0.039	0.202 $\pm$ 0.01	0.014 $\pm$ 0.0009
p210-F317I-BaF3	0.073 $\pm$ 0.008	0.855 $\pm$ 0.081	0.0546 $\pm$ 0.004	0.01 $\pm$ 0.001
p210-M351T-BaF3	0.045 $\pm$ 0.002	0.625 $\pm$ 0.253	0.017 $\pm$ 0.001	0.003 $\pm$ 0.0005
p210-Q252H-BaF3	0.14 $\pm$ 0.033	0.659 $\pm$ 0.072	0.023 $\pm$ 0.001	0.008 $\pm$ 0.001
p210-Y253F-BaF3	0.363 $\pm$ 0.001	>10	1.093 $\pm$ 0.029	0.001 $\pm$ 0.0006
p210-H369P-BaF3	0.440 $\pm$ 0.029	1.69 $\pm$ 0.177	0.025 $\pm$ 0.0044	0.004 $\pm$ 0.0002
p210-T315I-BaF3	9.25 $\pm$ 0.01	>10	>10	9.94 $\pm$ 4.0

<sup>a</sup>All  $\text{GI}_{50}$  values are presented as the mean  $\pm$  SEM ( $n = 3$ ).



**Figure 5.** Comparison of the binding modes of compound **18a** between ABL and c-KIT kinase. (A) Compound **18a** was docked into c-KIT kinase (PDB ID: 1T46). The c-KIT protein is shown in magenta. Compound **18a** was displayed as follows: carbon in green, nitrogen in blue, and oxygen in red. The key binding amino acid residues from the protein are displayed as follows: carbon in magenta, nitrogen in blue, and oxygen in red. (B) Superimposition of c-KIT (in magenta) and ABL (in gray, PDB ID: 2HYY) kinase in complex with compound **18a**.

**Table 5. Anti-proliferation Effects of 18a against Varieties of Intact Cancer Cell Lines<sup>a</sup>**

cell line	cell type	18a ( $\mu\text{M}$ )	Imatinib ( $\mu\text{M}$ )	Nilotinib ( $\mu\text{M}$ )	Dasatinib ( $\mu\text{M}$ )
K562	CML	0.014 $\pm$ 0.006	0.14 $\pm$ 0.001	0.002 $\pm$ 0.0002	<0.0003
KU812	CML	0.025 $\pm$ 0.002	0.16 $\pm$ 0.005	0.001 $\pm$ 0.0002	<0.0003
MEG-01	CML	0.016 $\pm$ 0.0056	0.24 $\pm$ 0.0267	0.016 $\pm$ 0.0014	<0.0003
MV4-11	AML	8 $\pm$ 0.4	>10	>10	3.6 $\pm$ 0.1
MOLM14	AML	>10	>10	>10	2.3 $\pm$ 0.09
U937	AML	>10	>10	>10	>10
HEL	AML	>10	5.3 $\pm$ 0.2	3.9 $\pm$ 0.05	5.3 $\pm$ 0.4
CHL	hamster lung cell	>10	>10	4.2 $\pm$ 0.6	0.27 $\pm$ 0.1

<sup>a</sup>All GI<sub>50</sub> values are presented as the mean  $\pm$  SEM ( $n = 3$ ).

concentration of 300 nM. BCR-ABL kinase downstream mediator Stat5, CrkL, and ERK phosphorylation was also significantly inhibited in a concentration-dependent manner. Interestingly, unlike Imatinib which had no effect on CrkL's phosphorylation, both **18a** and Dasatinib could affect CrkL's phosphorylation, which indicated a different pharmacology profile among them. The results also showed that phosphorylation of SRC kinase was much less potently inhibited than phosphorylation of BCR-ABL by **18a**, though they showed similar sensitivity to **18a** in the biochemical assay. This indicated that the BCR-ABL inhibitory activity of **18a** significantly contributed to the antiproliferative effect in these CML cell lines. P38 $\alpha$  phosphorylation was potently inhibited, which is in accordance with its biochemical inhibitory activities. In addition, even in the early 12 h, **18a** could dose-dependently arrest the cell cycle progression in the G0/G1 phase in these cells (Figure 7A). Upon 24 or 48 h of drug treatment, 100 nM concentration of **18a** could significantly induce apoptotic cell death (Figure 7B).

In the study of PK profiling in rats, compound **18a** exhibited good systemic exposure (AUC = 1715.51 ng/mL·h,  $C_{\text{max}}$  = 367.61 ng/mL), favorable oral bioavailability ( $F = 24.19\%$ ), and acceptable half-life ( $t_{1/2} = 4.33$  h) following oral administration of a single dose of 10 mg/kg (Table 6).

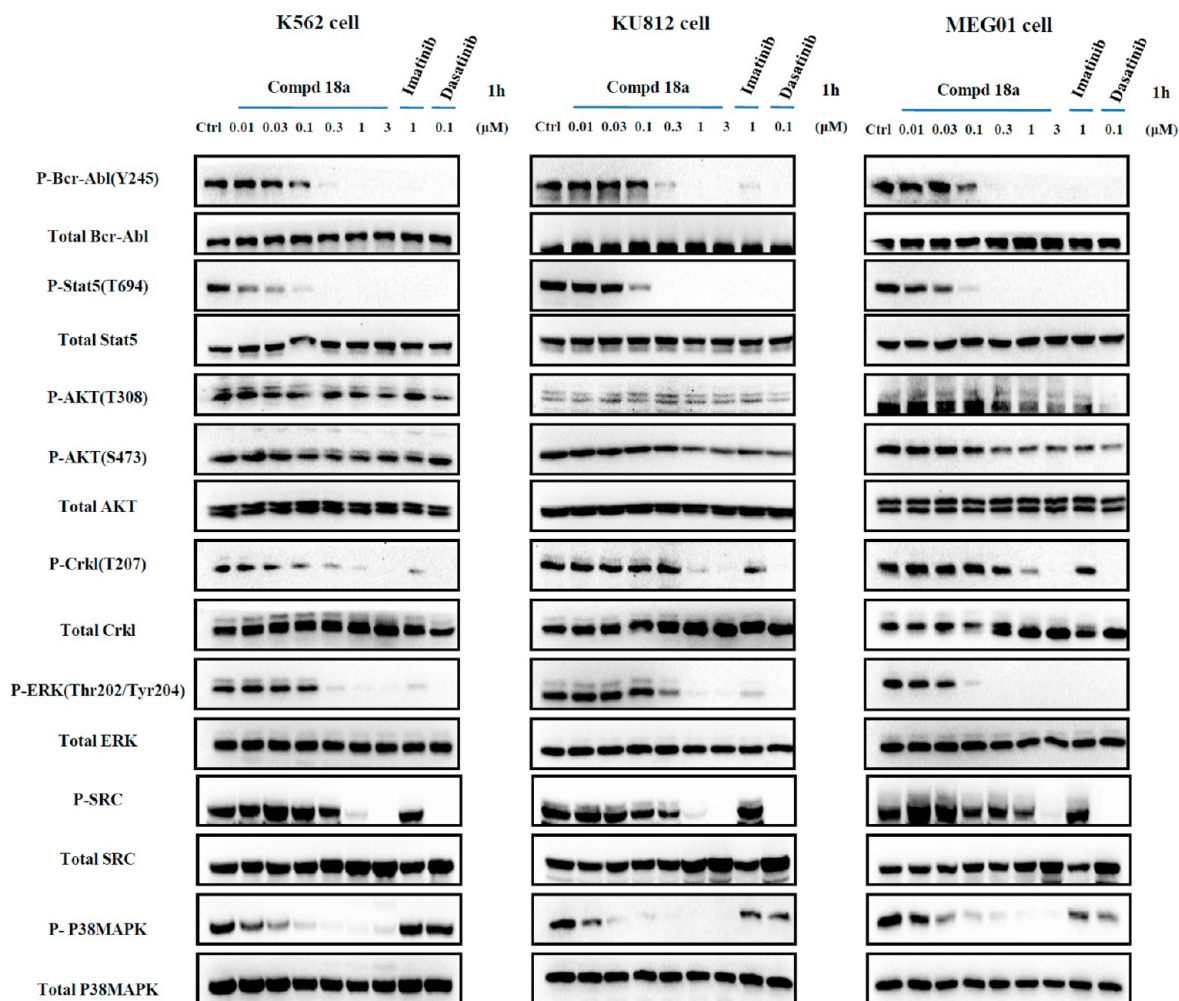
The in vivo antitumor study of **18a** was performed in the K562 cell inoculated xenograft mouse model. After 16 days of continuous treatment, compound **18a** dose-dependently inhibited the growth of the K562 tumor, and a 50 mg/kg/day dosage could almost completely suppress tumor progression (Figure 8A). All doses of **18a** were well tolerated, with no mortality and no significant body weight loss observed (Figure 8B). Compound **18a** displayed obvious antitumor efficacy (TGI = 48.3%) at 50 mg/kg/day dosage (Figure 8C,D). In addition, the immunohistochemistry (IHC) stain

revealed that the proliferation was effectively inhibited ( $K_i$ -67 lane), and significant apoptosis was induced (TUNEL lane) in the tumor (Figure 8E).

## CONCLUSIONS

Starting from a multiple target dihydropyrimidopyrimidine scaffold based compound **27** (GNF-7) bearing high BCR-ABL potency, we used a focused medicinal chemistry approach guided by computer-aided drug design to obtain an aminopyrimidine scaffold based compound **18a** (CHMFL-ABL-053) via 7 steps of chemical syntheses (11% overall yield), which possessed highly potent antiproliferative efficacy against BCR-ABL driven CML cell lines and exhibited a good safety window against other leukemia cell lines such as AML. Compound **18a** displayed a high selectivity profile. Besides the ABL kinase, it also inhibited structurally related SRC kinase and p38 $\alpha$  kinase, which is desired and might contribute positively to exert synergistic antileukemic effect of **18a** since SRC kinase is downstream of BCR-ABL and contributes to the proliferation and survival of the myeloid cell line, and p38 $\alpha$  kinase is involved in the BCR-ABL inhibitor induced apoptosis pathway.<sup>18,19</sup> Compared to clinically used BCR-ABL inhibitors Imatinib, Nilotinib, Bosutinib, and Dasatinib, compound **18a** completely abolished the c-KIT kinase inhibitory activity and exhibited better selectivity against DDR1/2 kinases. In addition, **18a** also showed a suitable PK profile and potent in vivo antitumor efficacy. It is worthy of note that although compound **18a** exhibits a good selectivity among kinome, we cannot exclude the possibility that it may also affect other effective targets that may also contribute to its antileukemia effect; for instance, Imatinib has been reported to inhibit the quinone oxidoreductase2 (NQO2).<sup>20</sup> Currently, **18a** is under extensive preclinical safety evaluation, and it might be a potential useful





**Figure 6.** Compound 18a's effects on the BCR-Abl kinase mediated signaling pathway in KU812, K562, and MEG-01 cancer cell lines. Cells were treated with 18a at the indicated concentrations for 1 h, and whole cell lysates were then subjected to Western blot analyses.

pharmacological candidate supplementary to the current BCR-ABL target therapies for the treatment of CML.

## EXPERIMENTAL PROCEDURES

**Chemical Synthesis.** All reagents and solvents were purchased from commercial sources and were used as received, unless specified otherwise, or prepared as described in the literature. All moisture-sensitive reactions were carried out using dry solvents under ultrapure argon protection. Glassware was dried in an oven at 140 °C for at least 12 h prior to use and then assembled quickly while hot, sealed with rubber septa, and allowed to cool under a stream of argon. Reactions were stirred magnetically using Teflon-coated magnetic stirring bars. Commercially available disposable syringes were used for transferring the reagents and solvents. LC/MS were performed on an Agilent 6224 TOF using an ESI source coupled to an Agilent 1260 Infinity HPLC system operating in reverse mode with an Agilent XDB-C18 column (4.6 × 50 mm, 1.8 μm) using a water/acetonitrile (each with 0.2% (v/v) formic acid) gradient at a flow rate at 0.4 mL/min. <sup>1</sup>H and <sup>13</sup>C spectra were recorded on a Bruker 400 MHz NMR spectrometer. Chemical shifts are expressed in ppm. In the NMR tabulation, s indicates singlet; d, doublet; t, triplet; q, quartet; and m, multiplet. Flash column chromatography was conducted using silica gel (Silicycle 40–64 μm). The purities of all compounds were determined to be >95% by HPLC.

**General Method A.** *N*-(3-Amino-4-methylphenyl)-3-(trifluoromethyl)benzamide (**3a**). To a solution of 4-methyl-3-nitroaniline (2.82 g, 18.5 mmol, 1.00 equiv) in DCM (30 mL) was added TEA (2.84 mL, 20.35 mmol, 1.10 equiv). Then a solution of 3-

(trifluoromethyl)benzoyl chloride (5.0 g, 19.5 mmol, 1.05 equiv) in DCM (15 mL) was dropwise added in 30 min at 0 °C under argon. The reaction mixture was stirred at 0 °C for 2 h, then was allowed to warm to room temperature overnight (10 h). The resulting solution was diluted with DCM (100 mL) and EtOAc (20 mL), washed with 1 M HCl (2 × 100 mL), 1 M NaOH (2 × 100 mL) and brine (100 mL), and dried over anhydrous MgSO<sub>4</sub>. Evaporation of the solvent afforded the crude product **2a**, which was used in the next step without further purification. To a solution of **2a** (18.5 mmol, 1.00 equiv) in methanol (20 mL) was added 10% Pd/C (0.2 g) at room temperature under argon. Then, the reaction mixture was stirred under a balloon of hydrogen for 20 h. The resulting mixture was filtered through a pad of Celite and washed with methanol. Evaporation of the filtrate provided the crude product, which was purified by silica gel flash chromatography (eluting with MeOH in DCM 0–4%) to give **3a** as a white solid (2.83 g, two steps yield 54%). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 9.97 (s, 1H), 8.30 (d, *J* = 14.1 Hz, 2H), 7.95 (d, *J* = 7.1 Hz, 1H), 7.78 (t, *J* = 7.2 Hz, 1H), 6.93 (d, *J* = 7.7 Hz, 1H), 6.64 (s, 1H), 6.47 (d, *J* = 7.7 Hz, 1H), 4.97 (s, 2H), 2.09 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 164.12, 147.36, 136.66, 136.15, 132.09, 130.95, 130.15, 129.90, 129.58, 128.39, 124.68, 120.93, 112.92, 112.85, 17.39. LC-MS (ESI, *m/z*): 295.0992 [M + H]<sup>+</sup>.

*N*-(3-Amino-4-methylphenyl)biphenyl-3-carboxamide (**3b**). (Method A) Yield 75%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.09 (s, 1H), 8.24 (s, 1H), 7.96 (d, *J* = 7.4 Hz, 1H), 7.85 (d, *J* = 7.4 Hz, 1H), 7.78 (d, *J* = 7.1 Hz, 2H), 7.62 (d, *J* = 7.5 Hz, 1H), 7.43 (d, *J* = 7.1 Hz, 1H), 7.22 (s, 1H), 6.92 (s, 2H), 4.92 (s, 2H), 2.08 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 165.48, 147.02, 140.69, 140.10, 138.07,

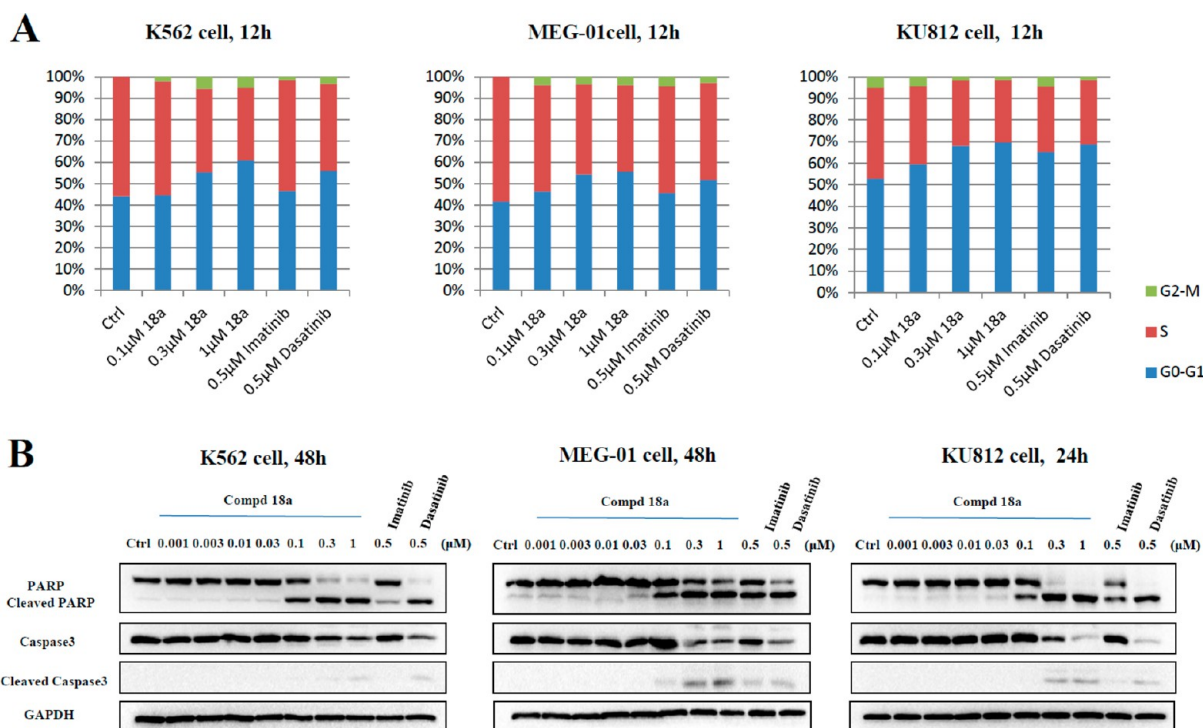


Figure 7. (A) Compound 18a arrested the cell cycle progression. (B) Compound 18a induced apoptosis in K562, KU812, and MEG-01 cell lines.

Table 6. Pharmacokinetic Study of 18a on Sprague Dawley Rats<sup>a</sup>

	$t_{1/2}$ (h)	$T_{max}$ (h)	$C_{max}$ (ng/mL)	$AUC_{(0-t)}$ (ng/mL·h)	$AUC_{(0-\infty)}$ (ng/mL·h)	$V_z$ mL/kg	$CL_z$ mL/h/kg	$MRT_{(0-\infty)}$ (h)	F %
IV 1 mg/kg mean	2.82	0.02	2395.22	635.98	720.69	6025.13	1538.39	3.1	NA
SD ( $n = 3$ )	0.53	0.00	449.56	208.84	256.99	1632.22	642.39	0.69	NA
PO 10 mg/kg mean	4.33	1.00	367.61	1715.51	1743.43	NA	NA	5.51	24.19
SD ( $n = 3$ )	1.11	0.87	202.21	1083.42	1091.08	NA	NA	0.50	NA

<sup>a</sup>Compound 18a was formulated as a clear solution in 5% DMSO, 40% PEG400, and 55% of 20% HP- $\beta$ -CD in water for intravenous and oral administration.

136.52, 130.20, 129.92, 129.47, 128.26, 127.40, 127.28, 126.23, 117.41, 109.47, 107.08, 17.50. LC-MS (ESI,  $m/z$ ): 303.1425  $[M + H]^+$ .

***N*-(3-Amino-4-methylphenyl)-3,5-dimethylbenzamide (3c).** (Method A) Yield 72%. <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ )  $\delta$  9.86 (s, 1H), 7.57 (s, 2H), 7.20 (s, 1H), 7.17 (s, 1H), 6.88 (s, 2H), 4.86 (s, 2H), 2.36 (s, 6H), 2.06 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO- $d_6$ )  $\delta$  165.84, 146.92, 138.20, 137.86, 135.90, 132.95, 130.14, 125.78, 117.20, 109.37, 106.95, 21.34, 17.46. LC-MS (ESI,  $m/z$ ): 255.1425  $[M + H]^+$ .

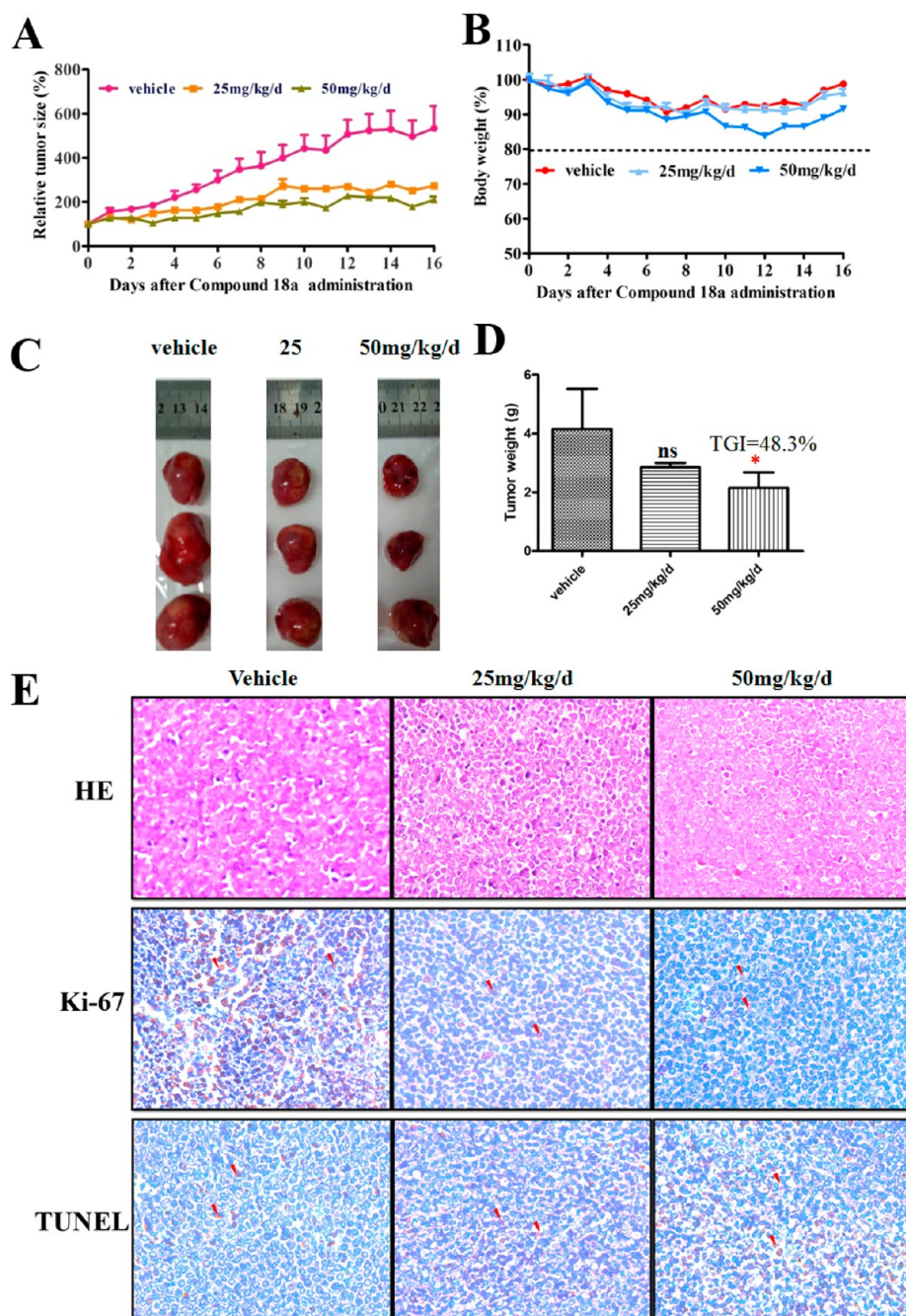
***N*-(3-Amino-4-methylphenyl)-3,5-di-*tert*-butylbenzamide (3d).** (Method A) Yield 71%. <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ )  $\delta$  9.92 (s, 1H), 7.77 (s, 2H), 7.59 (s, 1H), 7.11 (s, 1H), 6.89 (s, 2H), 2.07 (s, 3H), 1.34 (d,  $J = 13.6$  Hz, 18H). <sup>13</sup>C NMR (101 MHz, DMSO- $d_6$ )  $\delta$  166.31, 150.80, 146.92, 138.04, 135.41, 130.08, 125.26, 122.13, 117.31, 109.82, 107.46, 35.16, 31.68, 17.48. LC-MS (ESI,  $m/z$ ): 339.2365  $[M + H]^+$ .

***N*-(3-Amino-4-methylphenyl)-3-methylbenzamide (3e).** (Method A) Yield 65%. <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ )  $\delta$  10.50 (s, 1H), 8.07 (s, 1H), 7.79 (dd,  $J = 8.2, 4.8$  Hz, 2H), 7.65 (d,  $J = 8.2$  Hz, 1H), 7.41 (d,  $J = 4.5$  Hz, 2H), 7.29 (d,  $J = 8.3$  Hz, 1H), 2.40 (s, 3H), 2.36 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO- $d_6$ )  $\delta$  166.22, 138.45, 138.17, 135.05, 132.76, 131.70, 130.93, 128.75, 128.70, 127.37, 125.41, 120.43, 115.90, 21.43, 17.20. LC-MS (ESI,  $m/z$ ): 241.1272  $[M + H]^+$ .

**Ethyl 4-(Methylamino)-2-(methylthio)pyrimidine-5-carboxylate (5).** To a solution of ethyl 4-chloro-2-(methylthio)pyrimidine-5-carboxylate (5.00 g, 21.5 mol, 1.0 equiv) in THF (100 mL) was added TEA (6.9 mL, 49.45 mmol, 2.3 equiv) and methylamine hydrochloride (2.70 g, 49.45 mmol, 2.3 equiv) at 0 °C under argon. The reaction mixture was stirred at 0 °C for 1 h, then it was allowed to warm to room temperature overnight (14 h). The reaction mixture was

concentrated to remove the THF. The residue was diluted with water (100 mL) and extracted with EtOAc (3  $\times$  80 mL). The combined organic layers were washed with water (2  $\times$  80 mL) and brine (80 mL), and dried over anhydrous MgSO<sub>4</sub>. Evaporation of the solvent afforded the crude 5 as a white solid (4.6 g, 94%). <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ )  $\delta$  8.50 (d,  $J = 1.7$  Hz, 1H), 8.23 (s, 1H), 4.37–4.20 (m, 2H), 2.97 (dd,  $J = 4.8, 1.6$  Hz, 3H), 2.48 (d,  $J = 1.7$  Hz, 3H), 1.30 (td,  $J = 7.1, 1.6$  Hz, 3H). <sup>13</sup>C NMR (101 MHz, DMSO- $d_6$ )  $\delta$  175.41, 166.23, 160.16, 158.09, 101.25, 60.98, 40.65, 40.44, 40.23, 40.02, 39.82, 39.61, 39.40, 27.75, 14.50, 14.01. LC-MS (ESI,  $m/z$ ): 228.0730  $[M + H]^+$ .

**4-(Methylamino)-2-(methylthio)pyrimidine-5-carbaldehyde (7).** To a solution of ethyl 4-(methylamino)-2-(methylthio)pyrimidine-5-carboxylate (3.30 g, 14.5 mmol, 1.00 equiv) in anhydrous THF (30 mL) was added LAH (2.4 M in THF, 8.7 mL, 17.45 mmol, 1.20 equiv) at 0 °C under argon. The reaction mixture was then stirred at 0 °C for 1 h and was slowly warmed to room temperature for 14 h. The resulting mixture was concentrated to remove the solvent. The residue was quenched with ice-cold water and extracted with DCM (3  $\times$  50 mL). The combined organic layers were washed with water (2  $\times$  50 mL) and brine (50 mL), and dried over anhydrous MgSO<sub>4</sub>. Evaporation of the solvent afforded the crude 6. To a solution of 6 (14.5 mmol, 1.00 equiv) in anhydrous DCM (60 mL) was added activated MnO<sub>2</sub> (12.6 g, 145.0 mmol, 10 equiv) at room temperature under argon. Then, the reaction mixture was stirred at room temperature for 6 h. The resulting mixture was filtered and washed with DCM (2  $\times$  30 mL). Evaporation of the solvent afforded the crude product, which was purified by flash column chromatography (eluting with MeOH in DCM 0–2%) to give 7 as a yellow solid (1.95 g, two



**Figure 8.** Compound 18a's antitumor efficacy in the K562 xenograft model. Female nu/nu mice bearing an established control group and K562 tumor xenografts were treated with 18a at 25.0, 50.0 mg/kg/d, or vehicle. Daily oral administration was initiated when K562 tumors had reached a size of 200 to 400 mm<sup>3</sup>. Each group contained 5 animals. Data = mean ± SEM. (A) Tumor size measurement from K562 xenograft mice after 18a administration. Initial tumor size was set as 100%. (B) Body weight measurement from K562 xenograft mice after 18a administration. Initial body weight was set as 100%. (C) Representative photographs of tumors in each group after 25.0 or 50.0 mg/kg/d 18a or vehicle treatment. (D) Comparison of the final tumor weight in each group after a 16-day treatment period. (E) Representative micrographs of hematoxylin and eosin (HE), Ki-67, and TUNEL staining of tumor tissues with 18a treatment compared to the vehicle group. Note the specific nuclear staining of cells with morphology consistent with proliferation and apoptosis (E, red arrow).

steps yield 74%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 9.68 (s, 1H), 8.27 (s, 1H), 3.09 (d, J = 5.0 Hz, 3H), 2.54 (s, 3H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 190.79, 177.48, 162.76, 159.43, 109.43, 27.10, 14.27. LC-MS (ESI, m/z): 184.0470 [M + H]<sup>+</sup>.

**General Method B.** *N*-(4-Methyl-3-((4-(methylamino)-2-(methylthio)pyrimidin-5-yl)methylamino)phenyl)-3-(trifluoromethyl)benzamide (8a). To a solution of 4-(methylamino)-2-(methylthio)pyrimidine-5-carbaldehyde (1.10 g, 6.0 mmol, 1.0 equiv) and *N*-(3-amino-4-methylphenyl)-3-(trifluoromethyl)-

benzamide (1.76 g, 6.0 mmol, 1.0 equiv) in methanol (30 mL) was added acetic acid (0.7 mL, 12.0 mmol, 2.00 equiv) at 0 °C under argon. The reaction mixture was stirred at 0 °C for 30 min, then NaBH<sub>3</sub>(CN) was added portion-wise. The reaction mixture was stirred at 0 °C for 1 h, then was allowed to warm to room temperature for 24 h. The resulting mixture was concentrated to remove the solvent. The residue was diluted with water (50 mL) and extracted with EtOAc (3 × 50 mL). The combined organic layers were washed with water (50 mL) and brine (50 mL), and dried over anhydrous MgSO<sub>4</sub>. Evaporation of

the solvent afforded the crude product, which was purified by flash column chromatography (eluting with MeOH in DCM 0–2%) to yield **8a** as an off white solid (2.53 g, 92%). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.22 (s, 1H), 8.25 (s, 3H), 7.93 (s, 1H), 7.88 (s, 1H), 7.76 (s, 1H), 7.14 (s, 1H), 7.06 (s, 1H), 6.99 (s, 2H), 4.13 (s, 2H), 2.93 (s, 3H), 2.42 (s, 3H), 2.13 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 168.82, 164.25, 160.60, 152.67, 146.42, 138.25, 136.72, 132.20, 130.17, 130.02, 128.30, 124.69, 118.37, 110.91, 108.88, 102.51, 41.02, 28.01, 17.76, 13.76. LC-MS (ESI, *m/z*): 462.1453 [M + H]<sup>+</sup>.

*N*-(4-Methyl-3-((4-(methylamino)-2-(methylthio)pyrimidin-5-yl)methylamino)phenyl) Biphenyl 1-3-carboxamide (**8b**). (Method B) Yield 74%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.08 (s, 1H), 8.16 (s, 1H), 7.87 (dd, *J* = 14.4, 7.4 Hz, 3H), 7.77 (d, *J* = 7.3 Hz, 2H), 7.62–7.57 (m, 1H), 7.56–7.48 (m, 3H), 7.43 (d, *J* = 7.0 Hz, 1H), 7.14 (s, 1H), 7.06 (d, *J* = 7.8 Hz, 1H), 7.01 (s, 1H), 6.96 (d, *J* = 7.9 Hz, 1H), 5.44 (s, 1H), 4.11 (s, 2H), 2.91 (d, *J* = 4.0 Hz, 3H), 2.41 (s, 3H), 2.11 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 168.79, 165.67, 160.57, 152.69, 146.37, 140.70, 140.07, 138.59, 136.57, 130.14, 129.95, 129.48, 128.27, 127.40, 127.22, 126.25, 118.03, 110.95, 108.76, 102.45, 41.10, 27.81, 17.75, 13.78. LC-MS (ESI, *m/z*): 470.1943 [M + H]<sup>+</sup>.

3,5-Dimethyl-*N*-(4-methyl-3-((4-(methylamino)-2-(methylthio)pyrimidin-5-yl)methylamino)phenyl)benzamide (**8c**). (Method B) Yield 88%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 9.89 (s, 1H), 7.88 (s, 1H), 7.51 (s, 2H), 7.19 (s, 1H), 7.14 (s, 1H), 7.06 (d, *J* = 7.8 Hz, 1H), 6.99 (s, 1H), 6.93 (d, *J* = 7.8 Hz, 1H), 5.42 (s, 1H), 4.10 (s, 2H), 2.92 (d, *J* = 2.9 Hz, 3H), 2.42 (s, 3H), 2.35 (s, 6H), 2.10 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 168.78, 165.99, 160.58, 152.73, 146.30, 138.71, 137.88, 135.93, 132.97, 130.08, 125.74, 117.78, 110.96, 108.61, 102.30, 41.11, 27.80, 21.33, 17.73, 13.88. LC-MS (ESI, *m/z*): 422.1947 [M + H]<sup>+</sup>.

3,5-Di-*tert*-butyl-*N*-(4-methyl-3-((4-(methylamino)-2-(methylthio)pyrimidin-5-yl)methylamino)phenyl)benzamide (**8d**). (Method B) Yield 59%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 9.95 (s, 1H), 7.88 (s, 1H), 7.69 (s, 2H), 7.59 (s, 1H), 7.14 (s, 1H), 7.04 (s, 1H), 6.95 (s, 2H), 5.42 (s, 1H), 4.12 (s, 2H), 2.92 (s, 3H), 2.42 (s, 3H), 2.12 (s, 3H), 1.35 (s, 18H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 168.82, 166.72, 160.60, 152.81, 150.84, 146.33, 138.61, 135.61, 130.06, 125.36, 122.08, 117.94, 110.98, 109.07, 102.81, 41.12, 35.16, 31.67, 27.81, 17.77, 13.79. LC-MS (ESI, *m/z*): 506.2882 [M + H]<sup>+</sup>.

3-Methyl-*N*-(4-methyl-3-((4-(methylamino)-2-(methylthio)pyrimidin-5-yl)methylamino)phenyl)benzamide (**8e**). (Method B) Yield 69%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 9.93 (s, 1H), 7.87 (s, 1H), 7.72 (s, 2H), 7.38 (s, 2H), 7.13 (s, 1H), 7.07 (d, *J* = 7.1 Hz, 1H), 6.99 (s, 1H), 6.94 (d, *J* = 7.7 Hz, 1H), 5.43 (s, 1H), 4.10 (s, 2H), 2.92 (s, 3H), 2.42 (s, 3H), 2.40 (s, 3H), 2.10 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 168.78, 165.88, 160.57, 152.71, 146.32, 138.67, 138.04, 135.91, 132.27, 130.10, 128.64, 128.52, 125.19, 117.86, 110.95, 108.62, 102.32, 41.10, 27.81, 21.43, 17.73, 13.79. LC-MS (ESI, *m/z*): 408.1791 [M + H]<sup>+</sup>.

**General Method C.** *N*-(4-Methyl-3-(1-methyl-7-(methylthio)-2-oxo-1,2-dihydropyrimido[4,5-*d*]pyrimidin-3(4H)-yl)phenyl)-3-(trifluoromethyl)benzamide (**9a**). To a solution of *N*-(4-methyl-3-((4-(methylamino)-2-(methylthio)pyrimidin-5-yl)methylamino)phenyl)-3-(trifluoromethyl)benzamide (2.40 g, 5.21 mmol, 1.00 equiv) in anhydrous dioxane (26 mL) was added DIPEA (2.6 mL, 15.63 mmol, 3.00 equiv) at 0 °C under argon. Then, a solution of triphosgene (0.53 g, 1.77 mmol, 0.34 equiv) in dioxane (10 mL) was added. The reaction mixture was stirred at 0 °C for 1 h, then was allowed to warm to room temperature for 24 h. The resulting mixture was concentrated to remove the solvent. The residue was diluted with water (50 mL) and extracted with EtOAc (3 × 50 mL). The combined organic layers were washed with water (50 mL) and brine (50 mL), and dried over anhydrous MgSO<sub>4</sub>. Evaporation of the solvent afforded the crude product, which was purified by flash column chromatography (eluting with MeOH in DCM 0–2%) to yield **9a** as a yellow solid (1.05 g, 42%). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.59 (s, 1H), 8.33 (s, 1H), 8.30 (d, *J* = 7.7 Hz, 1H), 8.26 (s, 1H), 7.96 (d, *J* = 7.3 Hz, 1H), 7.86 (s, 1H), 7.79 (t, *J* = 7.5 Hz, 1H), 7.67 (d, *J* = 8.1 Hz, 1H), 7.31 (d, *J* = 8.1 Hz, 1H), 4.78 (d, *J* = 14.5 Hz, 1H), 4.59 (d, *J* = 14.6 Hz, 1H), 3.40 (s, 3H), 2.53 (s, 3H), 2.14 (s, 3H). <sup>13</sup>C NMR (101

MHz, DMSO-*d*<sub>6</sub>) δ 170.39, 164.32, 156.84, 152.73, 152.23, 141.37, 138.13, 136.06, 132.26, 131.37, 131.21, 130.16, 128.56, 124.68, 120.41, 119.79, 108.22, 60.20, 28.48, 17.29, 14.36. LC-MS (ESI, *m/z*): 488.1235 [M + H]<sup>+</sup>.

*N*-(4-Methyl-3-(1-methyl-7-(methylthio)-2-oxo-1,2-dihydropyrimido[4,5-*d*]pyrimidin-3(4H)-yl)phenyl)biphenyl-3-carboxamide (**9b**). (Method C) Yield 87%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.41 (s, 1H), 8.26 (d, *J* = 11.7 Hz, 2H), 7.96 (d, *J* = 7.0 Hz, 1H), 7.90 (d, *J* = 7.0 Hz, 1H), 7.86 (s, 1H), 7.79 (d, *J* = 7.0 Hz, 2H), 7.65 (dd, *J* = 16.8, 8.1 Hz, 2H), 7.52 (d, *J* = 7.1 Hz, 2H), 7.44 (d, *J* = 6.6 Hz, 1H), 7.32 (d, *J* = 8.1 Hz, 1H), 4.79 (d, *J* = 14.5 Hz, 1H), 4.60 (d, *J* = 14.5 Hz, 1H), 3.34 (s, 3H), 2.55 (s, 3H), 2.15 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 170.38, 165.75, 156.90, 152.77, 152.24, 141.38, 140.81, 139.99, 138.45, 135.93, 131.19, 131.05, 130.27, 129.63, 129.50, 128.33, 127.40, 127.28, 126.22, 120.31, 119.69, 108.31, 46.93, 28.52, 17.21, 14.07. LC-MS (ESI, *m/z*): 496.1730 [M + H]<sup>+</sup>.

3,5-Dimethyl-*N*-(4-methyl-3-(1-methyl-7-(methylthio)-2-oxo-1,2-dihydropyrimido[4,5-*d*]pyrimidin-3(4H)-yl)phenyl)benzamide (**9c**). (Method C) Yield 75%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.23 (s, 1H), 8.26 (s, 1H), 7.86 (s, 1H), 7.66 (d, *J* = 7.9 Hz, 1H), 7.58 (s, 2H), 7.28 (d, *J* = 8.0 Hz, 1H), 7.21 (s, 1H), 4.77 (d, *J* = 14.5 Hz, 1H), 4.58 (d, *J* = 14.6 Hz, 1H), 3.32 (s, 3H), 2.53 (s, 3H), 2.36 (s, 6H), 2.13 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 170.38, 166.08, 156.83, 152.72, 152.20, 141.31, 138.63, 138.00, 135.29, 133.30, 131.12, 130.78, 125.78, 120.10, 119.39, 108.23, 60.22, 46.92, 28.50, 21.33, 17.19, 14.06. LC-MS (ESI, *m/z*): 448.1735 [M + H]<sup>+</sup>.

3,5-Di-*tert*-butyl-*N*-(4-methyl-3-(1-methyl-7-(methylthio)-2-oxo-1,2-dihydropyrimido[4,5-*d*]pyrimidin-3(4H)-yl)phenyl)benzamide (**9d**). (Method C) Yield 80%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.12 (s, 1H), 8.12 (s, 1H), 7.63 (s, 3H), 7.52 (d, *J* = 8.2 Hz, 1H), 7.47 (s, 1H), 7.16 (d, *J* = 8.1 Hz, 1H), 4.62 (d, *J* = 14.4 Hz, 1H), 4.45 (d, *J* = 14.7 Hz, 1H), 3.18 (s, 3H), 2.38 (s, 3H), 1.99 (s, 3H), 1.20 (s, 18H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 170.41, 166.66, 156.84, 152.75, 152.27, 151.03, 141.27, 138.43, 134.72, 131.13, 130.99, 125.73, 122.13, 120.62, 120.01, 108.21, 46.61, 35.05, 31.51, 28.23, 17.28, 14.00. LC-MS (ESI, *m/z*): 532.2672 [M + H]<sup>+</sup>.

3-Methyl-*N*-(4-methyl-3-(1-methyl-7-(methylthio)-2-oxo-1,2-dihydropyrimido[4,5-*d*]pyrimidin-3(4H)-yl)phenyl)benzamide (**9e**). (Method C) Yield 75%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.27 (s, 1H), 8.26 (s, 1H), 7.85 (s, 1H), 7.78 (s, 1H), 7.77–7.74 (m, 1H), 7.64 (d, *J* = 7.9 Hz, 1H), 7.41 (s, 2H), 7.29 (d, *J* = 8.0 Hz, 1H), 4.77 (d, *J* = 14.3 Hz, 1H), 4.58 (d, *J* = 14.6 Hz, 1H), 3.32 (s, 3H), 2.53 (s, 3H), 2.40 (s, 3H), 2.13 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 170.37, 166.00, 156.85, 152.74, 152.22, 141.32, 138.54, 138.17, 135.27, 132.62, 131.15, 130.88, 128.77, 128.52, 125.24, 120.18, 119.47, 108.26, 46.91, 28.51, 21.43, 17.20, 14.06. LC-MS (ESI, *m/z*): 434.1581 [M + H]<sup>+</sup>.

**General Method D.** *N*-(4-Methyl-3-(1-methyl-7-(methylsulfonyl)-2-oxo-1,2-dihydropyrimido[4,5-*d*]pyrimidin-3(4H)-yl)phenyl)-3-(trifluoromethyl)benzamide (**10a**). To a solution of *N*-(4-methyl-3-(1-methyl-7-(methylthio)-2-oxo-1,2-dihydropyrimido[4,5-*d*]pyrimidin-3(4H)-yl)phenyl)-3-(trifluoromethyl)benzamide (480 mg, 1.00 mmol, 1.00 equiv) in DCM (20 mL) was added *m*-CBPA (362 mg, 2.10 mmol, 2.10 equiv) at 0 °C under argon. The reaction mixture was stirred at 0 °C for 2 h, then was allowed to warm to room temperature for 20 h. The resulting mixture was diluted with DCM (30 mL), washed with water (30 mL) and brine (30 mL), and dried over anhydrous MgSO<sub>4</sub>. Evaporation of the solvent afforded the crude product, which was purified by flash chromatography (eluting with MeOH in DCM 0–2%) to give the title compound **10a** as a white solid (0.36 g, 70%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.99 (s, 1H), 8.30 (s, 1H), 8.15 (s, 1H), 8.11 (d, *J* = 7.9 Hz, 1H), 7.81 (d, *J* = 1.7 Hz, 1H), 7.78 (d, *J* = 7.8 Hz, 1H), 7.60 (t, *J* = 7.8 Hz, 1H), 7.29 (s, 1H), 7.06 (d, *J* = 8.4 Hz, 1H), 4.85 (d, *J* = 16.0 Hz, 1H), 4.61 (d, *J* = 15.9 Hz, 1H), 3.54 (s, 3H), 3.37 (s, 3H), 1.74 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 166.13, 164.66, 158.22, 153.02, 151.52, 140.93, 138.63, 138.04, 135.30, 133.34, 131.21, 130.85, 125.77, 120.34, 119.41, 115.96, 47.17, 28.94, 21.34, 17.06. LC-MS (ESI, *m/z*): 520.1123 [M + H]<sup>+</sup>.

*N*-(4-Methyl-3-(1-methyl-7-(methylsulfonyl)-2-oxo-1,2-dihydropyrimido[4,5-*d*]pyrimidin-3(4H)-yl)phenyl)biphenyl-3-car-

**boxamide (10b).** (Method D) Yield 82%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 8.62 (d, *J* = 4.2 Hz, 1H), 8.26 (d, *J* = 4.3 Hz, 1H), 8.04–7.88 (m, 3H), 7.86–7.75 (m, 2H), 7.74–7.62 (m, 2H), 7.60–7.50 (m, 2H), 7.48–7.40 (m, 1H), 7.40–7.29 (m, 1H), 5.06–4.91 (m, 1H), 4.79 (dd, *J* = 15.2, 4.4 Hz, 1H), 3.44 (d, *J* = 4.2 Hz, 3H), 3.38 (t, *J* = 7.2 Hz, 3H), 2.17 (d, *J* = 4.0 Hz, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 165.70, 164.68, 158.24, 153.03, 151.55, 141.00, 140.82, 140.00, 138.41, 135.88, 131.29, 131.10, 130.31, 129.66, 129.51, 128.35, 127.41, 127.30, 126.24, 120.42, 119.57, 115.98, 47.19, 28.97, 17.21. LC-MS (ESI, *m/z*): 528.1630 [M + H]<sup>+</sup>.

**3,5-Dimethyl-N-(4-methyl-3-(1-methyl-7-(methylsulfonyl)-2-oxo-1,2-dihydropyrimido[4,5-*d*]pyrimidin-3(4H)-yl)phenyl)benzamide (10c).** (Method D) Yield 83%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.24 (s, 1H), 8.60 (s, 1H), 7.89 (s, 1H), 7.63 (d, *J* = 8.2 Hz, 1H), 7.57 (s, 2H), 7.30 (d, *J* = 8.2 Hz, 1H), 7.23 (s, 1H), 4.95 (d, *J* = 15.3 Hz, 1H), 4.77 (d, *J* = 15.6 Hz, 1H), 3.42 (s, 3H), 3.38 (s, 3H), 2.37 (s, 6H), 2.15 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 166.13, 164.66, 158.22, 153.02, 151.52, 140.93, 138.63, 138.04, 135.30, 133.34, 131.21, 130.85, 125.77, 120.34, 119.41, 115.96, 47.17, 39.48, 28.94, 21.34, 17.17. LC-MS (ESI, *m/z*): 480.1630 [M + H]<sup>+</sup>.

**3,5-Di-*tert*-butyl-N-(4-methyl-3-(1-methyl-7-(methylsulfonyl)-2-oxo-1,2-dihydropyrimido[4,5-*d*]pyrimidin-3(4H)-yl)phenyl)benzamide (10d).** (Method D) Yield 76%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.26 (s, 1H), 8.61 (s, 1H), 7.82 (s, 1H), 7.78 (s, 2H), 7.67 (d, *J* = 8.3 Hz, 1H), 7.62 (s, 1H), 7.33 (d, *J* = 8.2 Hz, 1H), 4.96 (d, *J* = 15.4 Hz, 1H), 4.79 (d, *J* = 15.4 Hz, 1H), 3.43 (d, *J* = 1.2 Hz, 3H), 3.38 (s, 3H), 2.16 (s, 3H), 1.36 (d, *J* = 1.1 Hz, 18H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 166.60, 164.68, 158.23, 153.02, 151.55, 151.01, 140.93, 138.52, 134.77, 131.18, 130.99, 125.75, 122.13, 120.82, 120.01, 115.97, 59.99, 46.74, 35.02, 31.51, 29.03, 17.30. LC-MS (ESI, *m/z*): 564.2570 [M + H]<sup>+</sup>.

**3-Methyl-N-(4-methyl-3-(1-methyl-7-(methylsulfonyl)-2-oxo-1,2-dihydropyrimido[4,5-*d*]pyrimidin-3(4H)-yl)phenyl)benzamide (10e).** (Method D) Yield 80%. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.43 (s, 1H), 7.82 (s, 1H), 7.73 (s, 1H), 7.69 (d, *J* = 2.8 Hz, 1H), 7.49 (dd, *J* = 6.0, 2.0 Hz, 1H), 7.37 (d, *J* = 4.3 Hz, 2H), 7.27 (d, *J* = 8.3 Hz, 1H), 4.96 (d, *J* = 15.8 Hz, 1H), 4.72 (d, *J* = 15.8 Hz, 1H), 3.51 (s, 3H), 3.39 (s, 3H), 2.43 (s, 3H), 2.19 (s, 3H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 170.27, 167.56, 161.26, 155.42, 154.84, 142.74, 141.31, 140.71, 137.51, 135.51, 134.35, 134.26, 131.33, 130.92, 127.38, 123.91, 122.30, 118.07, 50.10, 41.84, 31.71, 23.95, 19.71. LC-MS (ESI, *m/z*): 466.1480 [M + H]<sup>+</sup>.

**General Method E. N-(4-Methyl-3-(1-methyl-7-(4-methyl-3-nitrophenylamino)-2-oxo-1,2-dihydropyrimido[4,5-*d*]pyrimidin-3(4H)-yl)phenyl)-3-(trifluoromethyl)benzamide (11a).** To a solution of N-(4-methyl-3-(1-methyl-7-(methylsulfonyl)-2-oxo-1,2-dihydropyrimido[4,5-*d*]pyrimidin-3(4H)-yl)phenyl)-3-(trifluoromethyl)benzamide (450 mg, 0.88 mmol, 1.0 equiv) in anhydrous dioxane (2 mL) was added 4-methyl-3-nitroaniline (1.3 g, 8.80 mmol, 10.0 equiv) and TFA (0.32 mL, 8.80 mmol, 10.0 equiv) at room temperature under argon. The reaction mixture was then heated to 120 °C for 2 h. The resulting mixture was diluted with DCM (30 mL), washed with water (30 mL) and brine (30 mL), and dried over anhydrous MgSO<sub>4</sub>. Evaporation of the solvent afforded the crude product, which was purified by flash chromatography (eluting with MeOH in DCM 0–2%) to give the title compound **11a** as a white solid (320 mg, 61%). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.55 (s, 1H), 10.01 (s, 1H), 8.78 (s, 1H), 8.32 (s, 1H), 8.28 (d, *J* = 7.2 Hz, 1H), 8.21 (s, 1H), 7.98 (d, *J* = 6.8 Hz, 1H), 7.81 (d, *J* = 13.0 Hz, 3H), 7.66 (d, *J* = 7.6 Hz, 1H), 7.40 (d, *J* = 8.1 Hz, 1H), 7.32 (d, *J* = 7.9 Hz, 1H), 4.74 (d, *J* = 13.8 Hz, 1H), 4.56 (d, *J* = 13.9 Hz, 1H), 3.39 (s, 3H), 2.47 (s, 3H), 2.15 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 164.32, 159.06, 157.47, 153.67, 152.51, 149.09, 141.60, 140.11, 138.09, 136.09, 133.21, 132.28, 131.40, 131.23, 130.22, 128.64, 125.80, 125.31, 124.65, 123.79, 120.31, 119.82, 113.97, 103.95, 47.12, 28.72, 19.62, 17.25. LC-MS (ESI, *m/z*): 592.1849 [M + H]<sup>+</sup>.

Etherification method of **11i**, **11k**, and **11m**: to a solution of compound **10** (0.88 mmol, 1.0 equiv) in anhydrous dioxane (5 mL) was added ROH (8.8 mmol, 10.0 equiv) and K<sub>2</sub>CO<sub>3</sub> (8.8 mmol, 10.0 equiv) at room temperature under argon. The reaction mixture was stirred at room temperature for 24 h. Then, the resulting mixture was

concentrated to dryness. The residue was diluted with water and extracted with EtOAc. The organic layers were washed with water and brine, and dried. Evaporation of the solvent afforded the crude product **11**, which was purified by flash chromatography.

**N-(4-Methyl-3-(1-methyl-7-(4-methyl-3-nitrophenylamino)-2-oxo-1,2-dihydropyrimido[4,5-*d*]pyrimidin-3(4H)-yl)phenyl)biphenyl-3-carboxamide (11b).** (Method E) Yield 79%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.41 (s, 1H), 9.99 (s, 1H), 8.77 (s, 1H), 8.25 (s, 1H), 8.21 (s, 1H), 7.96 (d, *J* = 7.7 Hz, 1H), 7.87 (dd, *J* = 15.4, 9.7 Hz, 3H), 7.78 (d, *J* = 7.6 Hz, 2H), 7.69 (d, *J* = 8.3 Hz, 1H), 7.63 (t, *J* = 7.7 Hz, 1H), 7.52 (t, *J* = 7.5 Hz, 2H), 7.43 (d, *J* = 7.3 Hz, 1H), 7.39 (d, *J* = 8.6 Hz, 1H), 7.32 (d, *J* = 8.4 Hz, 1H), 4.74 (d, *J* = 14.1 Hz, 1H), 4.56 (d, *J* = 14.2 Hz, 1H), 3.39 (d, *J* = 5.2 Hz, 3H), 2.47 (s, 3H), 2.16 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 165.75, 159.07, 157.47, 153.68, 152.52, 149.11, 141.59, 140.81, 140.13, 139.99, 138.47, 135.93, 133.21, 131.18, 130.26, 129.62, 129.48, 128.31, 127.39, 127.30, 126.23, 125.31, 123.81, 120.23, 119.70, 114.00, 103.95, 47.16, 28.73, 19.61, 17.25. LC-MS (ESI, *m/z*): 600.2690 [M + H]<sup>+</sup>.

**3,5-Dimethyl-N-(4-methyl-3-(1-methyl-7-(4-methyl-3-nitrophenylamino)-2-oxo-1,2-dihydropyrimido[4,5-*d*]pyrimidin-3(4H)-yl)phenyl)benzamide (11c).** (Method E) Yield 82%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.21 (s, 1H), 9.99 (s, 1H), 8.77 (s, 1H), 8.21 (s, 1H), 7.84 (s, 2H), 7.65 (d, *J* = 6.1 Hz, 1H), 7.58 (s, 2H), 7.40 (s, 1H), 7.29 (d, *J* = 7.2 Hz, 1H), 7.22 (s, 1H), 4.73 (d, *J* = 13.5 Hz, 1H), 4.55 (d, *J* = 13.7 Hz, 1H), 3.39 (s, 3H), 2.47 (s, 3H), 2.36 (s, 6H), 2.15 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 166.10, 159.06, 157.47, 153.67, 152.50, 149.12, 141.54, 140.13, 138.60, 138.02, 135.32, 133.32, 133.22, 131.12, 130.84, 125.78, 125.33, 123.80, 120.05, 119.47, 113.99, 103.95, 47.15, 28.72, 21.33, 19.60, 17.23. LC-MS (ESI, *m/z*): 552.2290 [M + H]<sup>+</sup>.

**3,5-Di-*tert*-butyl-N-(4-methyl-3-(1-methyl-7-(4-methyl-3-nitrophenylamino)-2-oxo-1,2-dihydropyrimido[4,5-*d*]pyrimidin-3(4H)-yl)phenyl)benzamide (11d).** (Method E) Yield 73%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.23 (s, 1H), 9.99 (s, 1H), 8.78 (s, 1H), 8.22 (s, 1H), 7.85 (d, *J* = 8.0 Hz, 1H), 7.78 (s, 3H), 7.68 (d, *J* = 7.8 Hz, 1H), 7.62 (s, 1H), 7.40 (d, *J* = 8.1 Hz, 1H), 7.31 (d, *J* = 8.0 Hz, 1H), 4.74 (d, *J* = 14.0 Hz, 1H), 4.57 (d, *J* = 14.0 Hz, 1H), 3.40 (s, 3H), 2.47 (s, 3H), 2.16 (s, 3H), 1.35 (s, 18H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 166.59, 159.07, 157.49, 153.67, 152.53, 151.00, 149.12, 141.53, 140.13, 138.48, 134.80, 133.21, 131.09, 130.94, 125.30, 123.80, 122.13, 120.51, 120.03, 113.99, 103.96, 99.99, 49.24, 46.98, 34.81, 31.74, 28.68, 19.59, 17.08. LC-MS (ESI, *m/z*): 636.3230 [M + H]<sup>+</sup>.

**3-Methyl-N-(4-methyl-3-(1-methyl-7-(4-methyl-3-nitrophenylamino)-2-oxo-1,2-dihydropyrimido[4,5-*d*]pyrimidin-3(4H)-yl)phenyl)benzamide (11e).** (Method E) Yield 79%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.27 (s, 1H), 9.99 (s, 1H), 8.77 (s, 1H), 8.21 (s, 1H), 7.85 (s, 2H), 7.79 (s, 2H), 7.69–7.61 (m, 1H), 7.41 (d, *J* = 3.1 Hz, 3H), 7.30 (d, *J* = 7.7 Hz, 1H), 4.73 (d, *J* = 14.0 Hz, 1H), 4.55 (d, *J* = 14.0 Hz, 1H), 3.39 (s, 3H), 2.47 (s, 3H), 2.41 (s, 3H), 2.15 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 165.99, 159.07, 157.48, 153.67, 152.50, 149.12, 141.55, 140.13, 138.55, 138.17, 135.30, 133.21, 132.61, 131.14, 130.88, 128.76, 128.53, 125.25, 123.81, 120.09, 119.52, 113.99, 103.96, 55.36, 28.72, 21.43, 19.60, 17.24. LC-MS (ESI, *m/z*): 538.2125 [M + H]<sup>+</sup>.

**N-(3-(7-(3,4-Dimethylphenylamino)-1-methyl-2-oxo-1,2-dihydropyrimido[4,5-*d*]pyrimidin-3(4H)-yl)-4-methylphenyl)-3-(trifluoromethyl)benzamide (11f).** (Method E) Yield 88%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.56 (d, *J* = 12.6 Hz, 1H), 9.40 (s, 1H), 8.33 (s, 1H), 8.29 (d, *J* = 7.4 Hz, 1H), 8.13 (s, 1H), 7.97 (d, *J* = 7.0 Hz, 1H), 7.86–7.76 (m, 2H), 7.67 (d, *J* = 6.7 Hz, 1H), 7.58 (s, 1H), 7.50 (d, *J* = 7.7 Hz, 1H), 7.32 (d, *J* = 7.5 Hz, 1H), 7.04 (d, *J* = 7.7 Hz, 1H), 4.70 (d, *J* = 13.7 Hz, 1H), 4.52 (d, *J* = 14.0 Hz, 1H), 3.65 (s, 3H), 3.38 (s, 3H), 2.21 (s, 3H), 2.17 (s, 6H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 164.32, 159.61, 157.32, 153.67, 153.01, 152.71, 141.72, 138.72, 138.09, 136.31, 136.10, 132.27, 131.42, 131.31, 130.21, 129.84, 129.54, 129.38, 128.62, 124.66, 120.75, 119.85, 116.97, 102.63, 72.56, 28.24, 20.16, 19.12, 17.49. LC-MS (ESI, *m/z*): 561.2153 [M + H]<sup>+</sup>.

**N-(4-Methyl-3-(1-methyl-2-oxo-7-(*p*-tolylamino)-1,2-dihydropyrimido[4,5-*d*]pyrimidin-3(4H)-yl)phenyl)-3-(trifluoromethyl)benzamide (11g).** (Method E) Yield 89%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.55 (s, 1H), 9.48 (s, 1H), 8.33 (s, 1H),

8.29 (d,  $J = 7.3$  Hz, 1H), 8.14 (s, 1H), 7.97 (d,  $J = 7.1$  Hz, 1H), 7.85–7.77 (m, 2H), 7.67 (d,  $J = 7.6$  Hz, 2H), 7.32 (d,  $J = 7.6$  Hz, 1H), 7.10 (d,  $J = 7.3$  Hz, 2H), 4.70 (d,  $J = 13.7$  Hz, 1H), 4.52 (d,  $J = 13.8$  Hz, 1H), 3.65 (s, 3H), 2.26 (s, 3H), 2.16 (s, 3H).  $^{13}\text{C}$  NMR (101 MHz, DMSO- $d_6$ )  $\delta$  164.33, 159.55, 157.38, 153.60, 152.69, 141.70, 138.46, 138.09, 136.10, 132.27, 131.42, 131.21, 130.59, 130.21, 129.85, 129.65, 129.54, 129.34, 128.63, 124.66, 120.28, 119.85, 119.58, 119.47, 102.77, 72.14, 28.81, 21.02, 17.25. LC-MS (ESI,  $m/z$ ): 547.1200  $[\text{M} + \text{H}]^+$ .

*N*-(4-Methyl-3-(1-methyl-7-(4-methyl-3-(trifluoromethyl)phenylamino)-2-oxo-1,2-dihydropyrimido[4,5-*d*]pyrimidin-3(4*H*)-yl)phenyl)-3-(trifluoromethyl)benzamide (**11h**). (Method E) Yield 78%.  $^1\text{H}$  NMR (101 MHz, DMSO- $d_6$ )  $\delta$  164.33, 159.23, 157.43, 153.73, 152.55, 141.63, 139.33, 138.08, 136.09, 132.86, 132.29, 131.41, 131.23, 130.25, 129.82, 129.51, 128.64, 128.27, 127.91, 127.62, 126.53, 125.81, 124.62, 123.81, 123.10, 122.39, 120.30, 119.82, 115.93, 115.87, 103.59, 47.12, 28.60, 18.54, 17.26.  $^{13}\text{C}$  NMR (101 MHz, DMSO- $d_6$ )  $\delta$  164.33, 159.23, 157.43, 153.73, 152.55, 141.63, 139.33, 138.08, 136.09, 132.86, 132.29, 131.41, 131.23, 130.25, 129.82, 129.51, 128.64. LC-MS (ESI,  $m/z$ ): 615.1901  $[\text{M} + \text{H}]^+$ .

*N*-(4-Methyl-3-(1-methyl-7-(4-methyl-3-nitrophenoxy)-2-oxo-1,2-dihydropyrimido[4,5-*d*]pyrimidin-3(4*H*)-yl)phenyl)-3-(trifluoromethyl)benzamide (**11i**). (Etherification Method) Yield 90%.  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  9.15 (s, 1H), 8.16 (s, 1H), 8.13 (d,  $J = 8.0$  Hz, 1H), 8.03 (s, 1H), 7.97 (s, 1H), 7.80–7.74 (m, 2H), 7.61 (d,  $J = 7.8$  Hz, 1H), 7.44 (d,  $J = 1.4$  Hz, 2H), 7.23 (dd,  $J = 8.3, 2.0$  Hz, 1H), 7.03 (d,  $J = 8.4$  Hz, 1H), 4.72 (d,  $J = 15.2$  Hz, 1H), 4.50 (d,  $J = 15.0$  Hz, 1H), 3.48 (s, 3H), 2.66 (s, 3H), 1.66 (s, 3H).  $^{13}\text{C}$  NMR (101 MHz,  $\text{CDCl}_3$ )  $\delta$  164.43, 164.01, 159.24, 155.18, 153.74, 153.09, 150.91, 149.14, 139.55, 137.59, 135.37, 133.65, 133.48, 131.44, 131.11, 130.85, 129.16, 128.24, 126.66, 124.64, 124.35, 120.99, 120.07, 118.39, 111.24, 107.00, 46.97, 28.75, 20.26, 16.23. LC-MS (ESI,  $m/z$ ): 593.1680  $[\text{M} + \text{H}]^+$ .

*N*-(4-Methyl-3-(1-methyl-7-(3-morpholinopropylamino)-2-oxo-1,2-dihydropyrimido[4,5-*d*]pyrimidin-3(4*H*)-yl)phenyl)-3-(trifluoromethyl)benzamide (**11j**). (Method E) Yield 92%.  $^1\text{H}$  NMR (400 MHz, DMSO- $d_6$ )  $\delta$  10.62 (s, –1H), 8.33 (s, 1H), 8.29 (d,  $J = 7.4$  Hz, 1H), 7.95 (d,  $J = 10.8$  Hz, 2H), 7.78 (dd,  $J = 15.1, 7.1$  Hz, 2H), 7.67 (d,  $J = 8.1$  Hz, 1H), 7.29 (d,  $J = 8.1$  Hz, 1H), 7.16 (s, 1H), 4.62 (d,  $J = 13.6$  Hz, 1H), 4.42 (d,  $J = 13.6$  Hz, 1H), 3.62 (s, 6H), 3.34 (d,  $J = 5.6$  Hz, 2H), 3.19 (s, 3H), 2.48 (s, 6H), 2.13 (s, 3H), 1.75 (d,  $J = 5.6$  Hz, 2H).  $^{13}\text{C}$  NMR (101 MHz, DMSO- $d_6$ )  $\delta$  164.31, 162.05, 158.74, 157.32, 152.90, 141.80, 138.11, 136.08, 132.21, 131.34, 131.11, 130.12, 129.88, 129.56, 128.55, 125.78, 124.67, 123.07, 120.16, 119.77, 56.35, 53.83, 53.48, 49.02, 47.14, 42.13, 28.17, 25.96, 17.18, 12.74. LC-MS (ESI,  $m/z$ ): 584.2524  $[\text{M} + \text{H}]^+$ .

(*S*)-*tert*-Butyl 3-(8-methyl-6-(2-methyl-5-(3-(trifluoromethyl)benzamido)phenyl)-7-oxo-5,6,7,8-tetrahydropyrimido[4,5-*d*]pyrimidin-2-yloxy)piperidine-1-carboxylate (**11k**). (Etherification Method) Yield 90%.  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  9.24 (s, 1H), 8.17 (s, 1H), 8.13 (d,  $J = 7.8$  Hz, 1H), 8.01 (s, 1H), 7.75 (d,  $J = 6.9$  Hz, 1H), 7.57 (t,  $J = 7.8$  Hz, 1H), 7.27 (d,  $J = 10.4$  Hz, 1H), 7.02–7.00 (m, 1H), 4.68 (dd,  $J = 14.3, 3.5$  Hz, 1H), 4.45 (d,  $J = 14.3$  Hz, 1H), 4.27 (dt,  $J = 11.7, 6.4$  Hz, 1H), 3.72 (s, 2H), 3.66–3.57 (m, 2H), 3.52 (d,  $J = 6.6$  Hz, 3H), 2.27 (t,  $J = 16.8$  Hz, 2H), 1.64 (d,  $J = 6.3$  Hz, 3H), 1.49 (s, 9H).  $^{13}\text{C}$  NMR (101 MHz,  $\text{CDCl}_3$ )  $\delta$  162.31, 162.06, 151.57, 151.29, 137.66, 135.76, 133.66, 129.51, 129.05, 128.57, 126.97, 125.97, 123.19, 122.51, 120.55, 118.87, 118.06, 103.39, 77.36, 74.17, 66.23, 63.49, 44.96, 36.45, 27.76, 26.37, 14.31. LC-MS (ESI,  $m/z$ ): 627.2470  $[\text{M} + \text{H}]^+$ .

*tert*-Butyl 4-(8-methyl-6-(2-methyl-5-(3-(trifluoromethyl)benzamido)phenyl)-7-oxo-5,6,7,8-tetrahydropyrimido[4,5-*d*]pyrimidin-2-ylamino)piperidine-1-carboxylate (**11l**). (Method E) Yield 79%.  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  9.27 (s, 1H), 8.19 (s, 1H), 8.13 (d,  $J = 7.8$  Hz, 1H), 7.83 (s, 1H), 7.74 (d,  $J = 7.7$  Hz, 1H), 7.65 (s, 1H), 7.57 (t,  $J = 7.8$  Hz, 1H), 7.32 (d,  $J = 8.3$  Hz, 1H), 6.99 (d,  $J = 8.4$  Hz, 1H), 4.58 (d,  $J = 13.9$  Hz, 1H), 4.34 (d,  $J = 14.0$  Hz, 1H), 4.22–3.84 (m, 4H), 3.47 (s, 3H), 3.00 (t,  $J = 11.5$  Hz, 2H), 2.12–2.04 (m, 3H), 1.63 (s, 3H), 1.49 (s, 9H).  $^{13}\text{C}$  NMR (101 MHz,  $\text{CDCl}_3$ )  $\delta$  164.25, 161.02, 157.39, 154.82, 153.81, 152.98, 140.07, 137.71, 135.70, 131.46, 130.88, 130.73, 128.93, 124.46, 120.67, 120.12, 79.67, 48.42,

47.29, 32.12, 28.45, 28.19, 16.20. LC-MS (ESI,  $m/z$ ): 640.2490  $[\text{M} + \text{H}]^+$ .

*tert*-Butyl 4-(8-methyl-6-(2-methyl-5-(3-(trifluoromethyl)benzamido)phenyl)-7-oxo-5,6,7,8-tetrahydropyrimido[4,5-*d*]pyrimidin-2-yloxy)piperidine-1-carboxylate (**11m**). (Etherification Method) Yield 50%.  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  9.24 (s, 1H), 8.17 (s, 1H), 8.13 (d,  $J = 7.9$  Hz, 1H), 8.00 (s, 1H), 7.75 (d,  $J = 7.2$  Hz, 2H), 7.57 (t,  $J = 7.8$  Hz, 1H), 7.25 (dd,  $J = 8.3, 1.7$  Hz, 1H), 7.00 (d,  $J = 8.4$  Hz, 1H), 4.67 (d,  $J = 14.5$  Hz, 1H), 4.44 (d,  $J = 14.5$  Hz, 1H), 3.92–3.83 (m, 2H), 3.53 (s, 3H), 3.33–3.23 (m, 2H), 2.12–2.04 (m, 2H), 1.90–1.82 (m, 2H), 1.62 (s, 3H), 1.50 (s, 9H).  $^{13}\text{C}$  NMR (101 MHz,  $\text{CDCl}_3$ )  $\delta$  164.22, 164.01, 158.81, 154.87, 153.65, 153.39, 139.76, 137.78, 135.59, 131.47, 130.93, 130.63, 128.99, 127.99, 124.37, 120.79, 120.04, 105.00, 99.99, 79.56, 72.79, 46.97, 29.71, 28.60, 28.34, 16.18. LC-MS (ESI,  $m/z$ ): 641.2633  $[\text{M} + \text{H}]^+$ .

*N*-(4-Methyl-3-(1-methyl-7-(4-methyl-3-(methylamino)phenylamino)-2-oxo-1,2-dihydropyrimido[4,5-*d*]pyrimidin-3(4*H*)-yl)phenyl)-3-(trifluoromethyl)benzamide (**11n**). (Method E) Yield 79%.  $^1\text{H}$  NMR (400 MHz, DMSO- $d_6$ )  $\delta$  10.54 (s, 1H), 9.26 (s, 1H), 8.32 (s, 1H), 8.28 (d,  $J = 7.7$  Hz, 1H), 8.13 (s, 1H), 7.98 (d,  $J = 7.5$  Hz, 1H), 7.80 (t,  $J = 7.6$  Hz, 2H), 7.67 (d,  $J = 8.1$  Hz, 1H), 7.33 (d,  $J = 8.3$  Hz, 1H), 7.11 (s, 1H), 6.94–6.83 (m, 2H), 4.70 (d,  $J = 13.7$  Hz, 1H), 4.51 (d,  $J = 13.8$  Hz, 1H), 3.38 (s, 3H), 2.75 (d,  $J = 4.6$  Hz, 3H), 2.15 (s, 3H), 2.03 (s, 3H).  $^{13}\text{C}$  NMR (101 MHz, DMSO- $d_6$ )  $\delta$  164.34, 159.70, 157.29, 153.75, 152.74, 148.10, 141.74, 139.94, 138.06, 136.12, 132.29, 131.44, 131.22, 130.24, 129.71, 128.64, 124.62, 120.27, 119.88, 115.47, 111.29, 106.89, 102.28, 100.67, 47.19, 30.71, 28.70, 17.44, 17.26. LC-MS (ESI,  $m/z$ ): 576.2260  $[\text{M} + \text{H}]^+$ .

*N*-(3-(7-(3-(Dimethylamino)-4-methylphenylamino)-1-methyl-2-oxo-1,2-dihydropyrimido [4,5-*d*]pyrimidin-3(4*H*)-yl)-4-methylphenyl)-3-(trifluoromethyl)benzamide (**11o**). (Method E) Yield 65%.  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  9.21 (s, 1H), 8.21 (s, 1H), 8.15 (d,  $J = 7.6$  Hz, 1H), 7.96 (s, 1H), 7.77 (d,  $J = 7.5$  Hz, 1H), 7.68 (s, 1H), 7.59 (t,  $J = 7.7$  Hz, 1H), 7.44 (s, 1H), 7.36 (d,  $J = 8.1$  Hz, 1H), 7.29 (s, 1H), 7.16 (s, 2H), 7.03 (d,  $J = 8.1$  Hz, 1H), 4.65 (d,  $J = 13.9$  Hz, 1H), 4.41 (d,  $J = 14.0$  Hz, 1H), 3.58 (s, 3H), 2.77 (s, 6H), 2.34 (s, 3H), 1.67 (s, 3H).  $^{13}\text{C}$  NMR (101 MHz,  $\text{CDCl}_3$ )  $\delta$  164.25, 159.53, 157.30, 153.07, 137.70, 135.69, 131.33, 131.05, 129.03, 128.01, 126.47, 124.39, 120.71, 120.06, 114.84, 113.79, 110.27, 101.95, 101.61, 96.94, 47.38, 44.23, 29.71, 28.71, 17.93. LC-MS (ESI,  $m/z$ ): 590.2421  $[\text{M} + \text{H}]^+$ .

**General Method F.** *N*-(3-(7-(3-Amino-4-methylphenylamino)-1-methyl-2-oxo-1,2-dihydropyrimido[4,5-*d*]pyrimidin-3(4*H*)-yl)-4-methylphenyl)-3-(trifluoromethyl)benzamide (**12a**). To a solution of *N*-(4-methyl-3-(1-methyl-7-(4-methyl-3-nitrophenylamino)-2-oxo-1,2-dihydropyrimido[4,5-*d*]pyrimidin-3(4*H*)-yl)phenyl)-3-(trifluoromethyl)benzamide (50 mg, 0.085 mmol, 1.00 equiv) in methanol (5 mL) was added  $\text{SnCl}_4 \cdot 2\text{H}_2\text{O}$  (191 mg, 0.85 mmol, 10.0 equiv) at room temperature under argon. The reaction mixture was then heated to reflux for 14 h. The resulting mixture was then concentrated to dryness. The residue was diluted with water (50 mL) and added to 1 N NaOH to pH about 10. The mixture was extracted with EtOAc ( $3 \times 30$  mL). The combined organic layers were washed with water (50 mL) and brine (50 mL), and dried over anhydrous  $\text{MgSO}_4$ . Evaporation of the solvent afforded the crude product, which was purified by flash column chromatography (eluting with MeOH in DCM 0–5%) to yield **12a** as a white solid (40 mg, 80%).  $^1\text{H}$  NMR (400 MHz, DMSO- $d_6$ )  $\delta$  10.91 (s, 1H), 10.72 (s, 1H), 8.34 (s, 2H), 8.32 (s, 1H), 8.27 (s, 1H), 7.97 (d,  $J = 7.5$  Hz, 1H), 7.91 (s, 1H), 7.86 (s, 1H), 7.79 (t,  $J = 7.6$  Hz, 1H), 7.69 (d,  $J = 8.1$  Hz, 1H), 7.57 (d,  $J = 7.9$  Hz, 1H), 7.33 (dd,  $J = 8.1, 3.8$  Hz, 2H), 4.76 (d,  $J = 14.4$  Hz, 1H), 4.60 (d,  $J = 14.5$  Hz, 1H), 2.37 (s, 3H), 2.15 (s, 3H).  $^{13}\text{C}$  NMR (101 MHz, DMSO- $d_6$ )  $\delta$  164.37, 159.62, 151.49, 141.00, 138.19, 137.11, 136.00, 132.37, 132.01, 131.35, 131.28, 130.21, 129.79, 129.47, 128.64, 127.64, 125.80, 124.76, 124.73, 123.09, 120.77, 120.64, 119.71, 116.23, 104.28, 46.54, 29.59, 17.22, 17.17. LC-MS (ESI,  $m/z$ ): 562.1109  $[\text{M} + \text{H}]^+$ .

**N-Boc deprotection method of 12f and 12h:** To a solution of compound **11** (0.085 mmol, 1.0 equiv) in anhydrous dioxane (1 mL) was added 1 mL of 4 M HCl in dioxane at room temperature. The reaction mixture was stirred at room temperature for 30 min. The resulting mixture was concentrated to afford the title compound **12**.

*N*-(3-(7-(3-Amino-4-methylphenylamino)-1-methyl-2-oxo-1,2-dihydropyrimido[4,5-*d*] pyrimidin-3(4*H*)-yl)-4-methylphenyl)-biphenyl-3-carboxamide (**12b**). (Method F) Yield 85%. <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>OD) δ 8.17 (s, 1H), 7.96 (s, 1H), 7.90 (d, *J* = 7.7 Hz, 1H), 7.83–7.77 (m, 2H), 7.71–7.66 (m, 2H), 7.57 (t, *J* = 7.6 Hz, 2H), 7.46 (t, *J* = 7.5 Hz, 2H), 7.38 (d, *J* = 7.4 Hz, 1H), 7.29 (d, *J* = 8.3 Hz, 1H), 7.15 (s, 1H), 7.01 (q, *J* = 8.1 Hz, 2H), 4.51 (d, *J* = 14.1 Hz, 1H), 3.44 (s, 3H), 2.21 (s, 3H), 2.17 (s, 3H). <sup>13</sup>C NMR (101 MHz, CD<sub>3</sub>OD) δ 167.37, 158.81, 157.70, 153.39, 151.66, 141.58, 140.65, 140.12, 138.14, 137.64, 135.30, 131.60, 131.12, 130.35, 130.16, 128.86, 128.70, 127.56, 126.87, 126.11, 126.02, 122.56, 120.72, 119.67, 118.40, 111.61, 102.16, 48.90, 47.08, 28.01, 16.43. LC-MS (ESI, *m/z*): 570.2540 [M + H]<sup>+</sup>.

*N*-(3-(7-(3-Amino-4-methylphenylamino)-1-methyl-2-oxo-1,2-dihydropyrimido[4,5-*d*] pyrimidin-3(4*H*)-yl)-4-methylphenyl)-3,5-dimethylbenzamide (**12c**). (Method F) Yield 85%. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.43 (s, 1H), 7.91 (s, 1H), 7.73 (s, 1H), 7.49 (s, 2H), 7.45 (s, 1H), 7.35 (d, *J* = 8.2 Hz, 1H), 7.14 (d, *J* = 7.6 Hz, 2H), 7.08 (s, 1H), 7.02 (d, *J* = 8.1 Hz, 1H), 6.94 (d, *J* = 8.1 Hz, 1H), 4.65 (d, *J* = 14.0 Hz, 1H), 4.40 (d, *J* = 14.1 Hz, 1H), 3.49 (s, 3H), 2.37 (s, 6H), 2.16 (s, 3H), 2.01 (s, 3H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 166.11, 159.29, 157.47, 153.10, 152.45, 144.93, 140.58, 138.22, 137.48, 134.69, 133.31, 131.21, 131.12, 130.65, 125.08, 120.22, 119.44, 116.98, 109.97, 106.26, 102.09, 47.17, 29.88, 28.67, 21.01, 16.65. LC-MS (ESI, *m/z*): 522.2546 [M + H]<sup>+</sup>.

*N*-(3-(7-(3-Amino-4-methylphenylamino)-1-methyl-2-oxo-1,2-dihydropyrimido[4,5-*d*] pyrimidin-3(4*H*)-yl)-4-methylphenyl)-3,5-ditert-butylbenzamide (**12d**). (Method F) Yield 75%. <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>OD) δ 7.91 (s, 1H), 7.80 (s, 2H), 7.72 (s, 1H), 7.67 (s, 1H), 7.63 (d, *J* = 6.8 Hz, 1H), 7.22 (d, *J* = 8.3 Hz, 1H), 7.11 (s, 1H), 6.91 (s, 2H), 4.60 (d, *J* = 14.0 Hz, 1H), 4.40 (d, *J* = 14.0 Hz, 1H), 3.38 (d, *J* = 5.9 Hz, 3H), 2.11 (s, 6H), 1.37 (s, 18H). <sup>13</sup>C NMR (101 MHz, CD<sub>3</sub>OD) δ 168.26, 159.44, 157.14, 153.52, 152.68, 151.09, 145.12, 140.82, 138.54, 137.79, 134.29, 131.43, 130.86, 129.88, 125.72, 121.59, 120.59, 119.83, 117.04, 110.03, 106.83, 101.91, 99.99, 34.56, 30.48, 27.78, 15.87, 15.62. LC-MS (ESI, *m/z*): 606.3485 [M + H]<sup>+</sup>.

*N*-(3-(7-(3-Amino-4-methylphenylamino)-1-methyl-2-oxo-1,2-dihydropyrimido[4,5-*d*] pyrimidin-3(4*H*)-yl)-4-methylphenyl)-3-methylbenzamide (**12e**). (Method F) Yield 78%. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.73 (s, 1H), 7.69 (s, 2H), 7.52 (d, *J* = 8.1 Hz, 1H), 7.42–7.35 (m, 3H), 7.28 (d, *J* = 8.2 Hz, 1H), 7.14–6.90 (m, 3H), 4.75 (d, *J* = 13.9 Hz, 1H), 4.49 (d, *J* = 13.5 Hz, 1H), 3.46 (s, 3H), 2.44 (s, 3H), 2.21 (s, 3H), 2.18 (s, 3H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 170.83, 161.72, 157.13, 144.44, 142.29, 141.24, 138.56, 136.41, 135.38, 135.25, 134.46, 132.32, 131.83, 128.30, 128.25, 124.73, 123.53, 114.91, 100.04, 51.12, 33.38, 32.49, 25.04, 20.79. LC-MS (ESI, *m/z*): 508.2390 [M + H]<sup>+</sup>.

(*S*)-*N*-(4-Methyl-3-(1-methyl-2-oxo-7-(pyrrolidin-3-yloxy)-1,2-dihydropyrimido[4,5-*d*] pyrimidin-3(4*H*)-yl)phenyl)-3-(trifluoromethyl)benzamide (**12f**). (Deprotection Method) Yield 95%. <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>OD) δ 8.31 (s, 1H), 8.27 (s, 1H), 8.23 (d, *J* = 7.8 Hz, 1H), 7.96–7.88 (m, 2H), 7.74 (dd, *J* = 10.2, 5.1 Hz, 1H), 7.64–7.59 (m, 1H), 7.35 (d, *J* = 8.3 Hz, 1H), 4.93–4.91 (m, 1H), 4.74 (d, *J* = 14.7 Hz, 1H), 3.80 (s, 2H), 3.59 (d, *J* = 5.6 Hz, 1H), 3.52 (s, 3H), 2.54 (s, 2H), 2.24 (s, 3H), 2.06–2.00 (m, 2H). <sup>13</sup>C NMR (101 MHz, CD<sub>3</sub>OD) δ 165.47, 161.47, 158.68, 150.95, 144.49, 140.02, 137.64, 135.66, 131.80, 131.14, 130.97, 129.34, 128.48, 128.03, 124.22, 120.93, 119.48, 107.32, 78.97, 50.38, 46.15, 43.90, 30.30, 28.67, 15.64. LC-MS (ESI, *m/z*): 527.1950 [M + H]<sup>+</sup>.

*N*-(4-Methyl-3-(1-methyl-2-oxo-7-(piperidin-4-ylamino)-1,2-dihydropyrimido[4,5-*d*] pyrimidin-3(4*H*)-yl)phenyl)-3-(trifluoromethyl)benzamide (**12g**). (Deprotection Method) Yield 96%. <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>OD) δ 8.28 (m, 2H), 7.83 (m, 3H), 7.67 (s, 1H), 7.56 (s, 1H), 7.24 (s, 1H), 4.53 (m, 2H), 3.44 (s, 5H), 3.23 (br, 3H), 2.25 (br, 2H), 1.94 (m, 5H). <sup>13</sup>C NMR (101 MHz, CD<sub>3</sub>OD) δ 173.79, 171.60, 165.31, 152.87, 151.31, 140.10, 137.56, 135.53, 131.85, 131.14, 131.08, 130.36, 129.43, 128.08, 125.27, 124.25, 122.63, 120.94, 119.64, 46.65, 46.03, 42.64, 27.58, 19.34, 15.86. LC-MS (ESI, *m/z*): 540.2260 [M + H]<sup>+</sup>.

*N*-(4-Methyl-3-(1-methyl-2-oxo-7-(piperidin-4-yloxy)-1,2-dihydropyrimido[4,5-*d*]pyrimidin-3(4*H*)-yl)phenyl)-3-

(trifluoromethyl)benzamide (**12h**). (Deprotection Method) Yield 95%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.67 (s, 1H), 9.29 (s, 1H), 8.33 (s, 1H), 8.26 (s, 1H), 7.97 (d, *J* = 6.1 Hz, 1H), 7.86 (s, 1H), 7.79 (s, 1H), 7.68 (d, *J* = 6.4 Hz, 1H), 7.32 (d, *J* = 7.2 Hz, 1H), 4.76 (d, *J* = 13.6 Hz, 1H), 4.60 (d, *J* = 14.0 Hz, 1H), 3.31 (s, 3H), 3.23 (s, 2H), 3.13 (s, 2H), 2.20 (s, 2H), 2.13 (s, 3H), 2.01 (s, 2H). <sup>13</sup>C NMR (101 MHz, CD<sub>3</sub>OD) δ 163.77, 160.03, 158.58, 150.71, 148.91, 142.69, 139.21, 136.22, 134.31, 130.41, 129.71, 127.99, 126.66, 122.81, 119.38, 118.27, 99.97, 68.49, 45.19, 39.39, 26.79, 25.30, 14.35. LC-MS (ESI, *m/z*): 541.2301 [M + H]<sup>+</sup>.

*N*-(3-(7-(3-Amino-4-methylphenoxy)-1-methyl-2-oxo-1,2-dihydropyrimido[4,5-*d*]pyrimidin-3(4*H*)-yl)-4-methylphenyl)-3-(trifluoromethyl)benzamide (**12i**). (Method F) Yield 79%. <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>OD) δ 8.42 (s, 1H), 8.26 (s, 1H), 8.23 (d, *J* = 7.4 Hz, 1H), 7.91 (d, *J* = 6.8 Hz, 2H), 7.75 (t, *J* = 7.4 Hz, 1H), 7.61 (d, *J* = 8.2 Hz, 3H), 7.47 (s, 1H), 7.36 (d, *J* = 8.0 Hz, 1H), 7.18 (s, 1H), 4.80 (s, 2H), 3.37 (s, 3H), 2.50 (s, 3H), 2.26 (s, 3H). <sup>13</sup>C NMR (101 MHz, CD<sub>3</sub>OD) δ 165.55, 161.53, 150.95, 150.03, 139.99, 137.62, 135.71, 132.76, 132.17, 131.84, 131.15, 130.79, 130.71, 130.19, 129.33, 128.01, 124.21, 122.29, 121.58, 120.95, 119.56, 116.96, 109.88, 108.37, 48.48, 46.51, 28.44, 15.41. LC-MS (ESI, *m/z*): 563.1900 [M + H]<sup>+</sup>.

**General Method G. 4-(Methylamino)-2-(methylthio)pyrimidine-5-carboxylic Acid (**14a**)**. To a solution of ethyl 4-(methylamino)-2-(methylthio)pyrimidine-5-carboxylate (0.976g, 4.3 mmol, 1.0 equiv) in methanol (10 mL) and water (2 mL) was added NaOH (0.18 g, 4.4 mmol, 1.02 equiv) at room temperature under argon. The reaction mixture was stirred at room temperature for 20 h. The resulting mixture was concentrated to dryness. The residue was diluted with water (50 mL), acidified by 1 N HCl to pH 3. The white precipitate was filtered and dried to provide **14a** as a white solid (0.70 g, 80%). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 8.48 (s, 1H), 8.35 (s, 1H), 2.96 (d, *J* = 3.9 Hz, 3H), 2.47 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 175.07, 168.31, 160.62, 158.32, 101.76, 27.59, 14.00. TOF LCMS (*m/z*): 200.0423 [M + H]<sup>+</sup>.

**4-(Dimethylamino)-2-(methylthio)pyrimidine-5-carboxylic Acid (**14b**)**. (Method G) Yield 85%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 8.44 (s, 1H), 3.15 (s, 6H), 2.57 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 166.66, 159.23, 154.08, 137.14, 106.24, 40.50, 14.00. LC-MS (ESI, *m/z*): 214.0585 [M + H]<sup>+</sup>.

**General Method H. *N*-(2-Methyl-5-(3-(trifluoromethyl)benzamido)phenyl)-4-(methylamino)-2-(methylthio)pyrimidine-5-carboxamide (**15a**)**. To a solution of the 4-(methylamino)-2-(methylthio)pyrimidine-5-carboxylic acid (0.6 g, 3.0 mmol, 1.00 equiv) in anhydrous DMF (5 mL) was added *N*-(3-amino-4-methylphenyl)-3-(trifluoromethyl)benzamide (1.06 g, 3.6 mmol, 1.20 equiv), HATU (1.36 g, 3.6 mmol, 1.2 equiv), and DIPEA (1.6 mL, 6.0 mmol, 2.0 equiv) at room temperature under argon. The reaction mixture was stirred at room temperature for 20 h. The resulting mixture was concentrated to dryness. The residue was diluted with water (100 mL) and extracted with EtOAc (3 × 50 mL). The combined organic layers were washed with water (50 mL) and brine (50 mL), and dried over anhydrous MgSO<sub>4</sub>. Evaporation of the solvent afforded the crude product, which was purified by flash column chromatography (eluting with MeOH in DCM 0–5%) to yield **15a** as a white solid (1.14g, 80%). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.51 (s, 1H), 9.56 (s, 1H), 8.83 (s, 1H), 8.41–8.23 (m, 3H), 7.97 (d, *J* = 6.6 Hz, 1H), 7.80 (d, *J* = 6.8 Hz, 1H), 7.64 (d, *J* = 7.0 Hz, 1H), 7.26 (d, *J* = 7.3 Hz, 1H), 4.14 (s, 3H), 2.60 (s, 3H), 2.30 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 164.28, 160.68, 159.45, 137.46, 136.52, 136.19, 132.32, 130.67, 130.12, 129.79, 128.52, 124.69, 117.75, 116.03, 111.46, 55.28, 17.29, 13.99. LC-MS (ESI, *m/z*): 476.1235 [M + H]<sup>+</sup>.

**4-(Dimethylamino)-*N*-(2-methyl-5-(3-(trifluoromethyl)benzamido)phenyl)-2-(methylthio)pyrimidine-5-carboxamide (**15b**)**. (Method H) Yield 75%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.51 (s, 1H), 10.01 (s, 1H), 8.28 (d, *J* = 16.4 Hz, 3H), 7.97 (s, 1H), 7.89 (s, 1H), 7.79 (d, *J* = 6.1 Hz, 1H), 7.64 (s, 1H), 7.26 (s, 1H), 3.12 (s, 6H), 2.50 (d, *J* = 5.9 Hz, 3H), 2.26 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 170.61, 166.09, 164.41, 158.95, 155.95, 137.26, 136.37, 136.17, 132.27, 130.85, 130.17, 129.12, 128.67, 128.55, 124.67, 118.83, 117.98, 111.19, 17.86, 13.92. LC-MS (ESI, *m/z*): 490.1452 [M + H]<sup>+</sup>.

*N*-(2-Methyl-5-(3-(trifluoromethyl)benzamido)phenyl)-2-(methylthio)pyrimidine-5-carboxamide (**15c**). (Method H) Yield 90%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.50 (s, 1H), 9.95 (s, 1H), 8.77 (s, 1H), 8.33 (s, 1H), 8.29 (d, *J* = 7.3 Hz, 1H), 7.98–7.92 (m, 2H), 7.84 (s, 1H), 7.80 (d, *J* = 7.4 Hz, 1H), 7.63 (d, *J* = 7.7 Hz, 1H), 7.28 (d, *J* = 8.1 Hz, 1H), 2.50 (s, 3H), 2.23 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 173.52, 165.66, 164.33, 162.11, 156.16, 137.30, 136.31, 136.19, 132.28, 130.75, 130.14, 129.98, 128.54, 125.80, 124.66, 119.39, 118.95, 104.54, 17.86, 13.84. LC-MS (ESI, *m/z*): 447.1042 [M + H]<sup>+</sup>.

*N*-(2-Methyl-5-(3-(trifluoromethyl)benzamido)phenyl)-4-(methylamino)-2-(methylsulfonyl)pyrimidine-5-carboxamide (**16a**). (Method D) Yield 62%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.50 (s, 1H), 10.26 (s, 1H), 8.91 (s, 1H), 8.82 (s, 1H), 8.32 (s, 1H), 8.28 (d, *J* = 7.4 Hz, 1H), 7.98 (d, *J* = 7.3 Hz, 1H), 7.88 (s, 1H), 7.84–7.77 (m, 1H), 7.61 (d, *J* = 7.4 Hz, 1H), 7.30 (d, *J* = 8.3 Hz, 1H), 3.37 (s, 3H), 3.01 (s, 3H), 2.24 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 166.68, 164.38, 161.39, 155.60, 137.33, 136.18, 135.81, 132.30, 130.85, 130.20, 129.92, 128.60, 124.68, 119.28, 111.37, 39.16, 28.17, 17.87. LC-MS (ESI, *m/z*): 508.1190 [M + H]<sup>+</sup>.

4-(Dimethylamino)-*N*-(2-methyl-5-(3-(trifluoromethyl)benzamido)phenyl)-2-(methylsulfonyl)pyrimidine-5-carboxamide (**16b**). (Method D) Yield 72%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.53 (s, 1H), 10.23 (s, 1H), 8.56 (s, 1H), 8.30 (d, *J* = 14.1 Hz, 2H), 7.96 (s, 2H), 7.80 (s, 1H), 7.65 (d, *J* = 7.7 Hz, 1H), 7.28 (d, *J* = 6.5 Hz, 1H), 3.22 (s, 6H), 2.52 (s, 3H), 2.28 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 164.91, 164.71, 164.43, 159.16, 156.24, 137.36, 136.18, 135.94, 132.31, 130.97, 130.19, 129.48, 128.63, 128.38, 124.73, 118.87, 117.80, 117.09, 39.23, 17.88. LC-MS (ESI, *m/z*): 522.1355 [M + H]<sup>+</sup>.

*N*-(2-Methyl-5-(3-(trifluoromethyl)benzamido)phenyl)-2-(methylsulfonyl)pyrimidine-5-carboxamide (**16c**). (Method D) Yield 74%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.52 (s, 1H), 10.24 (s, 1H), 8.96 (s, 1H), 8.32 (s, 3H), 7.97 (s, 1H), 7.91 (s, 1H), 7.80 (s, 1H), 7.62 (d, *J* = 6.9 Hz, 1H), 7.30 (d, *J* = 7.3 Hz, 1H), 3.35 (s, 3H), 2.24 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 166.55, 164.40, 163.12, 156.67, 137.33, 136.15, 135.86, 133.82, 133.14, 132.28, 131.07, 130.87, 130.18, 129.91, 129.29, 128.55, 128.36, 125.77, 124.66, 123.10, 119.30, 66.42, 17.84. LC-MS (ESI, *m/z*): 479.0935 [M + H]<sup>+</sup>.

2-(4-Methyl-3-nitrophenylamino)-*N*-(2-methyl-5-(3-(trifluoromethyl)benzamido)phenyl)-4-(methylamino)pyrimidine-5-carboxamide (**17a**). (Method E) Yield 66%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.56 (s, 1H), 10.48 (s, 1H), 9.93 (s, 1H), 9.13 (s, 1H), 8.86 (s, 1H), 8.77 (s, 1H), 8.35–8.26 (m, 2H), 7.97 (d, *J* = 7.1 Hz, 1H), 7.86 (s, 1H), 7.80 (d, *J* = 7.5 Hz, 2H), 7.58 (d, *J* = 7.3 Hz, 1H), 7.46 (d, *J* = 8.3 Hz, 1H), 7.28 (d, *J* = 8.1 Hz, 1H), 3.07 (s, 3H), 2.50 (s, 3H), 2.23 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 165.03, 164.33, 161.96, 149.06, 138.72, 137.29, 136.23, 133.39, 132.27, 130.72, 130.16, 129.98, 128.42, 127.01, 124.90, 124.67, 123.04, 119.39, 118.94, 115.55, 102.03, 99.99, 28.29, 19.65, 17.90. LC-MS (ESI, *m/z*): 580.1850 [M + H]<sup>+</sup>.

4-(Dimethylamino)-2-(4-methyl-3-nitrophenylamino)-*N*-(2-methyl-5-(3-(trifluoromethyl)benzamido)phenyl)pyrimidine-5-carboxamide (**17b**). (Method E) Yield 65%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.52 (s, 1H), 10.38 (s, 1H), 10.02 (s, 1H), 8.77 (s, 1H), 8.36 (s, 1H), 8.32 (s, 1H), 8.28 (s, 1H), 7.98 (d, *J* = 7.2 Hz, 1H), 7.93 (s, 1H), 7.80 (d, *J* = 7.5 Hz, 1H), 7.76 (d, *J* = 8.0 Hz, 1H), 7.63 (d, *J* = 7.3 Hz, 1H), 7.45 (s, 1H), 7.26 (d, *J* = 7.9 Hz, 2H), 3.21 (s, 6H), 2.51 (s, 3H), 2.28 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 164.36, 160.30, 149.06, 138.87, 137.31, 136.42, 136.20, 133.41, 132.30, 130.83, 130.17, 129.80, 129.48, 128.62, 128.55, 126.70, 125.82, 124.74, 124.69, 123.11, 118.67, 118.14, 115.16, 108.39, 19.66, 17.93. LC-MS (ESI, *m/z*): 594.2013 [M + H]<sup>+</sup>.

2-(4-Methyl-3-nitrophenylamino)-*N*-(2-methyl-5-(3-(trifluoromethyl)benzamido)phenyl)pyrimidine-5-carboxamide (**17c**). (Method E) Yield 65%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.66 (s, 1H), 10.52 (s, 1H), 9.96 (s, 1H), 8.85 (s, 1H), 8.49 (s, 2H), 8.38–8.26 (m, 2H), 8.01–7.92 (m, 2H), 7.88 (s, 1H), 7.79 (d, *J* = 6.8 Hz, 1H), 7.60 (d, *J* = 7.3 Hz, 1H), 7.44 (d, *J* = 7.9 Hz, 1H), 7.27 (d, *J* = 7.7 Hz, 1H), 2.49 (d, *J* = 14.5 Hz, 3H), 2.24 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 164.66, 164.34, 163.37, 149.43, 138.32, 137.31,

136.20, 133.25, 132.25, 130.75, 130.10, 129.90, 128.51, 126.84, 125.06, 124.69, 119.34, 118.96, 115.69, 101.80, 19.36, 17.87. LC-MS (ESI, *m/z*): 551.1583 [M + H]<sup>+</sup>.

2-(3-Amino-4-methylphenylamino)-*N*-(2-methyl-5-(3-(trifluoromethyl)benzamido)phenyl)-4-(methylamino)pyrimidine-5-carboxamide (**18a**). (Method F) Yield 63%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.48 (s, 1H), 9.68 (s, 1H), 9.27 (s, 1H), 8.69 (s, 2H), 8.30 (d, *J* = 10.0 Hz, 2H), 7.97 (s, 1H), 7.81 (s, 2H), 7.59 (d, *J* = 6.2 Hz, 1H), 7.26 (d, *J* = 7.2 Hz, 1H), 7.14 (s, 1H), 6.93 (s, 1H), 6.83 (s, 1H), 4.74 (s, 2H), 3.18 (s, 1H), 3.00 (s, 3H), 2.22 (s, 3H), 2.02 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 166.27, 164.29, 162.49, 160.61, 157.36, 146.74, 139.21, 137.20, 136.81, 136.23, 132.30, 130.62, 130.19, 130.07, 128.54, 124.68, 119.46, 118.55, 115.52, 108.72, 106.10, 40.61, 40.41, 40.20, 39.99, 39.78, 39.58, 39.36, 27.85, 17.96, 17.38. LC-MS (ESI, *m/z*): 550.2110 [M + H]<sup>+</sup>.

2-(3-Amino-4-methylphenylamino)-4-(dimethylamino)-*N*-(2-methyl-5-(3-(trifluoromethyl)benzamido)phenyl)pyrimidine-5-carboxamide (**18b**). (Method F) Yield 73%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.50 (s, 1H), 9.78 (s, 1H), 9.02 (s, 1H), 8.32 (s, 1H), 8.29 (d, *J* = 7.9 Hz, 1H), 8.23 (s, 1H), 7.97 (d, *J* = 7.2 Hz, 1H), 7.89 (s, 1H), 7.79 (s, 1H), 7.73 (s, 1H), 7.68 (s, 1H), 7.63 (d, *J* = 8.3 Hz, 1H), 7.24 (d, *J* = 8.2 Hz, 1H), 7.08 (s, 1H), 6.87 (d, *J* = 7.9 Hz, 1H), 6.80 (d, *J* = 8.0 Hz, 1H), 3.11 (s, 6H), 2.26 (s, 3H), 2.01 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 167.42, 166.58, 164.31, 160.96, 159.40, 157.81, 146.71, 139.51, 137.25, 136.24, 132.31, 132.19, 131.98, 130.71, 130.15, 130.04, 129.13, 128.59, 125.83, 124.68, 118.22, 115.10, 108.40, 105.78, 30.48, 18.94. LC-MS (ESI, *m/z*): 564.2263 [M + H]<sup>+</sup>.

2-(3-Amino-4-methylphenylamino)-*N*-(2-methyl-5-(3-(trifluoromethyl)benzamido)phenyl)pyrimidine-5-carboxamide (**18c**). (Method F) Yield 72%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.49 (s, 1H), 9.67 (s, 1H), 9.18 (s, 1H), 8.77 (s, 1H), 8.34 (s, 1H), 8.30 (d, *J* = 7.3 Hz, 1H), 7.96 (d, *J* = 7.0 Hz, 1H), 7.84 (s, 1H), 7.80 (d, *J* = 7.3 Hz, 1H), 7.62 (d, *J* = 7.4 Hz, 2H), 7.27 (d, *J* = 7.5 Hz, 2H), 6.94 (d, *J* = 7.5 Hz, 1H), 6.83 (d, *J* = 7.9 Hz, 1H), 2.24 (s, 3H), 2.06 (d, *J* = 15.4 Hz, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 166.12, 164.32, 163.51, 160.81, 158.08, 146.70, 139.18, 137.23, 136.86, 136.24, 132.28, 130.64, 130.14, 130.05, 129.96, 128.52, 125.82, 124.66, 123.11, 119.48, 118.59, 115.64, 109.02, 106.52, 60.22, 17.94, 17.32, 14.51. LC-MS (ESI, *m/z*): 521.1843 [M + H]<sup>+</sup>.

*tert*-Butyl 3-amino-4-methylphenylcarbamate (**20**). To a solution of 4-methyl-3-nitroaniline (5.00 g, 32.9 mmol, 1.00 equiv) in anhydrous THF (50 mL) was added (Boc)<sub>2</sub>O (7.88 g, 36.1 mmol, 1.10 equiv) and DMAP (0.3 g) at 0 °C under argon. Then the reaction mixture was allowed to warm to room temperature for 1 h. After that, the reaction mixture was heated to reflux for 20 h. The resulting mixture was then concentrated to dryness. The residue was diluted with water (100 mL) and extracted with EtOAc (3 × 100 mL). The combined organic layers were washed with 1 N HCl (100 mL), water (100 mL), and brine (100 mL), and dried over anhydrous MgSO<sub>4</sub>. Evaporation of the solvent afforded the *tert*-butyl 4-methyl-3-nitrophenylcarbamate **19**, which was used in the next step without further purification. To a solution of crude **19** (6.90 g, 27.38 mmol, 1.00 equiv) in methanol (50 mL) was added 10% Pd/C (0.69 g, 10%) at room temperature under argon. Then, the reaction mixture was stirred under a balloon of hydrogen for 2 h. The resulting mixture was filtered and washed with methanol. The filtrate was concentrated to afford the crude product **20**, which was crystallized from EtOAc/hexanes as a needle solid (5 g, 83%). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 8.92 (s, 1H), 6.87 (s, 1H), 6.76 (d, *J* = 7.7 Hz, 1H), 6.53 (d, *J* = 6.8 Hz, 1H), 4.73 (s, 2H), 1.99 (s, 3H), 1.47 (s, 9H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 153.22, 146.94, 138.40, 130.13, 115.54, 107.28, 104.82, 78.81, 28.67, 17.27. LC-MS (ESI, *m/z*): 223.1375 [M + H]<sup>+</sup>.

*tert*-Butyl 4-Methyl-3-(4-(methylamino)-2-(methylthio)pyrimidine-5-carboxamido)phenylcarbamate (**21**). To a solution of 4-(methylamino)-2-(methylthio)pyrimidine-5-carboxylic acid (4.0 g, 20.0 mmol, 1.00 equiv) in anhydrous DMF (20 mL) was added *tert*-butyl 3-amino-4-methylphenylcarbamate (4.98 g, 3.6 mmol, 1.10 equiv), HATU (9.88 g, 26.0 mmol, 1.2 equiv), and DIPEA (12 mL, 70.0 mmol, 3.5 equiv) at room temperature under argon. The reaction mixture was stirred at room temperature for 20 h. The resulting



mixture was concentrated to dryness. The residue was diluted with water (100 mL) and extracted with EtOAc (3 × 100 mL). The combined organic layers were washed with water (100 mL) and brine (100 mL), and dried over anhydrous MgSO<sub>4</sub>. Evaporation of the solvent afforded the crude product, which was purified by flash column chromatography (eluting with MeOH in DCM 0–5%) to offer **21** as a white solid (6.60 g, 82%). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 9.89 (s, 1H), 9.33 (s, 1H), 8.67 (s, 2H), 7.51 (s, 1H), 7.22 (d, *J* = 8.0 Hz, 1H), 7.13 (d, *J* = 8.1 Hz, 1H), 2.96 (d, *J* = 3.1 Hz, 3H), 2.52 (s, 3H), 2.13 (s, 3H), 1.48 (s, 9H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 173.60, 165.68, 160.55, 155.22, 153.24, 138.04, 136.26, 130.63, 127.94, 117.00, 116.72, 104.82, 79.40, 28.60, 27.58, 17.72, 13.98. LC-MS (ESI, *m/z*): 404.1685 [M + H]<sup>+</sup>.

*N*-(5-Amino-2-methylphenyl)-4-(methylamino)-2-(methylthio)pyrimidine-5-carboxamide (**22**). To a solution of *tert*-butyl 4-methyl-3-(4-(methylamino)-2-(methylthio)pyrimidine-5-carboxamido)-phenylcarbamate (4.03 g, 10.0 mmol, 1.00 equiv) in methanol (15 mL) was added 10 mL of 4 M HCl (in methanol) at room temperature under argon. The reaction mixture was stirred at room temperature for 2 h. The resulting mixture was concentrated to provide **22** as an off white solid (4.2 g, 95%). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.72 (s, 1H), 9.51 (s, 1H), 9.00 (s, 1H), 7.41 (s, 1H), 7.26 (s, 1H), 3.08 (s, 3H), 2.64 (s, 3H), 2.25 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 168.07, 163.84, 159.40, 147.58, 136.68, 134.31, 132.62, 131.88, 131.10, 129.92, 121.65, 104.86, 28.66, 18.17, 14.02. LC-MS (ESI, *m/z*): 304.1163 [M + H]<sup>+</sup>.

**General Method I.** *N*-(5-(2-Methoxybenzamido)-2-methylphenyl)-4-(methylamino)-2-(methylthio)pyrimidine-5-carboxamide (**23a**). To a solution of *N*-(5-amino-2-methylphenyl)-4-(methylamino)-2-(methylthio)pyrimidine-5-carboxamide (0.40 g, 0.9 mmol, 1.00 equiv) in anhydrous THF (10 mL) was added a solution of 2-methylbenzoyl chloride (0.13 mL, 0.93 mmol, 1.05 equiv) and DIPEA (0.96 mL, 5.4 mmol, 6.0 equiv) at 0 °C under argon. The reaction mixture was stirred at 0 °C under argon for 2 h. The resulting mixture was concentrated to dryness. The residue was diluted with water (50 mL) and extracted with EtOAc (3 × 50 mL). The combined organic layers were washed with water (50 mL) and brine (50 mL), and dried over anhydrous MgSO<sub>4</sub>. Evaporation of the solvent afforded the crude product, which was purified by flash column chromatography (eluting with MeOH in DCM 0–5%) to offer **23a** as a white solid (0.34 g, 89%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.44 (s, 1H), 8.13 (d, *J* = 7.2 Hz, 1H), 7.75 (s, 1H), 7.45 (dd, *J* = 19.1, 12.3 Hz, 1H), 7.37 (d, *J* = 7.5 Hz, 1H), 7.15 (d, *J* = 8.0 Hz, 1H), 7.09–7.01 (m, 1H), 6.97 (d, *J* = 8.0 Hz, 1H), 3.99 (s, 3H), 3.00 (s, 3H), 2.51 (s, 3H), 2.24 (d, *J* = 28.4 Hz, 3H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 174.33, 166.08, 163.51, 160.55, 157.25, 153.78, 136.32, 135.46, 133.34, 132.19, 130.86, 129.33, 121.51, 121.41, 118.79, 118.16, 111.53, 104.18, 56.16, 27.08, 17.44, 14.00. LC-MS (ESI, *m/z*): 438.1560 [M + H]<sup>+</sup>.

*N*-(5-(2-Methoxybenzamido)-2-methylphenyl)-4-(methylamino)-2-(methylthio)pyrimidine-5-carboxamide (**23b**). (Method I) <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.25 (s, 1H), 9.98 (s, 1H), 8.70 (s, 1H), 8.66 (s, 1H), 7.84 (s, 1H), 7.60 (d, *J* = 8.3 Hz, 1H), 7.56 (d, *J* = 6.8 Hz, 1H), 7.52 (s, 1H), 7.46 (d, *J* = 6.6 Hz, 1H), 7.25 (d, *J* = 8.1 Hz, 1H), 7.17 (t, *J* = 8.9 Hz, 2H), 3.85 (s, 3H), 2.96 (d, *J* = 4.2 Hz, 3H), 2.21 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 173.63, 165.77, 165.56, 160.56, 159.66, 155.25, 137.60, 136.76, 136.18, 130.62, 130.13, 129.99, 129.72, 122.02, 120.33, 119.38, 118.90, 117.74, 114.41, 113.36, 104.88, 55.80, 27.38, 17.72, 13.99. LC-MS (ESI, *m/z*): 438.1537 [M + H]<sup>+</sup>.

*N*-(5-(4-Methoxybenzamido)-2-methylphenyl)-4-(methylamino)-2-(methylthio)pyrimidine-5-carboxamide (**23c**). (Method I) Yield 79%. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.35 (s, 1H), 7.74 (d, *J* = 7.3 Hz, 2H), 7.63 (s, 1H), 7.28 (d, *J* = 6.4 Hz, 1H), 7.03 (d, *J* = 7.5 Hz, 1H), 6.82 (d, *J* = 7.4 Hz, 2H), 3.78 (s, 3H), 2.96 (s, 3H), 2.49 (s, 3H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 174.27, 166.21, 166.08, 162.32, 160.46, 153.83, 136.45, 135.09, 130.69, 129.43, 129.17, 126.75, 119.13, 118.52, 113.61, 104.06, 55.32, 27.05, 17.30, 13.97. LC-MS (ESI, *m/z*): 438.1522 [M + H]<sup>+</sup>.

*N*-(5-(3,4-Dimethoxybenzamido)-2-methylphenyl)-4-(methylamino)-2-(methylthio)pyrimidine-5-carboxamide (**23d**). (Method I)

Yield 85%. <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>OD) δ 8.95 (s, 1H), 8.88 (s, 1H), 8.83 (s, 1H), 8.74 (s, 1H), 8.16 (s, 1H), 7.72 (s, 2H), 7.59 (d, *J* = 7.5 Hz, 1H), 7.36 (d, *J* = 8.1 Hz, 1H), 7.10 (d, *J* = 8.1 Hz, 1H), 4.21 (s, 3H), 4.17 (s, 3H), 3.32 (d, *J* = 4.2 Hz, 3H), 2.86 (s, 3H), 2.45 (s, 3H). <sup>13</sup>C NMR (101 MHz, CD<sub>3</sub>OD) δ 170.53, 162.15, 161.83, 156.59, 149.92, 147.99, 144.88, 132.48, 131.34, 126.76, 124.71, 123.08, 116.24, 114.80, 113.96, 106.69, 106.28, 100.14, 56.49, 52.02, 23.33, 13.46, 10.16. LC-MS (ESI, *m/z*): 468.26329 [M + H]<sup>+</sup>.

*N*-(5-(3,5-Dimethoxybenzamido)-2-methylphenyl)-4-(methylamino)-2-(methylthio)pyrimidine-5-carboxamide (**23e**). (Method I) Yield 84%. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.98 (s, 1H), 7.19 (s, 1H), 6.96 (d, *J* = 7.2 Hz, 1H), 6.71 (d, *J* = 8.0 Hz, 1H), 6.55 (s, 2H), 6.11 (s, 1H), 4.10 (s, 6H), 2.56 (s, 3H), 2.06 (s, 3H), 1.73 (s, 3H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 176.97, 169.65, 169.21, 163.56, 163.32, 156.55, 139.54, 139.27, 137.96, 133.41, 133.24, 122.37, 122.11, 108.09, 106.77, 106.45, 57.96, 29.56, 19.83, 16.35. LC-MS (ESI, *m/z*): 468.1630 [M + H]<sup>+</sup>.

*N*-(2-Methyl-5-(3,4,5-trimethoxybenzamido)phenyl)-4-(methylamino)-2-(methylthio)pyrimidine-5-carboxamide (**23f**). (Method I) Yield 81%. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.54–8.45 (m, 1H), 7.81–7.72 (m, 1H), 7.51–7.41 (m, 1H), 7.29–7.15 (m, 3H), 3.93–3.88 (m, 6H), 3.87–3.84 (m, 3H), 3.07–3.02 (m, 3H), 2.58–2.53 (m, 3H), 2.25–2.19 (m, 3H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 176.83, 169.20, 169.08, 163.16, 156.46, 155.54, 143.37, 139.28, 137.90, 133.18, 132.99, 132.63, 122.17, 122.06, 107.61, 106.65, 62.70, 58.27, 29.24, 19.66, 15.93. LC-MS (ESI, *m/z*): 498.1782 [M + H]<sup>+</sup>.

*N*-(5-(Benzod[1,3]dioxole-5-carboxamido)-2-methylphenyl)-4-(methylamino)-2-(methylthio)pyrimidine-5-carboxamide (**23g**). (Method I) Yield 80%. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.73 (s, 1H), 7.92 (s, 1H), 7.76–7.65 (m, 2H), 7.64 (s, 1H), 7.45 (d, *J* = 5.1 Hz, 1H), 7.12 (d, *J* = 4.7 Hz, 1H), 6.31 (s, 2H), 3.10 (s, 2H), 2.85 (s, 3H), 2.49 (s, 3H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 177.26, 169.44, 169.23, 163.58, 156.83, 153.63, 150.92, 139.59, 138.17, 133.77, 133.18, 131.68, 125.45, 122.54, 122.11, 110.91, 110.79, 107.05, 104.81, 41.42, 30.04, 20.29, 16.89. LC-MS (ESI, *m/z*): 452.1319 [M + H]<sup>+</sup>.

*N*-(5-(2,3-Dihydrobenzo[1,4]dioxine-6-carboxamido)-2-methylphenyl)-4-(methylamino)-2-(methylthio)pyrimidine-5-carboxamide (**23h**). (Method I) Yield 87%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.07 (s, 1H), 9.95 (s, 1H), 8.68 (d, *J* = 12.8 Hz, 2H), 7.83 (s, 1H), 7.56 (s, 3H), 7.23 (s, 1H), 7.00 (s, 1H), 4.32 (s, 4H), 2.97 (s, 3H), 2.52 (d, *J* = 4.6 Hz, 3H), 2.20 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 173.63, 165.75, 164.85, 160.55, 155.22, 146.83, 143.41, 137.78, 136.13, 130.58, 129.46, 128.16, 121.67, 119.23, 118.77, 117.30, 117.14, 104.89, 64.87, 64.50, 27.58, 17.86, 13.99. LC-MS (ESI, *m/z*): 467.1485 [M + H]<sup>+</sup>.

*N*-(5-(2-Methoxybenzamido)-2-methylphenyl)-4-(methylamino)-2-(methylsulfonyl)pyrimidine-5-carboxamide (**24a**). (Method D) Yield 60%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.30 (s, 1H), 10.16 (s, 1H), 8.92 (s, 1H), 8.86 (s, 1H), 7.88 (s, 1H), 7.65 (s, 1H), 7.52 (s, 2H), 7.26 (d, *J* = 7.9 Hz, 1H), 7.18 (d, *J* = 6.8 Hz, 1H), 7.08 (s, 1H), 3.91 (s, 3H), 3.02 (s, 3H), 2.38 (s, 3H), 2.22 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 166.66, 164.85, 164.37, 161.41, 156.97, 155.64, 137.62, 135.85, 132.53, 130.85, 130.15, 129.52, 128.38, 125.29, 120.99, 118.49, 112.49, 111.25, 56.47, 39.17, 28.01, 17.88. LC-MS (ESI, *m/z*): 470.1735 [M + H]<sup>+</sup>.

*N*-(5-(3-Methoxybenzamido)-2-methylphenyl)-4-(methylamino)-2-(methylsulfonyl)pyrimidine-5-carboxamide (**24b**). (Method D) Yield 81%. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 9.27 (s, 1H), 8.64 (s, 1H), 8.38 (s, 1H), 8.28 (s, 1H), 8.05 (s, 1H), 7.33 (s, 2H), 7.25 (d, *J* = 6.9 Hz, 1H), 7.10–6.93 (m, 3H), 3.77 (s, 3H), 3.21 (s, 3H), 2.88 (s, 3H), 2.04 (s, 3H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 166.67, 165.98, 164.82, 160.86, 159.68, 154.38, 136.15, 135.78, 134.93, 130.80, 130.26, 129.58, 119.44, 118.87, 117.90, 117.08, 112.68, 111.01, 55.48, 38.56, 27.73, 17.64. LC-MS (ESI, *m/z*): 470.1426 [M + H]<sup>+</sup>.

*N*-(5-(4-Methoxybenzamido)-2-methylphenyl)-4-(methylamino)-2-(methylsulfonyl)pyrimidine-5-carboxamide (**24c**). (Method D) Yield 87%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.26 (s, 1H), 10.13 (s, 1H), 8.91 (s, 1H), 8.84 (s, 1H), 7.98 (d, *J* = 7.4 Hz, 3H), 7.88 (s, 1H), 7.59 (d, *J* = 6.7 Hz, 1H), 7.26 (d, *J* = 7.5 Hz, 1H), 7.07 (d, *J* = 7.5 Hz, 3H), 3.85 (s, 3H), 3.38 (s, 3H), 3.01 (s, 3H), 2.22 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 166.65, 165.26, 164.34, 162.38, 161.39,

155.57, 137.91, 135.68, 130.72, 130.03, 129.30, 127.36, 119.09, 114.08, 111.34, 55.90, 39.16, 28.17, 17.85. LC-MS (ESI,  $m/z$ ): 470.1430 [M + H]<sup>+</sup>.

*N*-(5-(3,4-Dimethoxybenzamido)-2-methylphenyl)-4-(methylamino)-2-(methylsulfonyl)pyrimidine-5-carboxamide (**24d**). (Method D) Yield 90%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.25 (s, 1H), 10.10 (s, 1H), 8.90 (s, 1H), 8.83 (s, 1H), 7.85 (s, 1H), 7.64 (d, *J* = 7.5 Hz, 1H), 7.60 (d, *J* = 8.6 Hz, 1H), 7.56 (s, 1H), 7.27 (d, *J* = 8.1 Hz, 2H), 7.09 (d, *J* = 8.0 Hz, 1H), 3.86 (s, 6H), 3.38 (s, 3H), 3.01 (d, *J* = 3.6 Hz, 3H), 2.22 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 166.64, 165.25, 164.35, 161.38, 155.55, 152.13, 148.78, 137.83, 135.68, 133.17, 131.11, 130.72, 129.35, 128.38, 127.35, 121.53, 119.26, 111.50, 56.15, 39.15, 28.17, 17.83. LC-MS (ESI,  $m/z$ ): 500.1535 [M + H]<sup>+</sup>.

*N*-(5-(3,5-Dimethoxybenzamido)-2-methylphenyl)-4-(methylamino)-2-(methylsulfonyl)pyrimidine-5-carboxamide (**24e**). (Method D) Yield 76%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.17 (s, 1H), 9.66 (s, 1H), 9.24 (s, 1H), 8.70 (s, 2H), 7.79 (s, 1H), 7.58 (d, *J* = 8.0 Hz, 1H), 7.24 (d, *J* = 8.0 Hz, 1H), 7.14 (d, *J* = 8.1 Hz, 3H), 6.94 (d, *J* = 7.8 Hz, 1H), 6.83 (d, *J* = 7.9 Hz, 1H), 6.72 (s, 1H), 4.73 (s, 2H), 3.84 (s, 6H), 3.39 (s, 5H), 3.00 (d, *J* = 3.1 Hz, 3H), 2.21 (s, 3H), 2.02 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 166.65, 165.38, 164.37, 161.39, 160.86, 155.58, 137.56, 137.33, 135.72, 131.09, 130.76, 129.67, 129.30, 119.28, 111.33, 106.08, 103.81, 55.96, 55.49, 28.44, 28.16, 17.85, 17.72. LC-MS (ESI,  $m/z$ ): 500.1530 [M + H]<sup>+</sup>.

*N*-(2-Methyl-5-(3,4,5-trimethoxybenzamido)phenyl)-4-(methylamino)-2-(methylsulfonyl)pyrimidine-5-carboxamide (**24f**). (Method D) Yield 82%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.28 (s, 1H), 10.18 (s, 1H), 8.92 (s, 1H), 8.84 (s, 1H), 7.85 (s, 1H), 7.62 (d, *J* = 6.1 Hz, 1H), 7.32 (s, 3H), 3.89 (s, 6H), 3.75 (s, 3H), 3.39 (s, 3H), 3.02 (s, 3H), 2.25 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 166.65, 165.23, 164.40, 161.39, 155.57, 153.11, 140.81, 137.59, 135.73, 130.77, 130.36, 129.63, 119.42, 111.34, 105.75, 60.85, 56.13, 28.52, 18.00, 15.43. LC-MS (ESI,  $m/z$ ): 530.1645 [M + H]<sup>+</sup>.

*N*-(5-(Benzo[d][1,3]dioxole-5-carboxamido)-2-methylphenyl)-4-(methylamino)-2-(methylsulfonyl)pyrimidine-5-carboxamide (**24g**). (Method D) Yield 72%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.26 (s, 1H), 10.10 (s, 1H), 8.91 (s, 1H), 8.84 (s, 1H), 7.87 (s, 1H), 7.57 (dd, *J* = 18.1, 9.8 Hz, 3H), 7.26 (d, *J* = 7.8 Hz, 1H), 7.06 (d, *J* = 7.8 Hz, 1H), 6.14 (s, 2H), 3.38 (s, 3H), 3.01 (d, *J* = 3.1 Hz, 3H), 2.22 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 166.65, 164.86, 164.35, 161.39, 155.57, 150.52, 147.85, 137.79, 135.69, 130.73, 129.42, 129.11, 123.30, 119.10, 111.34, 108.39, 108.16, 102.29, 39.15, 28.16, 17.85. LC-MS (ESI,  $m/z$ ): 484.1222 [M + H]<sup>+</sup>.

*N*-(5-(2,3-Dihydrobenzo[b][1,4]dioxine-6-carboxamido)-2-methylphenyl)-4-(methylamino)-2-(methylsulfonyl)pyrimidine-5-carboxamide (**24h**). (Method D) Yield 79%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.07 (d, *J* = 12.6 Hz, 2H), 9.85 (s, 1H), 9.17 (s, 1H), 8.41 (s, 1H), 7.80 (s, 1H), 7.28–7.18 (m, 2H), 6.99 (d, *J* = 7.3 Hz, 2H), 4.32 (s, 4H), 2.93 (s, 3H), 2.50 (s, 3H), 2.17 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 164.89, 164.62, 155.57, 153.77, 146.84, 146.58, 143.42, 137.84, 135.92, 130.62, 129.41, 128.17, 121.66, 119.14, 118.84, 117.31, 117.14, 64.89, 64.51, 46.12, 28.21, 17.84. LC-MS (ESI,  $m/z$ ): 498.1375 [M + H]<sup>+</sup>.

*N*-(5-(2-Methoxybenzamido)-2-methylphenyl)-4-(methylamino)-2-(methylsulfonyl)pyrimidine-5-carboxamide (**25a**). (Method E) Yield 76%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.61 (s, 1H), 10.12 (s, 1H), 9.95 (s, 1H), 9.17 (s, 1H), 8.86 (s, 1H), 8.76 (s, 1H), 7.82 (s, 2H), 7.64 (d, *J* = 6.8 Hz, 1H), 7.55–7.43 (m, 3H), 7.21 (dd, *J* = 18.5, 8.2 Hz, 2H), 7.08 (d, *J* = 6.7 Hz, 1H), 3.91 (s, 3H), 3.06 (s, 3H), 2.50 (s, 3H), 2.20 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 164.95, 164.81, 161.91, 156.95, 149.03, 138.65, 137.57, 136.26, 133.41, 132.46, 130.72, 130.12, 129.55, 127.06, 125.39, 124.91, 120.97, 118.57, 118.19, 115.54, 112.48, 101.98, 56.36, 28.31, 19.69, 17.91. LC-MS (ESI,  $m/z$ ): 542.2080 [M + H]<sup>+</sup>.

*N*-(5-(3-Methoxybenzamido)-2-methylphenyl)-2-(4-methyl-3-nitrophenylamino)-4-(methylamino)pyrimidine-5-carboxamide (**25b**). (Method E) Yield 86%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.65 (s, 1H), 10.24 (s, 1H), 9.95 (s, 1H), 9.18 (s, 1H), 8.86 (s, 1H), 8.77 (s, 1H), 7.85 (s, 1H), 7.79 (d, *J* = 8.0 Hz, 1H), 7.56 (s, 2H), 7.50 (s, 1H), 7.47 (s, 1H), 7.45 (s, 1H), 7.25 (d, *J* = 8.1 Hz, 1H), 7.16 (d, *J* = 7.7 Hz, 1H), 3.85 (s, 3H), 3.06 (s, 4H), 2.50 (s, 3H), 2.22 (s, 4H).

<sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 165.58, 164.89, 161.88, 159.66, 149.02, 137.60, 136.78, 136.12, 133.42, 130.65, 130.00, 129.66, 127.17, 124.95, 120.32, 119.29, 118.88, 117.71, 115.60, 113.37, 102.04, 55.79, 28.34, 19.70, 17.89. LC-MS (ESI,  $m/z$ ): 542.2083 [M + H]<sup>+</sup>.

*N*-(5-(4-Methoxybenzamido)-2-methylphenyl)-2-(4-methyl-3-nitrophenylamino)-4-(methylamino)pyrimidine-5-carboxamide (**25c**). (Method E) Yield 78%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.34 (s, 1H), 10.10 (s, 1H), 9.89 (s, 1H), 9.03 (s, 1H), 8.87 (s, 1H), 8.74 (s, 1H), 7.97 (d, *J* = 8.5 Hz, 2H), 7.81 (d, *J* = 9.6 Hz, 2H), 7.54 (d, *J* = 8.2 Hz, 1H), 7.46 (d, *J* = 8.4 Hz, 1H), 7.23 (d, *J* = 8.1 Hz, 1H), 7.07 (d, *J* = 8.6 Hz, 2H), 3.85 (s, 3H), 3.04 (d, *J* = 3.7 Hz, 3H), 2.20 (s, 3H), 2.00 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 170.84, 165.26, 162.36, 162.04, 158.65, 153.87, 149.07, 138.96, 137.80, 136.19, 133.36, 130.59, 130.01, 129.41, 127.38, 126.69, 124.83, 119.25, 118.77, 115.37, 114.07, 60.41, 55.69, 21.44, 14.80. LC-MS (ESI,  $m/z$ ): 542.2090 [M + H]<sup>+</sup>.

*N*-(5-(3,4-Dimethoxybenzamido)-2-methylphenyl)-2-(4-methyl-3-nitrophenylamino)-4-(methylamino)pyrimidine-5-carboxamide (**25d**). (Method E) Yield 75%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.58 (s, 1H), 10.09 (s, 1H), 9.92 (s, 1H), 9.14 (s, 1H), 8.86 (s, 1H), 8.76 (s, 1H), 7.80 (d, *J* = 13.2 Hz, 2H), 7.63 (d, *J* = 8.1 Hz, 1H), 7.56 (d, *J* = 8.0 Hz, 2H), 7.46 (d, *J* = 8.2 Hz, 1H), 7.24 (d, *J* = 8.0 Hz, 1H), 7.09 (d, *J* = 8.0 Hz, 1H), 3.85 (s, 6H), 3.06 (s, 3H), 2.50 (s, 3H), 2.21 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 165.24, 164.98, 161.93, 152.10, 149.03, 148.77, 137.77, 136.10, 133.41, 130.60, 129.40, 127.40, 127.05, 124.91, 121.51, 119.37, 118.91, 115.53, 111.48, 111.40, 102.01, 56.10, 28.30, 19.69, 17.91. LC-MS (ESI,  $m/z$ ): 572.2185 [M + H]<sup>+</sup>.

*N*-(5-(3,5-Dimethoxybenzamido)-2-methylphenyl)-2-(4-methyl-3-nitrophenylamino)-4-(methylamino)pyrimidine-5-carboxamide (**25e**). (Method E) Yield 70%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.46 (s, 1H), 10.19 (s, 1H), 9.92 (s, 1H), 9.09 (s, 1H), 8.85 (s, 1H), 8.74 (s, 1H), 7.79 (d, *J* = 9.7 Hz, 2H), 7.55 (d, *J* = 7.8 Hz, 1H), 7.46 (d, *J* = 8.1 Hz, 1H), 7.25 (d, *J* = 8.2 Hz, 1H), 7.10 (s, 2H), 6.71 (s, 1H), 3.82 (s, 9H), 2.51 (s, 3H), 2.21 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 165.40, 165.07, 161.95, 160.85, 149.04, 138.71, 137.46, 137.36, 136.16, 133.40, 130.66, 129.77, 127.00, 124.92, 119.42, 118.95, 115.51, 106.05, 103.78, 101.99, 55.96, 28.26, 19.66, 17.87. LC-MS (ESI,  $m/z$ ): 572.2183 [M + H]<sup>+</sup>.

2-(4-Methyl-3-nitrophenylamino)-*N*-(2-methyl-5-(3,4,5-trimethoxybenzamido)phenyl)-4-(methylamino)pyrimidine-5-carboxamide (**25f**). (Method E) Yield 65%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.75 (s, 1H), 10.15 (s, 1H), 9.95 (s, 1H), 9.22 (s, 1H), 8.86 (s, 1H), 8.78 (s, 1H), 7.80 (s, 1H), 7.77 (s, 1H), 7.57 (d, *J* = 7.5 Hz, 1H), 7.46 (d, *J* = 8.3 Hz, 1H), 7.30 (s, 2H), 7.26 (d, *J* = 8.1 Hz, 1H), 3.88 (s, 6H), 3.74 (s, 3H), 3.07 (s, 3H), 2.50 (s, 3H), 2.23 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 165.18, 164.82, 161.85, 153.09, 148.98, 140.78, 138.37, 137.55, 136.09, 133.42, 130.65, 130.42, 129.62, 127.25, 124.94, 119.49, 119.06, 115.61, 105.73, 102.03, 60.63, 56.13, 28.09, 20.07, 17.99. LC-MS (ESI,  $m/z$ ): 602.2293 [M + H]<sup>+</sup>.

*N*-(5-(Benzo[d][1,3]dioxole-5-carboxamido)-2-methylphenyl)-2-(4-methyl-3-nitrophenylamino)-4-(methylamino)pyrimidine-5-carboxamide (**25g**). (Method E) Yield 79%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.83 (s, 1H), 10.07 (s, 1H), 9.95 (s, 1H), 9.27 (s, 1H), 8.84 (s, 1H), 8.78 (s, 1H), 7.85 (s, 1H), 7.75 (d, *J* = 7.2 Hz, 1H), 7.59 (d, *J* = 7.8 Hz, 1H), 7.54 (d, *J* = 9.5 Hz, 2H), 7.44 (d, *J* = 7.9 Hz, 1H), 7.23 (d, *J* = 7.9 Hz, 1H), 7.04 (d, *J* = 7.8 Hz, 1H), 6.13 (s, 2H), 3.06 (s, 3H), 2.49 (s, 3H), 2.22 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 164.83, 161.76, 159.62, 150.49, 148.89, 147.89, 147.83, 137.74, 135.98, 133.43, 130.62, 129.44, 129.14, 124.94, 123.27, 119.18, 115.66, 108.34, 108.15, 102.26, 102.11, 28.23, 19.93, 17.85. LC-MS (ESI,  $m/z$ ): 556.1870 [M + H]<sup>+</sup>.

*N*-(5-(2,3-Dihydrobenzo[b][1,4]dioxine-6-carboxamido)-2-methylphenyl)-2-(4-methyl-3-nitrophenylamino)-4-(methylamino)pyrimidine-5-carboxamide (**25h**). (Method E) Yield 67%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.58 (s, 1H), 10.06 (s, 1H), 9.92 (s, 1H), 9.16 (s, 1H), 8.86 (s, 1H), 8.77 (s, 1H), 7.83 (d, *J* = 12.8 Hz, 2H), 7.51 (d, *J* = 25.3 Hz, 3H), 7.24 (s, 1H), 7.00 (s, 1H), 4.32 (s, 4H), 3.06 (s, 3H), 2.51 (s, 3H), 2.21 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 164.86, 161.93, 152.17, 149.05, 146.85, 143.42, 138.57, 137.78, 136.07, 133.42, 130.60, 129.42, 128.16, 127.11, 124.97, 121.66, 119.19, 118.75,

117.31, 117.14, 115.62, 102.06, 64.89, 64.52, 28.33, 19.70, 17.88. LC-MS (ESI,  $m/z$ ): 570.2030 [M + H]<sup>+</sup>.

**2-(3-Amino-4-methylphenylamino)-N-(5-(2-methoxybenzamido)-2-methylphenyl)-4-(methylamino)pyrimidine-5-carboxamide (26a).** (Method F) Yield 63%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.22 (s, 1H), 9.67 (s, 1H), 9.25 (s, 1H), 8.71 (s, 2H), 7.82 (s, 1H), 7.58 (s, 2H), 7.55–7.41 (m, 2H), 7.24 (d, *J* = 7.6 Hz, 1H), 7.16 (s, 2H), 6.95 (d, *J* = 6.2 Hz, 1H), 6.84 (s, 1H), 3.85 (s, 3H), 3.01 (s, 3H), 2.22 (s, 3H), 2.03 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 166.27, 165.55, 162.51, 160.62, 159.66, 157.34, 146.73, 139.22, 137.52, 136.83, 136.73, 130.52, 130.07, 130.01, 129.71, 120.33, 119.43, 118.52, 117.72, 115.54, 113.35, 108.76, 106.13, 99.99, 55.80, 27.84, 17.95, 17.38. LC-MS (ESI,  $m/z$ ): 512.2340 [M + H]<sup>+</sup>.

**2-(3-Amino-4-methylphenylamino)-N-(5-(3-methoxybenzamido)-2-methylphenyl)-4-(methylamino)pyrimidine-5-carboxamide (26b).** (Method F) Yield 63%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.22 (s, 1H), 9.67 (s, 1H), 9.25 (s, 1H), 8.71 (s, 2H), 7.82 (s, 1H), 7.58 (s, 2H), 7.55–7.41 (m, 2H), 7.24 (d, *J* = 7.6 Hz, 1H), 7.16 (s, 2H), 6.95 (d, *J* = 6.2 Hz, 1H), 6.84 (s, 1H), 3.85 (s, 3H), 3.01 (s, 3H), 2.22 (s, 3H), 2.03 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 166.27, 165.55, 162.51, 160.62, 159.66, 157.34, 146.73, 139.22, 137.52, 136.83, 136.73, 130.52, 130.07, 130.01, 129.71, 120.33, 119.43, 118.52, 117.72, 115.54, 113.35, 108.76, 106.13, 99.99, 55.80, 27.84, 17.95, 17.38. LC-MS (ESI,  $m/z$ ): 512.2340 [M + H]<sup>+</sup>.

**2-(3-Amino-4-methylphenylamino)-N-(5-(4-methoxybenzamido)-2-methylphenyl)-4-(methylamino)pyrimidine-5-carboxamide (26c).** (Method F) Yield 83%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.23 (s, 1H), 9.81 (s, 1H), 9.38 (s, 1H), 8.85 (s, 2H), 8.12 (d, *J* = 8.4 Hz, 2H), 7.96 (s, 1H), 7.71 (d, *J* = 6.6 Hz, 1H), 7.35 (d, *J* = 8.2 Hz, 1H), 7.30 (s, 1H), 7.20 (d, *J* = 8.4 Hz, 2H), 7.10 (d, *J* = 6.6 Hz, 1H), 6.97 (d, *J* = 7.9 Hz, 1H), 3.97 (s, 3H), 3.57 (s, 3H), 2.34 (s, 3H), 2.17 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 166.30, 165.26, 162.52, 162.35, 160.63, 157.33, 146.71, 139.23, 137.78, 136.69, 130.49, 130.10, 130.03, 129.44, 127.46, 119.38, 118.48, 115.59, 114.06, 108.82, 106.17, 100.20, 55.87, 27.84, 17.93, 17.36. LC-MS (ESI,  $m/z$ ): 512.2340 [M + H]<sup>+</sup>.

**2-(3-Amino-4-methylphenylamino)-N-(5-(3,4-dimethoxybenzamido)-2-methylphenyl)-4-(methylamino)pyrimidine-5-carboxamide (26d).** (Method F) Yield 81%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.06 (s, 1H), 9.65 (s, 1H), 9.23 (s, 1H), 8.70 (s, 2H), 7.78 (s, 1H), 7.64 (d, *J* = 8.3 Hz, 1H), 7.57 (d, *J* = 9.2 Hz, 2H), 7.23 (d, *J* = 8.1 Hz, 1H), 7.15 (s, 1H), 7.09 (d, *J* = 8.4 Hz, 1H), 6.94 (d, *J* = 7.8 Hz, 1H), 6.83 (d, *J* = 8.0 Hz, 1H), 3.86 (s, 3H), 3.40 (s, 3H), 3.00 (d, *J* = 4.3 Hz, 3H), 2.21 (s, 3H), 2.02 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 166.28, 165.22, 162.50, 160.61, 157.32, 152.09, 148.78, 146.72, 139.21, 137.69, 136.68, 130.48, 130.08, 129.45, 127.45, 121.51, 119.50, 118.57, 115.56, 111.49, 111.42, 108.77, 106.14, 100.17, 56.25, 27.83, 17.71, 17.07. LC-MS (ESI,  $m/z$ ): 542.2445 [M + H]<sup>+</sup>.

**2-(3-Amino-4-methylphenylamino)-N-(5-(3,5-dimethoxybenzamido)-2-methylphenyl)-4-(methylamino)pyrimidine-5-carboxamide (26e).** (Method F) Yield 75%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.17 (s, 1H), 9.66 (s, 1H), 9.24 (s, 1H), 8.70 (s, 2H), 7.79 (s, 1H), 7.58 (d, *J* = 8.0 Hz, 1H), 7.24 (d, *J* = 8.0 Hz, 1H), 7.14 (d, *J* = 8.1 Hz, 3H), 6.94 (d, *J* = 7.8 Hz, 1H), 6.83 (d, *J* = 7.9 Hz, 1H), 6.72 (s, 1H), 4.73 (s, 2H), 3.84 (s, 6H), 3.39 (s, 3H), 3.00 (d, *J* = 3.1 Hz, 3H), 2.21 (s, 3H), 2.02 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 166.28, 165.33, 162.50, 160.86, 160.62, 157.33, 146.73, 139.21, 137.43, 136.72, 130.53, 130.08, 129.75, 119.51, 118.58, 115.56, 108.76, 106.13, 106.06, 103.81, 100.15, 55.97, 27.83, 17.93, 17.36. LC-MS (ESI,  $m/z$ ): 542.2440 [M + H]<sup>+</sup>.

**2-(3-Amino-4-methylphenylamino)-N-(2-methyl-5-(3,4,5-trimethoxybenzamido)phenyl)-4-(methylamino)pyrimidine-5-carboxamide (26f).** (Method F) Yield 65%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.13 (s, 1H), 9.66 (s, 1H), 9.24 (s, 1H), 8.71 (s, 2H), 7.77 (s, 1H), 7.59 (d, *J* = 7.4 Hz, 1H), 7.32 (s, 2H), 7.25 (d, *J* = 7.8 Hz, 1H), 7.16 (s, 1H), 6.95 (d, *J* = 7.1 Hz, 1H), 6.83 (d, *J* = 7.5 Hz, 1H), 4.74 (s, 2H), 3.89 (s, 6H), 3.75 (s, 3H), 3.01 (s, 3H), 2.22 (s, 3H), 2.03 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 166.29, 165.15, 162.49, 160.60, 157.32, 153.09, 146.70, 140.77, 139.19, 137.45, 136.72, 130.53, 130.44, 130.07, 129.68, 119.64, 118.64, 115.57, 108.77, 106.14, 105.74,

60.59, 56.56, 27.83, 17.91, 17.35. LC-MS (ESI,  $m/z$ ): 572.2550 [M + H]<sup>+</sup>.

**2-(3-Amino-4-methylphenylamino)-N-(5-(benzo[d][1,3]dioxole-5-carboxamido)-2-methylphenyl)-4-(methylamino)pyrimidine-5-carboxamide (26g).** (Method F) Yield 87%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.07 (s, 1H), 9.66 (s, 1H), 9.24 (s, 1H), 8.71 (s, 2H), 7.81 (s, 1H), 7.61 (d, *J* = 8.1 Hz, 1H), 7.56 (d, *J* = 8.4 Hz, 2H), 7.22 (d, *J* = 8.0 Hz, 1H), 7.16 (s, 1H), 7.07 (d, *J* = 8.0 Hz, 1H), 6.95 (d, *J* = 7.6 Hz, 1H), 6.83 (d, *J* = 7.8 Hz, 1H), 6.14 (s, 2H), 4.73 (s, 2H), 3.01 (s, 3H), 2.21 (s, 3H), 2.03 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 166.28, 164.81, 162.51, 160.62, 157.34, 150.48, 147.85, 146.73, 139.23, 137.65, 136.70, 130.49, 130.09, 129.51, 129.21, 123.28, 119.36, 118.45, 115.56, 108.78, 108.39, 108.16, 106.14, 102.27, 27.84, 17.93, 17.36. LC-MS (ESI,  $m/z$ ): 526.2130 [M + H]<sup>+</sup>.

**2-(3-Amino-4-methylphenylamino)-N-(5-(2,3-dihydrobenzo[b][1,4]dioxine-6-carboxamido)-2-methylphenyl)-4-(methylamino)pyrimidine-5-carboxamide (26h).** (Method F) Yield 72%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 8.72 (s, 1H), 7.84 (s, 1H), 7.55 (dd, *J* = 12.6, 3.9 Hz, 3H), 7.27–7.15 (m, 2H), 7.08–6.94 (m, 2H), 6.84 (d, *J* = 8.1 Hz, 1H), 4.31 (s, 4H), 3.01 (s, 3H), 2.21 (s, 3H), 2.06 (d, *J* = 17.7 Hz, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 166.17, 164.78, 162.42, 160.55, 157.34, 146.83, 146.64, 143.41, 139.13, 137.61, 136.57, 130.48, 130.10, 129.42, 128.17, 121.67, 119.19, 118.33, 117.31, 117.15, 115.56, 108.74, 106.01, 100.14, 64.87, 64.49, 27.69, 17.91, 17.32. LC-MS (ESI,  $m/z$ ): 540.2290 [M + H]<sup>+</sup>.

**Cell Lines and Cell Culture.** BaF3, P210-BaF3, Tel-ABL-BaF3, K562 (CML), Ku812 (CML), MEG-01 (CML), MV4-11 (AML), MOLM14 (AML), REC-1 (human B-cell lymphoma cell), OCI-AML-3 (AML), U937 (AML), Kasumi-1 (AML), HEL (AML), CHL (hamster lung cell), and CHO (hamster ovary cell) were used. All of the cells were grown in a humidified incubator at 37 °C under 5% CO<sub>2</sub>. CHO cells were maintained in DMEM supplemented with 10% FBS and 1% penicillin/streptomycin. BaF3, P210/BaF3, Tel-ABL-BaF3, K562, Ku812, MEG-01, MV4-11, MOLM14, REC-1, OCI-AML-3, U937, Kasumi-1, HEL, and CHL cells were grown in Roswell Park Memorial Institute (RPMI) 1640 medium supplemented with 10% FBS and 1% penicillin/streptomycin. Cells were grown in tissue culture flasks until they were 85–95% confluent prior to use. These nonadherent cells were collected by spin down at 700 rpm/min for 4 min.

**General Proliferation Protocol for Nonadherent Cells.** A density of 2 to 3 × 10<sup>4</sup> cells/mL cells were mixed with various concentrations of compounds, then 100 μL was added to each well and incubated for 72 h. Cell viability was determined using the CellTiter-Glo (Promega, USA) or CCK-8 (Bebuy, China). Both assays were performed according to the manufacturer's instructions. For the CellTiter-Glo assay, luminescence was determined in a multilabel reader (Envision, PerkinElmer, USA). For the CCK-8 assay, absorbance was measured in a microplate reader (iMARK, Bio-Rad, USA) at 450 and 650 nm. Data were normalized to the control group (DMSO). GI<sub>50</sub>s were calculated using Prism 5.0 (GraphPad Software, San Diego, CA).

**TEL-Isogenic Cell Generation.** Retroviral constructs for BaF3-FLT3 mutants were made based on the pMSCVpuro (Clontech) backbone. For TEL-FLT3 vector, the first 1 kb of human TEL gene with an artificial myristoylation sequence (MGCGCSSHPEDD) was cloned into the pMSCVpuro retroviral vector, followed by a 3xFLAG tag sequence and a stop codon. Then, the kinase domain coding the sequence of FLT3 was inserted in-frame between the TEL and 3xFLAG sequences. For full-length expression vectors, the coding sequences of FLT3 variants were directly cloned in a pMSCVpuro vector with a 3xFLAG tag at the C-terminal end. All mutagenesis studies were performed using QuikChange Site-Directed Mutagenesis Kit (Stratagene) following the manufacturer's instructions. The retrovirus was packaged in HEK293T cells by transfecting FLT3-containing MSCV vectors together with two helper plasmids. Virus supernatants were harvested 48 h after transfection and filtered before infection. Then, BaF3 cells were infected with harvested virus supernatants using the spinoculation protocol, and stable cell lines were obtained by puromycin selection for 48 h. The IL-3

concentrations in the culture medium were gradually withdrawn until cells were able to grow in the absence of IL-3.

**Signaling Pathway Study.** KU812, K562, and MEG-01 cells were treated with DMSO, serially diluted compound **18a**, 1  $\mu\text{M}$  Imatinib, and 0.1  $\mu\text{M}$  Dasatinib for 1 h. Cells were then washed in PBS and lysed in cell lysis buffer. Phospho-c-Abl (Tyr245)(73E5) rabbit mAb #2868, c-Abl antibody #2862, STAT5 (3H7) rabbit mAb #9358, phospho-STAT5 (Tyr694)(C71E5) rabbit mAb #9314, Akt (pan)-(C67E7) rabbit mAb #4691, phospho-Akt (Thr308) (244F9) rabbit mAb #4056, phospho-Akt (Ser473) (D9E) XP rabbit mAb #4060, phospho-Crkl (Tyr207) antibody #3181, Crkl (32H4) mouse mAb #3182, phospho-p44/42 MAPK (Erk1/2) (Thr202/Tyr204) (197G2) rabbit mAb #4377, and p44/42 MAPK (Erk1/2) (137F5) rabbit mAb #4695 antibody (Cell Signaling Technology) were used for immunoblotting.

**Apoptosis Effect Examination.** KU812, K562, and MEG-01 cells were treated with DMSO, serially diluted compound **18a**, 0.5  $\mu\text{M}$  Imatinib, and 0.5  $\mu\text{M}$  Dasatinib for the indicated periods. Cells were then washed in PBS and lysed in cell lysis buffer. PARP (46D11) rabbit mAb #9532, caspase-3 (8G10) rabbit mAb #9665, and GAPDH (14C10) rabbit mAb #2118 antibody (Cell Signaling Technology) were used for immunoblotting.

**Cell Cycle Analysis.** KU812, K562, and MEG-01 cells were treated with DMSO, serially diluted compound **18a**, 0.5  $\mu\text{M}$  Imatinib, and 0.5  $\mu\text{M}$  Dasatinib for the indicated periods. The cells were fixed in 70% cold ethanol and incubated at  $-20\text{ }^\circ\text{C}$  overnight then stained with PI/RNase staining buffer (BD Pharmingen). Flow cytometry was performed using FACS Calibur (BD), and results were analyzed by ModFit software.

**In Vivo Pharmacodynamics Studies.** Compound **18a** was dissolved in 55% saline containing 5% DMSO and 40% PEG400 by vortex. The final concentration of the stock solution was 1 mg/mL for administration. Six–eight week old male Sprague–Dawley rats were fasted overnight before starting drug treatment via intravenous and oral administration. Animal blood collection time points were as follows: for group 1, 3, and 5 (intravenous), 1 min, 5 min, 15 min, 30 min, 1 h, 2 h, 4 h, 6 h, and 8 h before and after administration were selected; for group 2, 4, and 6 (oral), 5 min, 15 min, 30 min, 1 h, 2 h, 4 h, 6 h, 8 h, and 24 h before and after dosing. Each time, about 0.2 mL of blood was collected through the jugular vein adding heparin for anticoagulation and kept on ice. Then, plasma was separated by centrifugation at 8000 rpm for 6 min at  $2\text{--}8\text{ }^\circ\text{C}$ . The obtained plasma was stored at  $-80\text{ }^\circ\text{C}$  before analysis. After finishing the test, all surviving animals will be transferred to the repository or euthanized ( $\text{CO}_2$  asphyxiation).

**K562 Xenograft Model.** Six week old female nu/nu mice were purchased from the Shanghai Experimental Center, Chinese Academy of Sciences (Shanghai, China). All animals were maintained in a specific pathogen-free facility and used according to the animal care regulations of Hefei Institutes of Physical Science, Chinese Academy of Sciences (Hefei, China), and all efforts were made to minimize animal suffering. To obtain orthotopic xenograft of human mammary tumor in the mice, cells were harvested during exponential growth. Six million K562 cells in PBS were suspended in a 1:1 mixture with Matrigel (BD Biosciences) and injected into the subcutaneous space on the right flank of nu/nu mice. Daily oral administration was initiated when K562 tumors had reached a size of 200 to 400  $\text{mm}^3$ . Animals were then randomized into treatment groups of 5 mice each for efficacy studies. Compound **18a** was delivered daily in a PEG300 solution (30% PEG300/10% ethanol in  $\text{ddH}_2\text{O}$ ) by oral gavage. A range of doses of **18a** or its vehicle was administered, as indicated in the Figure 8 legend. Body weight and tumor growth were measured daily after **18a** treatment. Tumor volumes were calculated as follows: tumor volume ( $\text{mm}^3$ ) =  $[(W^2 \times L)/2]$  in which width ( $W$ ) is defined as the smaller of the two measurements, and length ( $L$ ) is defined as the larger of the two measurements.

**HE Staining.** HE staining was carried out according to the previous report.<sup>21</sup> First, the sections were hydrated and then the slide was dipped into a Coplin jar containing Mayer's hematoxylin and agitated for 30 s. After rinsing the slide in  $\text{H}_2\text{O}$  for 1 min, it was stained with

1% eosin Y solution for 10–30 s with agitation. Subsequently, the sections were dehydrated with two changes of 95% alcohol and two changes of 100% alcohol for 30 s each, and then the alcohol was extracted with two changes of xylene. Finally, one or two drops of mounting medium was added and covered with a coverslip.

**K<sub>i</sub>-67 Staining.** For IHC demonstration of K<sub>i</sub>-67, tissue sections were quenched for endogenous peroxides and placed in an antigen retrieval solution (0.01 M citrate buffer, PH 6.0) for 15 min in a microwave oven at  $100\text{ }^\circ\text{C}$  at 600W. After incubation in the casein block, mouse mAb anti-K<sub>i</sub>-67 (ZSGB-BIO, China) was applied to the sections at dilutions of 1:50. Incubations with primary antibodies lasted overnight at  $4\text{ }^\circ\text{C}$ . The secondary detection system was used to visualize antibody binding. Staining was developed with DAB, and the slides were counterstained with hematoxylin, dehydrated, and mounted.

**TUNEL Staining.** TUNEL staining was performed using the POD in Situ Cell Death Detection kit (Roche, USA). Briefly, sections were deparaffinized in xylene, rehydrated in decreasing concentration of ethanol, and then treated by nuclease free Proteinase K for 15 min at room temperature before endogenous peroxidase was blocked in 3%  $\text{H}_2\text{O}_2$  in methanol. Terminal deoxynucleotidyl transferase (TdT) in reaction buffer was applied to sections for 1 h at  $37\text{ }^\circ\text{C}$ . Following washes, the slides were covered by converter-POD solution for 30 min at  $37\text{ }^\circ\text{C}$ . Apoptotic cells were detected after incubation in 3,3'-diaminobenzidine (DAB) chromogen (Beyotime Biotechnology, China) for approximately 8 min, and the slides were counterstained with hematoxylin.

**Molecular Modeling.** Molecular docking of small molecules to ABL1 kinase was performed with the software Autodock 4.0.<sup>22</sup> The ABL kinase structure (PDB ID: 2HYY) including the chain A of the kinase domain was used as the receptor, and polar hydrogen atoms were added to the receptor structure. All small molecules were constructed using the online-demo CORINA server. The grip map was adjusted as a dimension of  $64 \times 54 \times 50$  points with a spacing of 0.375 Å. The default parameters were used, and a total of 50 runs were performed with a Lamarckian genetic algorithm. The docked models were then clustered and sorted by binding energy.

## ■ ASSOCIATED CONTENT

### ● Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.jmedchem.5b01618.

DiscoverX's KinomeScan selectivity profiling data of compound **18a** and binding  $K_d$  of several clinical inhibitors determined by DiscoverX's binding assay, which was obtained from a previous publication (PDF) SMILES data (CSV)

## ■ AUTHOR INFORMATION

### Corresponding Authors

\*(Q.L.) E-mail: qslu97@hmfl.ac.cn.

\*(J.L.) E-mail: jingliu@hmfl.ac.cn.

### Author Contributions

<sup>○</sup>X.L., X.L., B.W., and F.Z. contributed equally to this work.

### Notes

The authors declare the following competing financial interest(s): Dr. Shanchun Zhang is a shareholder of Hefei Cosource Medicine Technology Co. LTD.

## ■ ACKNOWLEDGMENTS

We are grateful for the Scientific Research Grant of Hefei Science Center of CAS (SRG-HSC # 2015SRG-HSC022). J.L., Q.L., and W.W. are supported by the grant of "Cross-disciplinary Collaborative Teams Program for Science,

Technology and Innovation (2014-2016)" from Chinese Academy of Sciences. Z.Z. is supported by Anhui Province Natural Science Foundation Annual Key Program (grant number: 1301023011). We are also grateful for the China "Thousand Talent Program" support for Q.L. and the "Hundred Talent Program" of the Chinese Academy of Sciences support for J.L. and W.W. X. Liang was supported by the National Natural Science Foundation of China (grant number: 81402797). J.L. is also supported by the Top-Notch Young Talents Program of China.

## ■ ABBREVIATIONS USED

CML, chronic myeloid leukemia; AML, acute myeloid leukemia; MCL, mantle cell lymphoma; BCR, break point cluster region; ABL kinase, abelson kinase; HATU, 1-[bis(dimethylamino)methylene]-1H-1,2,3-triazolo[4,5-b]pyridinium 3-oxid hexafluorophosphate; DIPEA, N,N-diisopropylethylamine; DMF, dimethylformamide; DMAP, 4-dimethylaminopyridine; LAH, lithium aluminum hydride; TGI, tumor growth inhibition

## ■ REFERENCES

- (1) Sawyers, C. L.; Hochhaus, A.; Feldman, E.; Goldman, J. M.; Miller, C. B.; Ottmann, O. G.; Schiffer, C. A.; Talpaz, M.; Guilhot, F.; Deininger, M. W. N.; Fischer, T.; O'Brien, S. G.; Stone, R. M.; Gambacorti-Passerini, C. B.; Russell, N. H.; Reiffers, J. J.; Shea, T. C.; Chapuis, B.; Coutre, S.; Tura, S.; Morra, E.; Larson, R. A.; Saven, A.; Peschel, C.; Gratwohl, A.; Mandelli, F.; Ben-Am, M.; Gathmann, I.; Capdeville, R.; Paquette, R. L.; Druker, B. J. Imatinib induces hematologic and cytogenetic responses in patients with chronic myelogenous leukemia in myeloid blast crisis: results of a phase II study. *Blood* **2002**, *99* (10), 3530–3539.
- (2) Rana, A.; Hussain Shah, S.; Rehman, N.; Ali, S.; Muhammad Ali, G.; Bhatti, S.; Ahmad Farooqi, A. Chronic Myeloid Leukemia: Attributes Of Break Point Cluster Region-Abelson (BCR-ABL). *J. Cancer Res. Exp. Oncol.* **2011**, *3* (6), 5.
- (3) Greuber, E. K.; Smith-Pearson, P.; Wang, J.; Pendergast, A. M. Role of ABL family kinases in cancer: from leukaemia to solid tumours. *Nat. Rev. Cancer* **2013**, *13* (8), 559–571.
- (4) Deininger, M. W. N.; Goldman, J. M.; Melo, J. V. The molecular biology of chronic myeloid leukemia. *Blood* **2000**, *96* (10), 3343–3356.
- (5) Schindler, T.; Bornmann, W.; Pellicena, P.; Miller, W. T.; Clarkson, B.; Kuriyan, J. Structural mechanism for STI-571 inhibition of Abelson tyrosine kinase. *Science* **2000**, *289* (5486), 1938–1942.
- (6) Deininger, M.; Buchdunger, E.; Druker, B. J. The development of imatinib as a therapeutic agent for chronic myeloid leukemia. *Blood* **2005**, *105* (7), 2640–2653.
- (7) Weisberg, E.; Manley, P. W.; Breitenstein, W.; Bruggen, J.; Cowan-Jacob, S. W.; Ray, A.; Huntly, B.; Fabbro, D.; Fendrich, G.; Hall-Meyers, E.; Kung, A. L.; Mestan, J.; Daley, G. Q.; Callahan, L.; Catley, L.; Cavazza, C.; Mohammed, A.; Neuberg, D.; Wright, R. D.; Gilliland, D. G.; Griffin, J. D. Characterization of AMN107, a selective inhibitor of native and mutant Bcr-Abl. *Cancer Cell* **2005**, *7* (2), 129–141.
- (8) Shah, N. P.; Tran, C.; Lee, F. Y.; Chen, P.; Norris, D.; Sawyers, C. L. Overriding imatinib resistance with a novel ABL kinase inhibitor. *Science* **2004**, *305* (5682), 399–401.
- (9) Boschelli, D. H.; Ye, F.; Wang, Y. D.; Dutia, M.; Johnson, S. L.; Wu, B. Q.; Miller, K.; Powell, D. W.; Yaczko, D.; Young, M.; Tischler, M.; Arndt, K.; Discafani, C.; Etienne, C.; Gibbons, J.; Grod, J.; Lucas, J.; Weber, J. M.; Boschelli, F. Optimization of 4-phenylamino-3-quinolinecarbonitriles as potent inhibitors of Src kinase activity. *J. Med. Chem.* **2001**, *44* (23), 3965–3977.
- (10) O'Hare, T.; Shakespeare, W. C.; Zhu, X. T.; Eide, C. A.; Rivera, V. M.; Wang, F.; Adrian, L. T.; Zhou, T. J.; Huang, W. S.; Xu, Q. H.; Metcalf, C. A.; Tyner, J. W.; Loriaux, M. M.; Corbin, A. S.; Wardwell, S.; Ning, Y. Y.; Keats, J. A.; Wang, Y. H.; Sundaramoorthi, R.; Thomas, M.; Zhou, D.; Snodgrass, J.; Commodore, L.; Sawyer, T. K.; Dalgarno, D. C.; Deininger, M. W. N.; Druker, B. J.; Clackson, T. AP24534, a Pan-BCR-ABL Inhibitor for Chronic Myeloid Leukemia, Potently Inhibits the T315I Mutant and Overcomes Mutation-Based Resistance. *Cancer Cell* **2009**, *16* (5), 401–412.
- (11) Niwa, T.; Asaki, T.; Kimura, S. NS-187 (INNO-406), a Bcr-Abl/Lyn dual tyrosine kinase inhibitor. *Anal. Chem. Insights* **2007**, *2*, 93.
- (12) Carpinelli, P.; Ceruti, R.; Giorgini, M. L.; Cappella, P.; Gianellini, L.; Croci, V.; Degrassi, A.; Texido, G.; Rocchetti, M.; Vianello, P.; Rusconi, L.; Storici, P.; Zugnoni, P.; Arrigoni, C.; Soncini, C.; Alli, C.; Patton, V.; Marsiglio, A.; Ballinari, D.; Pesenti, E.; Fancelli, D.; Moll, J. PHA-739358, a potent inhibitor of Aurora kinases with a selective target inhibition profile relevant to cancer. *Mol. Cancer Ther.* **2007**, *6* (12), 3158–3168.
- (13) Chan, W. W.; Wise, S. C.; Kaufman, M. D.; Ahn, Y. M.; Ensinger, C. L.; Haack, T.; Hood, M. M.; Jones, J.; Lord, J. W.; Lu, W. P.; Miller, D.; Patt, W. C.; Smith, B. D.; Petillo, P. A.; Rutkoski, T. J.; Telikepalli, H.; Vogeti, L.; Yao, T.; Chun, L.; Clark, R.; Evangelista, P.; Gavrilescu, L. C.; Lazarides, K.; Zaleskas, V. M.; Stewart, L. J.; Van Etten, R. A.; Flynn, D. L. Conformational Control Inhibition of the BCR-ABL1 Tyrosine Kinase, Including the Gatekeeper T315I Mutant, by the Switch-Control Inhibitor DCC-2036. *Cancer Cell* **2011**, *19* (4), 556–568.
- (14) Ren, X. M.; Pan, X. F.; Zhang, Z.; Wang, D. P.; Lu, X. Y.; Li, Y. P.; Wen, D. H.; Long, H. Y.; Luo, J. F.; Feng, Y. B.; Zhuang, X. X.; Zhang, F. X.; Liu, J. Q.; Leng, F.; Lang, X. F.; Bai, Y.; She, M. Q.; Tu, Z. C.; Pan, J. X.; Ding, K. Identification of GZD824 as an Orally Bioavailable Inhibitor That Targets Phosphorylated and Non-phosphorylated Breakpoint Cluster Region-Abelson (Bcr-Abl) Kinase and Overcomes Clinically Acquired Mutation-Induced Resistance against Imatinib. *J. Med. Chem.* **2013**, *56* (3), 879–894.
- (15) Zhang, J. M.; Adrian, F. J.; Jahnke, W.; Cowan-Jacob, S. W.; Li, A. G.; Iacob, R. E.; Sim, T.; Powers, J.; Dierks, C.; Sun, F. X.; Guo, G. R.; Ding, Q.; Okram, B.; Choi, Y.; Wojciechowski, A.; Deng, X. M.; Liu, G. X.; Fendrich, G.; Strauss, A.; Vajpai, N.; Grzesiek, S.; Tuntland, T.; Liu, Y.; Bursulaya, B.; Azam, M.; Manley, P. W.; Engen, J. R.; Daley, G. Q.; Warmuth, M.; Gray, N. S. Targeting Bcr-Abl by combining allosteric with ATP-binding-site inhibitors. *Nature* **2010**, *463* (7280), 501–U116.
- (16) Nonami, A.; Sattler, M.; Weisberg, E.; Liu, Q. S.; Zhang, J. M.; Patricelli, M. P.; Christie, A. L.; Saur, A. M.; Kohl, N. E.; Kung, A. L.; Yoon, H.; Sim, T.; Gray, N. S.; Griffin, J. D. Identification of novel therapeutic targets in acute leukemias with NRAS mutations using a pharmacologic approach. *Blood* **2015**, *125* (20), 3133–3143.
- (17) Choi, H. G.; Ren, P. D.; Adrian, F.; Sun, F. X.; Lee, H. S.; Wang, X.; Ding, Q. A.; Zhang, G. B.; Xie, Y. P.; Zhang, J. M.; Liu, Y.; Tuntland, T.; Warmuth, M.; Manley, P. W.; Mestan, J.; Gray, N. S.; Sim, T. A Type-II Kinase Inhibitor Capable of Inhibiting the T315I "Gatekeeper" Mutant of Bcr-Abl. *J. Med. Chem.* **2010**, *53* (15), 5439–5448.
- (18) Martinelli, G.; Soverini, S.; Rosti, G.; Baccarani, M. Dual tyrosine kinase inhibitors in chronic myeloid leukemia. *Leukemia* **2005**, *19* (11), 1872–1879.
- (19) Dong, Y.; Xiong, M.; Duan, L.; Liu, Z.; Niu, T.; Luo, Y.; Wu, X.; Xu, C.; Lu, C. H2AX phosphorylation regulated by p38 is involved in Bim expression and apoptosis in chronic myelogenous leukemia cells induced by imatinib. *Apoptosis* **2014**, *19* (8), 1281–1292.
- (20) Winger, J. A.; Hantschel, O.; Superti-Furga, G.; Kuriyan, J. The structure of the leukemia drug imatinib bound to human quinone reductase 2 (NQO2). *BMC Struct. Biol.* **2009**, *9*, 7.
- (21) Fischer, A. H.; Jacobson, K. A.; Rose, J.; Zeller, R. Hematoxylin and eosin staining of tissue and cell sections. *CSH Protoc.* **2008**, DOI: 10.1101/pdb.prot4986.
- (22) Morris, G. M.; Huey, R.; Lindstrom, W.; Sanner, M. F.; Belew, R. K.; Goodsell, D. S.; Olson, A. J. AutoDock4 and AutoDockTools4: Automated docking with selective receptor flexibility. *J. Comput. Chem.* **2009**, *30* (16), 2785–2791.